

Package ‘cageminer’

January 21, 2022

Title Candidate Gene Miner

Version 1.1.0

Description This package aims to integrate GWAS-derived SNPs and coexpression networks to mine candidate genes associated with a particular phenotype. For that, users must define a set of guide genes, which are known genes involved in the studied phenotype. Additionally, the mined candidates can be given a score that favor candidates that are hubs and/or transcription factors. The scores can then be used to rank and select the top n most promising genes for downstream experiments.

License GPL-3

URL <https://github.com/almeidasilvaf/cageminer>

BugReports <https://support.bioconductor.org/t/cageminer>

biocViews Software, SNP, FunctionalPrediction, GenomeWideAssociation, GeneExpression, NetworkEnrichment, VariantAnnotation, FunctionalGenomics, Network

Encoding UTF-8

LazyData false

Roxygen list(markdown = TRUE)

RoxygenNote 7.1.1

Imports ggplot2, ggbio, ggtext, GenomeInfoDb, GenomicRanges, IRanges, reshape2, methods, BioNERO

Depends R (>= 4.1)

Suggests testthat (>= 3.0.0), SummarizedExperiment, knitr, BiocStyle, rmarkdown

Config/testthat/edition 3

VignetteBuilder knitr

git_url <https://git.bioconductor.org/packages/cageminer>

git_branch master

git_last_commit 55c80b9

git_last_commit_date 2021-10-26

Date/Publication 2022-01-21

Author Fabrício Almeida-Silva [aut, cre]

(<https://orcid.org/0000-0002-5314-2964>),

Thiago Venancio [aut] (<https://orcid.org/0000-0002-2215-8082>)

Maintainer Fabrício Almeida-Silva <fabricio_almeidasilva@hotmail.com>

R topics documented:

chr_length	2
gcn	3
gene_ranges	3
guides	4
hubs	4
mine2	5
mined_candidates	5
mine_candidates	6
mine_step1	7
mine_step2	8
mine_step3	9
pepper_se	10
plot_snp_circos	10
plot_snp_distribution	11
score_genes	12
simulate_windows	12
snp_pos	13
tfs	14

Index **15**

chr_length	<i>Pepper chromosome lengths</i>
------------	----------------------------------

Description

Lengths of pepper chromosomes 1-12 in a GRanges object. The genome for which lengths were calculated (v1.55) was downloaded from <http://peppergenome.snu.ac.kr/download.php>

Usage

```
data(chr_length)
```

Format

A GRanges object

Examples

```
data(chr_length)
```

gcn	<i>Simulation of the output list from BioNERO::exp2gcn() with pepper data</i>
-----	---

Description

This object is a list as returned by BioNERO::exp2gcn(), but only the element genes_and_modules is included. For running time issues, only genes in the cyan module were kept in the element genes_and_modules. All other list elements have been assigned NULL. The network was inferred using the code from the vignette.

Usage

```
data(gcn)
```

Format

A list with the elements returned by BioNERO::exp2gcn().

Examples

```
data(gcn)
```

gene_ranges	<i>Genomic coordinates of pepper genes</i>
-------------	--

Description

GRanges object with genomic coordinates of pepper genes downloaded from <http://peppergenome.snu.ac.kr/download.php>.

Usage

```
data(gene_ranges)
```

Format

A GRanges object

Examples

```
data(gene_ranges)
```

guides

Guide genes associated with defense and resistance to oomycetes

Description

The GO annotation was retrieved from PLAZA 4.0 Dicots.

Usage

```
data(guides)
```

Format

A data frame with genes in the first column and GO description in the second column.

References

Van Bel, M., Diels, T., Vancaester, E., Kreft, L., Botzki, A., Van de Peer, Y., ... & Vandepoele, K. (2018). PLAZA 4.0: an integrative resource for functional, evolutionary and comparative plant genomics. *Nucleic acids research*, 46(D1), D1190-D1196.

Examples

```
data(guides)
```

hubs

Example hub genes for the network stored in the gcn object

Description

The data frame was created using the code from the vignette.

Usage

```
data(hubs)
```

Format

Data frame with gene IDs, module and intramodular degree.

Examples

```
data(hubs)
```

`mine2`*Example output from mine_step2()*

Description

The list was created using the example code from `mine_step()`.

Usage

```
data(mine2)
```

Format

List with elements 'candidates' (character vector) and 'enrichment' (data frame).

Examples

```
data(mine2)
```

`mined_candidates`*Example output from mined_candidates()*

Description

The data frame was created using the code from the vignette.

Usage

```
data(mined_candidates)
```

Format

Data frame with an example of the output from `mined_candidates`

Examples

```
data(mined_candidates)
```

mine_candidates *Mine high-confidence candidate genes in a single step*

Description

Mine high-confidence candidate genes in a single step

Usage

```
mine_candidates(
  gene_ranges = NULL,
  marker_ranges = NULL,
  window = 2,
  expand_intervals = TRUE,
  gene_col = "ID",
  exp = NULL,
  gcn = NULL,
  guides = NULL,
  metadata,
  sample_group,
  min_cor = 0.2,
  alpha = 0.05,
  continuous = FALSE
)
```

Arguments

gene_ranges	A GRanges object with genomic coordinates of all genes in the genome.
marker_ranges	Genomic positions of SNPs. For a single trait, a GRanges object. For multiple traits, a GRangesList or CompressedGRangesList object, with each element of the list representing SNP positions for a particular trait.
window	Sliding window (in Mb) upstream and downstream relative to each SNP. Default: 2.
expand_intervals	Logical indicating whether or not to expand markers that are represented by intervals. This is particularly useful if users want to use a custom interval defined by linkage disequilibrium, for example. Default: TRUE.
gene_col	Column of the GRanges object containing gene ID. Default: "ID", the default for gff/gff3 files imported with rtracklayer::import.
exp	Expression data frame with genes in row names and samples in column names or a SummarizedExperiment object.
gcn	Gene coexpression network returned by BioNERO::exp2gcn().
guides	Guide genes as a character vector or as a data frame with genes in the first column and gene annotation class in the second column.

metadata	Sample metadata with samples in row names and sample information in the first column. Ignored if exp is a SummarizedExperiment object, as the colData will be extracted from the object.
sample_group	Level of sample metadata to be used for filtering in gene-trait correlation.
min_cor	Minimum correlation value for <code>BioNERO::gene_significance()</code> . Default: 0.2
alpha	Numeric indicating significance level. Default: 0.05
continuous	Logical indicating whether metadata is continuous or not. Default: FALSE

Value

A data frame with mined candidate genes and their correlation to the condition of interest.

Examples

```
data(pepper_se)
data(snp_pos)
data(gene_ranges)
data(guides)
data(gcn)
set.seed(1)
candidates <- mine_candidates(gene_ranges, snp_pos, exp = pepper_se,
                             gcn = gcn, guides = guides,
                             sample_group = "PRR_stress")
```

mine_step1

Step 1: Get all putative candidate genes for a given sliding window

Description

For a user-defined sliding window relative to each SNP, this function will subset all genes whose genomic positions overlap with the sliding window.

Usage

```
mine_step1(gene_ranges, marker_ranges, window = 2, expand_intervals = TRUE)
```

Arguments

gene_ranges	A GRanges object with genomic coordinates of all genes in the genome.
marker_ranges	Genomic positions of SNPs. For a single trait, a GRanges object. For multiple traits, a GRangesList or CompressedGRangesList object, with each element of the list representing SNP positions for a particular trait.
window	Sliding window (in Mb) upstream and downstream relative to each SNP. Default: 2.
expand_intervals	Logical indicating whether or not to expand markers that are represented by intervals. This is particularly useful if users want to use a custom interval defined by linkage disequilibrium, for example. Default: TRUE.

Value

A GRanges or GRangesList object with the genomic positions of all putative candidate genes.

See Also

[findOverlaps-methods](#)

Examples

```
data(snp_pos)
data(gene_ranges)
genes <- mine_step1(gene_ranges, snp_pos, window = 2)
```

mine_step2

Step 2: Get candidates in modules enriched in guide genes

Description

Step 2: Get candidates in modules enriched in guide genes

Usage

```
mine_step2(exp, gcn, guides, candidates)
```

Arguments

exp	Expression data frame with genes in row names and samples in column names or a SummarizedExperiment object.
gcn	Gene coexpression network returned by <code>BioNERO::exp2gcn()</code> .
guides	Guide genes as a character vector or as a data frame with genes in the first column and gene annotation class in the second column.
candidates	Character vector of all candidates genes to be inspected.

Value

A list of 2 elements:

candidates Character vector of candidates after step 2

enrichment Data frame of results for enrichment analysis

Examples

```
data(pepper_se)
data(guides)
data(gcn)
set.seed(1)
mine2 <- mine_step2(pepper_se, gcn = gcn, guides = guides$Gene,
                    candidates = rownames(pepper_se))
```

`mine_step3`*Step 3: Select candidates based on gene significance*

Description

Step 3: Select candidates based on gene significance

Usage

```
mine_step3(  
  exp,  
  metadata,  
  candidates,  
  sample_group,  
  min_cor = 0.2,  
  alpha = 0.05,  
  continuous = FALSE  
)
```

Arguments

<code>exp</code>	Expression data frame with genes in row names and samples in column names or a SummarizedExperiment object.
<code>metadata</code>	Sample metadata with samples in row names and sample information in the first column. Ignored if <code>exp</code> is a SummarizedExperiment object, as the <code>colData</code> will be extracted from the object.
<code>candidates</code>	Character vector of candidate genes to be inspected.
<code>sample_group</code>	Level of sample metadata to be used for filtering in gene-trait correlation.
<code>min_cor</code>	Minimum correlation value for <code>BioNERO::gene_significance()</code> . Default: 0.2
<code>alpha</code>	Numeric indicating significance level. Default: 0.05
<code>continuous</code>	Logical indicating whether metadata is continuous or not. Default: FALSE

Value

A data frame with mined candidate genes and their correlation to the condition of interest.

Examples

```
data(pepper_se)  
data(snp_pos)  
data(gene_ranges)  
data(guides)  
data(gcn)  
data(mine2)  
set.seed(1)  
mine3 <- mine_step3(pepper_se, candidates = mine2$candidates,  
  sample_group = "PRR_stress")
```

 pepper_se

Gene expression data from Kim et al., 2018.

Description

The data were filtered to keep only the top 4000 genes with highest RPKM values in PRR stress-related samples.

Usage

```
data(pepper_se)
```

Format

A SummarizedExperiment object.

References

Kim, MS., Kim, S., Jeon, J. et al. Global gene expression profiling for fruit organs and pathogen infections in the pepper, *Capsicum annuum* L.. *Sci Data* 5, 180103 (2018). <https://doi.org/10.1038/sdata.2018.103>

Examples

```
data(pepper_se)
```

 plot_snp_circos

Circos plot of SNP distribution across chromosomes

Description

Circos plot of SNP distribution across chromosomes

Usage

```
plot_snp_circos(genome_ranges, gene_ranges, marker_ranges)
```

Arguments

genome_ranges A GRanges object with chromosome lengths.

gene_ranges A GRanges object with genomic coordinates of all genes in the genome.

marker_ranges Genomic positions of SNPs. For a single trait, a GRanges object. For multiple traits, a GRangesList or CompressedGRangesList object, with each element of the list representing SNP positions for a particular trait.

Value

A ggplot object with a circos plot of molecular marker distribution across chromosomes.

Examples

```
data(snp_pos)
data(gene_ranges)
data(chr_length)
p <- plot_snp_circos(chr_length, gene_ranges, snp_pos)
```

`plot_snp_distribution` *Plot a barplot of SNP distribution across chromosomes*

Description

Plot a barplot of SNP distribution across chromosomes

Usage

```
plot_snp_distribution(marker_ranges)
```

Arguments

`marker_ranges` Genomic positions of SNPs. For a single trait, a GRanges object. For multiple traits, a GRangesList or CompressedGRangesList object, with each element of the list representing SNP positions for a particular trait. List elements must have names for proper labelling.

Value

A ggplot object.

Examples

```
data(snp_pos)
p <- plot_snp_distribution(snp_pos)
```

score_genes	<i>Score candidate genes and select the top n genes</i>
-------------	---

Description

Score candidate genes and select the top n genes

Usage

```
score_genes(mined_candidates, hubs = NULL, tfs = NULL, pick_top = 10)
```

Arguments

mined_candidates	Data frame resulting from mine_candidates() or mine_step().
hubs	Character vector of hub genes.
tfs	Character vector of transcription factors.
pick_top	Number of top genes to select. Default: 10.

Value

Data frame with top n candidates and their scores.

Examples

```
data(tfs)
data(hubs)
data(mined_candidates)
set.seed(1)
scored <- score_genes(mined_candidates, hubs$Gene, tfs$Gene_ID)
```

simulate_windows	<i>Simulate number of genes for each sliding window</i>
------------------	---

Description

This function counts genes that are contained in sliding windows related to each SNP.

Usage

```
simulate_windows(
  gene_ranges,
  marker_ranges,
  windows = seq(0.1, 2, by = 0.1),
  expand_intervals = TRUE
)
```

Arguments

gene_ranges	A GRanges object with genomic coordinates of all genes in the genome.
marker_ranges	Genomic positions of SNPs. For a single trait, a GRanges object. For multiple traits, a GRangesList or CompressedGRangesList object, with each element of the list representing SNP positions for a particular trait.
windows	Sliding windows (in Mb) upstream and downstream relative to each SNP. Default: seq(0.1, 2, by = 0.1).
expand_intervals	Logical indicating whether or not to expand markers that are represented by intervals. This is particularly useful if users want to use a custom interval defined by linkage disequilibrium, for example. Default: TRUE.

Details

By default, the function creates 20 sliding windows by expanding upstream and downstream boundaries for each SNP from 0.1 Mb (100 kb) to 2 Mb.

Value

A ggplot object summarizing the results of the simulations.

See Also

[findOverlaps-methods](#)

Examples

```
data(snp_pos)
data(gene_ranges)
simulate_windows(gene_ranges, snp_pos)
```

snp_pos	<i>Capsicum annuum</i> SNPs associated with resistance to <i>Phytophthora</i> root rot.
---------	---

Description

The SNPs in this data set were retrieved from Siddique et al., 2019, and they are associated to resistance to *Phytophthora* root rot.

Usage

```
data(snp_pos)
```

Format

A GRanges object.

References

Siddique, M.I., Lee, H.Y., Ro, N.Y. et al. Identifying candidate genes for *Phytophthora capsici* resistance in pepper (*Capsicum annuum*) via genotyping-by-sequencing-based QTL mapping and genome-wide association study. *Sci Rep* 9, 9962 (2019). <https://doi.org/10.1038/s41598-019-46342-1>

Examples

```
data(snp_pos)
```

tfs	<i>Pepper transcription factors</i>
-----	-------------------------------------

Description

Pepper transcription factors and their families retrieved from PlantTFDB 4.0.

Usage

```
data(tfs)
```

Format

A data frame with gene IDs in the first column and TF families in the second column.

References

Jin, J., Tian, F., Yang, D. C., Meng, Y. Q., Kong, L., Luo, J., & Gao, G. (2016). PlantTFDB 4.0: toward a central hub for transcription factors and regulatory interactions in plants. *Nucleic acids research*, gkw982.

Examples

```
data(tfs)
```

Index

* datasets

- chr_length, [2](#)
- gcn, [3](#)
- gene_ranges, [3](#)
- guides, [4](#)
- hubs, [4](#)
- mine2, [5](#)
- mined_candidates, [5](#)
- pepper_se, [10](#)
- snp_pos, [13](#)
- tfs, [14](#)

chr_length, [2](#)

gcn, [3](#)

gene_ranges, [3](#)

guides, [4](#)

hubs, [4](#)

mine2, [5](#)

mine_candidates, [6](#)

mine_step1, [7](#)

mine_step2, [8](#)

mine_step3, [9](#)

mined_candidates, [5](#)

pepper_se, [10](#)

plot_snp_circos, [10](#)

plot_snp_distribution, [11](#)

score_genes, [12](#)

simulate_windows, [12](#)

snp_pos, [13](#)

tfs, [14](#)