

# Package ‘TimeSeriesExperiment’

October 12, 2021

**Type** Package

**Title** Analysis for short time-series data

**Version** 1.11.1

**Date** 2021-08-31

**Description** TimeSeriesExperiment is a visualization and analysis toolbox for short time course data. The package includes dimensionality reduction, clustering, two-sample differential expression testing and gene ranking techniques. Additionally, it also provides methods for retrieving enriched pathways.

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**Depends** R (>= 4.1), S4Vectors (>= 0.19.23), SummarizedExperiment (>= 1.11.6)

**VignetteBuilder** knitr

**Imports** dynamicTreeCut, dplyr, edgeR, DESeq2, ggplot2 (>= 3.0.0), graphics, Hmisc, limma, methods, magrittr, proxy, stats, tibble, tidyr, vegan, viridis, utils

**Suggests** Biobase, BiocFileCache (>= 1.5.8), circlize, ComplexHeatmap, GO.db, grDevices, grid, knitr, org.Mm.eg.db, org.Hs.eg.db, MASS, RColorBrewer, rmarkdown, UpSetR,

**Collate** TimeSeriesExperiment-class.R TimeSeriesExperiment-methods.R  
utils.R pca.R clustering.R differential\_expression.R  
pathway\_over\_representation.R plots.R data.R

**Encoding** UTF-8

**RoxygenNote** 7.1.1

**biocViews** TimeCourse, Sequencing, RNASeq, Microbiome, GeneExpression, ImmunoOncology, Transcription, Normalization, DifferentialExpression, PrincipalComponent, Clustering, Visualization, Pathways

**BugReports** <https://github.com/nlhuong/TimeSeriesExperiment/issues>

**URL** <https://github.com/nlhuong/TimeSeriesExperiment>

**git\_url** <https://git.bioconductor.org/packages/TimeSeriesExperiment>

**git\_branch** master

**git\_last\_commit** fcb85c7

**git\_last\_commit\_date** 2021-08-31

**Date/Publication** 2021-10-12

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---

`.addLagsToTimeSeries` *Add differences to the time-course data.*

---

**Description**

This function add lag difference features to the time-course data.

**Usage**

```
.addLagsToTimeSeries(timeseries, lambda)
```

**Arguments**

<code>timeseries</code>	a data matrix or data.frame where each column corresponds to consecutive time point.
<code>lambda</code>	Weights for each lag difference, for time-course data. Length of lambda specifies number of lags to include. By default lag of order one and two are included with coefficients 0.5 and 0.25 respectively.

**Value**

a data matrix with added difference lags.

---

`.imputeData` *Imputing missing samples*

---

**Description**

Estimates assay values for missing timepoints

**Usage**

```
.imputeData(raw.data, sample.data)
```

**Arguments**

<code>raw.data</code>	Raw input data with columns corresponding to samples (observations) and rows to features
<code>sample.data</code>	Optional. A data.frame object were rows are samples (observations) and columns are sample attributes (e.g. group/condition, replicate, timepoint)

**Details**

Sometimes datasets include a missing sample. Here we impute the expression values by taking the mean expression of two samples from the same group and replicate (same individual) at two times surrounding the time of the missing sample

**Value**

list of modified raw.data and sample.data

---

addLags	<i>Add differences to the time-course data.</i>
---------	---

---

**Description**

This function concatenates lags to time-course data stored in elements of the `timeSeries` slot.

**Usage**

```
addLags(object, lambda = c(0.5, 0.25))
```

**Arguments**

object	A <code>TimeSeriesExperiment</code> object
lambda	Weights for each lag difference, for time-course data. Length of lambda specifies number of lags to include. By default lag of order one and two are included with coefficients 0.5 and 0.25 respectively.

**Value**

Returns `TimeSeriesExperiment` object with lags added to elements in `timeSeries` slot.

**Examples**

```
data("endoderm_small")
endoderm_small <- collapseReplicates(endoderm_small)
endoderm_small <- makeTimeSeries(endoderm_small)
endoderm_small <- addLags(endoderm_small)
head(timeSeries(endoderm_small, "ts"))
head(timeSeries(endoderm_small, "ts_collapsed"))
```

---

assayCollapsed	<i>Collapsed data.</i>
----------------	------------------------

---

### Description

Assay and colData collapsed over replicates. The values can be computed with set with [collapseReplicates](#) function.

### Usage

```
assayCollapsed(object, ...)  
  
## S4 method for signature 'TimeSeriesExperiment'  
assayCollapsed(object)  
  
assayCollapsed(object, ...) <- value  
  
## S4 replacement method for signature 'TimeSeriesExperiment'  
assayCollapsed(object) <- value  
  
colDataCollapsed(object, ...)  
  
## S4 method for signature 'TimeSeriesExperiment'  
colDataCollapsed(object)  
  
colDataCollapsed(object, ...) <- value  
  
## S4 replacement method for signature 'TimeSeriesExperiment'  
colDataCollapsed(object) <- value
```

### Arguments

object	a TimeSeriesExperiment object.
...	arguments to other functions.
value	a numerical matrix

### Value

a [DataFrame](#)

### Examples

```
data("endoderm_small")  
endoderm_small <- collapseReplicates(endoderm_small)  
head(assayCollapsed(endoderm_small))  
  
data("endoderm_small")
```

```
endoderm_small <- collapseReplicates(endoderm_small)
head(colDataCollapsed(endoderm_small))
```

---

assignClusterDynamic *Cluster assignment using dynamic branch cutting.*

---

### Description

This function computes the cluster assignment for hierarchical clustering results using dynamic branch cutting.

### Usage

```
assignClusterDynamic(hclst, max_height = 0.9, ...)
```

### Arguments

hclst	an object of class hclust representing the clustering of features of X.
max_height	a fraction of the total tree height used to compute maxTreeHeight argument for <a href="#">cutreeDynamic</a> function from dynamicTreeCut package.
...	other parameters for <a href="#">cutreeDynamic</a> .

### Value

a data.frame containing cluster assignments.

---

assignClusterStatic *Cluster assignment using static branch cutting.*

---

### Description

This function computes the cluster assignment for hierarchical clustering results using static branch cutting.

### Usage

```
assignClusterStatic(hclst, h = NULL, k = NULL)
```

### Arguments

hclst	an object of class hclust representing the clustering of features of X.
h	a the fraction of the max tree height at which to cut and assign labels.
k	an integer scalar or vector with the desired number of groups

### Value

a data.frame containing cluster assignments.

---

clusterAssignment      *Cluster analysis results*

---

## Description

Getter methods for `clusterAssignment` slot of a `TimeSeriesExperiment` object. The slot is a list with with elements named: `'settings'`, `'hclust'`, `'cluster_map'`, `'clust_centroids'` storing results from running `clusterTimeSeries` function.

## Usage

```
clusterAssignment(object, ...)  
  
## S4 method for signature 'TimeSeriesExperiment'  
clusterAssignment(object, name = NULL)  
  
clusterMap(object, ...)  
  
## S4 method for signature 'TimeSeriesExperiment'  
clusterMap(object)
```

## Arguments

<code>object</code>	a <code>TimeSeriesExperiment</code> object.
<code>...</code>	arguments to other functions.
<code>name</code>	one of elements of <code>'clusterAssignment'</code> slot: <code>'settings'</code> , <code>'hclust'</code> , <code>'cluster_map'</code> , <code>'clust_centroids'</code> . If <code>NULL</code> , all elements are returned.

## Value

a `data.frame`

## Examples

```
data("endoderm_small")  
endoderm_small <- clusterTimeSeries(endoderm_small)  
clusterAssignment(endoderm_small, name = 'settings')  
head(clusterAssignment(endoderm_small, name = 'final_cluster_map'))  
head(clusterAssignment(endoderm_small, name = 'clust_centroids'))
```

---

`clusterData`*Cluster time series data.*

---

### Description

Perform cluster assignment using hierarchical clustering and static or dynamic branch cutting. If 'subset' is specified the clustering is performed on the subset of the data, and the rest of the rows are assigned based on the distances to the centroids of computed clusters.

### Usage

```
clusterData(  
  X,  
  dist = "euclidean",  
  dynamic = FALSE,  
  hclust_params = list(),  
  static_cut_params = list(h = 0.5),  
  dynamic_cut_params = list(max_height = 0.9)  
)
```

### Arguments

`X` a data matrix or data.frame where rows are time series.

`dist` the distance metric for the dissimilarity used for clustering.

`dynamic` whether dynamic branch cutting should be done for cluster assignment.

`hclust_params` parameters for [hclust](#) function.

`static_cut_params` parameters for [assignClusterStatic](#).

`dynamic_cut_params` parameters for [assignClusterDynamic](#).

### Value

a list with the hclust object, as well as `clust_map` and `clust_centroids` data.frames.

### Examples

```
data("endoderm_small")  
endoderm_small <- normalizeData(endoderm_small)  
X <- SummarizedExperiment::assays(endoderm_small)$norm  
clust_res <- clusterData(X)  
head(clust_res$clust_centroids)  
head(clust_res$clust_map)
```



---

clusterTimeSeries      *Cluster time series features.*

---

### Description

Find the cluster assignment for timecourse features. Clustering computed for top "n.top.feats" features most variable over time in each of the selected "groups" using time-series expression (collapsed over replicates). The cluster assignment of the remaining genes is based on the distance to the closest cluster centroid previously obtained. Hierarchical clustering is performed and both static and dynamic branch cutting algorithm are available for assigning cluster membership.

### Usage

```
clusterTimeSeries(  
  object,  
  n.top.feats = 1000,  
  groups.selected = "all",  
  lambda = c(0.5, 0.25),  
  clust.params = list()  
)
```

### Arguments

object	A TimeSeriesExperiment object
n.top.feats	A number of top most variable time-course features to use for clustering.
groups.selected	One or multiple groups from object@group to take into account when aggregating time-course features.
lambda	Weights for each lag difference, for time-course data. Length of lambda specifies number of lags to include. By default lag of order one and two are included with coefficients 0.5 and 0.25 respectively.
clust.params	A list containing arguments for hierarchical clustering. For details see <a href="#">clusterData</a> .

### Value

a TimeSeriesExperiment object with cluster assignment stored in cluster.map slot.

### Examples

```
data("endoderm_small")  
endoderm_small <- clusterTimeSeries(endoderm_small)  
head(clusterMap(endoderm_small))
```

---

```
colData<-, TimeSeriesExperiment, ANY-method  
      Column data for TimeSeriesExperiment
```

---

## Description

colData() slots holds information on individual samples including corresponding sample name in column 'sample' as well as time, group and replicate.

## Usage

```
## S4 replacement method for signature 'TimeSeriesExperiment, ANY'  
colData(x, ...) <- value
```

## Arguments

x	a TimeSeriesExperiment object
...	arguments to other functions.
value	a <a href="#">DataFrame</a> with new sample information

## Details

The setter also updates the information in timepoint, replicate and group slots and resets the time-series analysis results to NULL.

## Value

a [S4Vectors::DataFrame](#)

## Examples

```
data("endoderm_small")  
head(colData(endoderm_small))  
newdf <- colData(endoderm_small)  
newdf$random <- sample(ncol(endoderm_small), ncol(endoderm_small))  
colData(endoderm_small) <- newdf  
head(colData(endoderm_small))
```

---

collapseReplicates      *Collapse data over replicates.*

---

### Description

This function aggregates the data over replicates, i.e. returns collapse data for each group and at each time point.

### Usage

```
collapseReplicates(object, FUN = mean)
```

### Arguments

object	A TimeSeriesExperiment object
FUN	the aggregate function. Default is mean

### Value

Returns TimeSeriesExperiment object after collapsing over replicates. Collapsed data is stored sample.data.collapsed and data.collapsed slots.

### Examples

```
data("endoderm_small")
endoderm_small <- collapseReplicates(endoderm_small)
assayCollapsed(endoderm_small)[1:10, 1:6]
```

---

colnames<- , TimeSeriesExperiment-method

*Row and column name getters and setters for TimeSeriesExperiment object.*

---

### Description

colnames(), rownames() can be used to reset the dimensions, column and row names respectively. The data will be updated across all slots of TimeSeriesExperiment.

### Usage

```
## S4 replacement method for signature 'TimeSeriesExperiment'
colnames(x) <- value
```

```
## S4 replacement method for signature 'TimeSeriesExperiment'
rownames(x) <- value
```

**Arguments**

`x` a TimeSeriesExperiment object.  
`value` a character vector or a list of two character vectors with new dimension names.

**Details**

Setting `colnames()` automatically updates information in `dimensionReduction` slot.

**Value**

a character vector

**Examples**

```
data("endoderm_small")
head(colnames(endoderm_small))
colnames(endoderm_small) <- paste0("Smp", 1:ncol(endoderm_small))
head(colnames(endoderm_small))
```

---

dataToTimeSeries      *Data to time-series*

---

**Description**

Function that splits data to time series for each replicate included.

**Usage**

```
dataToTimeSeries(X, timepoint, group = NULL, replicate = NULL)
```

**Arguments**

`X` a data matrix or data.frame where columns correspond to samples and rows to features.  
`timepoint` a vector of length equal to `ncol(X)` indicating the time variable corresponding to the sample (data column).  
`group` a vector of length equal to `ncol(X)` indicating the group membership corresponding to the sample (data column).  
`replicate` a vector of length equal to `ncol(X)` indicating the replicate variable corresponding to the sample (data column).

**Value**

a data.frame with ordered time series for each replicate.

## Examples

```
X <- matrix(rnorm(1000), ncol = 50)
group <- rep(c("A", "B"), each = 25)
replicate <- rep(paste0("rep", 1:5), each = 5)
time <- rep(1:5, 10)
tc <- dataToTimeSeries(X, time, replicate, group)
head(tc)
```

---

differentialExpression

*Differential Expression for TimeSeriesExperiment*

---

## Description

Getter method for differentialExpression slot of a TimeSeriesExperiment. The slot is a list with differential expression results possibly containing elements named 'timepoint\_de', and 'trajectory\_de' computed with [timepointDE](#) and [trajectoryDE](#) functions.

## Usage

```
differentialExpression(object, ...)
```

```
## S4 method for signature 'TimeSeriesExperiment'
differentialExpression(object, name = NULL)
```

## Arguments

object	a TimeSeriesExperiment object.
...	arguments to other functions.
name	one of elements of 'differentialExpression' slot: 'timepoint_de', 'trajectory_de'. If NULL, all elements are returned.

## Value

a data.frame or a list of data.frames

## Examples

```
data("endoderm_small")
endoderm_small <- trajectoryDE(endoderm_small)
head(differentialExpression(endoderm_small, "trajectory_de"))
```

---

dimensionReduction      *Dimension reduction results*

---

### Description

Getter methods for dimensionReduction slot of a TimeSeriesExperiment object. The slot is a list of data.frames: 'pca\_sample', 'pca\_feature' and 'pca\_eigs' storing results from a PCA projection.

### Usage

```
dimensionReduction(object, ...)

## S4 method for signature 'TimeSeriesExperiment'
dimensionReduction(object, name = NULL)
```

### Arguments

object	a TimeSeriesExperiment object.
...	arguments to other functions.
name	one of elements of 'dimensionReduction' slot: 'pca_sample', 'pca_feature' and 'pca_eigs' for returning the entire list. If NULL, all elements are returned.

### Value

a data.frame or a list of data.frames

### Examples

```
data("endoderm_small")
endoderm_small <- runPCA(endoderm_small)
head(dimensionReduction(endoderm_small, "pca_sample")[, 1:3])
```

---

endoderm\_small      *Endoderm differentiation*

---

### Description

A 'TimeSeriesExperiment' object containing gene expression data, feature and sample data from a study by Blake et al. "A Comparative Study Of Endoderm Differentiation In Humans And Chimpanzees". Here we include only a small fraction of the data including top 250 most variable genes, to serve as a toy example for TimeSeriesExperiment.

### Usage

```
data(endoderm_small)
```

**Format**

A TimeSeriesExperiment object

**Source**

<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE98411>

**References**

Blake et al. (2017) ([bioRxiv](#))

---

filterFeatures	<i>Filter features</i>
----------------	------------------------

---

**Description**

Filters TimeSeriesExperiment object to keep only chosen features. All relevant slots are updates.

**Usage**

```
filterFeatures(object, features)
```

**Arguments**

object	TimeSeriesExperiment object
features	features (genes) to keep

**Details**

The slots for collapsed data and time series formatted data are filtered accordingly, but `dimensionReduction`, `clusterAssignment` and `differentialExpression` are reset to NULL as different set of features would output in different results.

**Value**

"TimeSeriesExperiment" object

**Examples**

```
data("endoderm_small")
features <- 1:100
endoderm_small <- filterFeatures(endoderm_small, features)
endoderm_small
```

---

groups	<i>Group information</i>
--------	--------------------------

---

## Description

Group information

## Usage

```
groups(object, ...)  
  
## S4 method for signature 'TimeSeriesExperiment'  
groups(object)  
  
groups(object, ...) <- value  
  
## S4 replacement method for signature 'TimeSeriesExperiment'  
groups(object) <- value
```

## Arguments

object	a TimeSeriesExperiment object.
...	arguments to other functions.
value	a character vector with new group membership

## Value

a character vector

## Examples

```
data("endoderm_small")  
head(groups(endoderm_small))  
groups(endoderm_small) <- sample(c("A", "B"),  
  ncol(endoderm_small), replace = TRUE)  
head(groups(endoderm_small))
```



---

makeTimeSeries	<i>Convert data to time-series</i>
----------------	------------------------------------

---

## Description

This function converts the wide data matrix to time-course long data.frame format where each row gives data values over time (at each time point) for each feature, group, and replicate.

## Usage

```
makeTimeSeries(  
  object,  
  feature.trans.method = "var_stab",  
  var.stabilize.method = "asinh"  
)
```

## Arguments

`object` A TimeSeriesExperiment object

`feature.trans.method` Method for feature normalization. Default "none". Currently supports only "none" (no transformation), "scale\_feat\_sum" (scaling by feature sum), or "var\_stab" (variance stabilization). Default is "var\_stab".

`var.stabilize.method` Method for variance stabilization (VST). Currently, supports "none" (no VST), "log1p" (log plus one), "asinh" (inverse hyperbolic sine) or "deseq" ([varianceStabilizingTransform](#) function from DESeq2 package). Default is "log1p".

## Value

Returns TimeSeriesExperiment object after conversion to time-course format. Converted data is stored in timecourse.data slot.

## Examples

```
data("endoderm_small")  
endoderm_small <- makeTimeSeries(endoderm_small)  
names(timeSeries(endoderm_small))  
head(timeSeries(endoderm_small)[[1]])
```

---

`makeTimeSeriesExperimentFromExpressionSet`*TimeSeriesExperiment constructor from ExpressionSet*

---

## Description

`TimeSeriesExperiment` constructor initializes the `TimeSeriesExperiment` object from `ExpressionSet` and populates the time, replicate, and group slots.

## Usage

```
makeTimeSeriesExperimentFromExpressionSet(  
  eset,  
  timepoint = NULL,  
  group = NULL,  
  replicate = NULL  
)
```

## Arguments

<code>eset</code>	ExpressionSet object
<code>timepoint</code>	a vector indicating timepoint at which each sample was collected or a character string equal to one of the column names of a supplied ExpressionSet.
<code>group</code>	a vector indicating a group membership for each sample or a character string equal to one of the column names of a supplied ExpressionSet. If not specified, the group is set to 'G1' for each sample.
<code>replicate</code>	a vector indicating a replicate id of each sample or a character string equal to one of the column names of a supplied ExpressionSet. If not specified, the replicate is set to 'R1' for each sample.

## Details

`TimeSeriesExperiment` is an extension of `SummarizedExperiment` class.

## Value

Returns an initialized `TimeSeriesExperiment` object.

## Examples

```
raw <- matrix(runif(3000), ncol = 30)  
pheno.data <- data.frame(  
  time = rep(rep(1:5, each = 3), 2),  
  replicate = rep(1:3, 10),  
  group = rep(1:2, each = 15))  
feature.data <- data.frame(  
  feature = paste0("F", 1:100)
```

```

)
## Not run:
library(Biobase)
test_eset <- ExpressionSet(
  raw, phenoData = AnnotatedDataFrame(pheno.data),
  featureData = AnnotatedDataFrame(feature.data))
test_TimeSeriesExperiment <- makeTimeSeriesExperimentFromExpressionSet(
  test_eset, timepoint = "time", group = "group",
  replicate = "replicate")
test_TimeSeriesExperiment

## End(Not run)

```

---

```
makeTimeSeriesExperimentFromSummarizedExperiment
```

*TimeSeriesExperiment constructor from SummarizedExperiment*

---

## Description

TimeSeriesExperiment constructor initializes the object from [SummarizedExperiment::SummarizedExperiment](#) and populates the time, replicate, and group slots.

## Usage

```

makeTimeSeriesExperimentFromSummarizedExperiment(
  se,
  timepoint = NULL,
  group = NULL,
  replicate = NULL
)

```

## Arguments

se	<a href="#">SummarizedExperiment::SummarizedExperiment</a> object
timepoint	a vector indicating timepoint at which each sample was collected or a character string equal to one of the column names of a supplied SummarizedExperiment.
group	a vector indicating a group membership for each sample or a character string equal to one of the column names of a supplied SummarizedExperiment If not specified, the group is set to 'G1' for each sample.
replicate	a vector indicating a replicate id of each sample or a character string equal to one of the column names of a supplied SummarizedExperiment If not specified, the replicate is set to 'R1' for each sample.

## Details

TimeSeriesExperiment is an extension of SummarizedExperiment class.

**Value**

Returns an initialized TimeSeriesExperiment object.

**Examples**

```
raw <- matrix(runif(3000), ncol = 30)
pheno.data <- data.frame(
  time = rep(rep(1:5, each = 3), 2),
  replicate = rep(1:3, 10),
  group = rep(1:2, each = 15))
feature.data <- data.frame(
  feature = paste0("F", 1:100)
)
test_sumexp <- SummarizedExperiment::SummarizedExperiment(
  assays = list(counts = raw),
  rowData = feature.data, colData = pheno.data)
test_TimeSeriesExperiment <-
makeTimeSeriesExperimentFromSummarizedExperiment(
  test_sumexp, timepoint = "time", group = "group",
  replicate = "replicate")
test_TimeSeriesExperiment
```

---

meltMatrix

*Melt data matrix.*

---

**Description**

Convert data matrix to a long format

**Usage**

```
meltMatrix(X)
```

**Arguments**

X                    a data matrix.

**Value**

a data matrix in a long format.

**Examples**

```
Z <- matrix(rnorm(100), 20)
Z.m <- meltMatrix(Z)
head(Z.m)
```

---

normalizeData	<i>Normalize Data</i>
---------------	-----------------------

---

**Description**

Normalize data the assay data.

**Usage**

```
normalizeData(  
  object,  
  sample.norm.method = "scale_common_factor",  
  column.scale.factor = 1e+06  
)
```

**Arguments**

`object` A TimeSeriesExperiment object or a data matrix/data frame.

`sample.norm.method` Method for sample normalization. Currently supports only scaling to a common factor, "scale\_common\_factor" which with `column.scale.factor = 1e+06` is equivalent to CPM normalization.

`column.scale.factor` Sets the scale factor for sample-level normalization

**Value**

Returns TimeSeriesExperiment object after normalization. Normalized data is stored data slot.

**Examples**

```
data("endoderm_small")  
names(assays(endoderm_small))  
endoderm_small <- normalizeData(endoderm_small)  
assays(endoderm_small)$norm[1:10, 1:6]
```

---

pathwayEnrichment	<i>Pathway enrichment testing.</i>
-------------------	------------------------------------

---

**Description**

This is a wrapper around [goana](#) function from limma package for testing over-representation of gene ontology (GO) terms or KEGG pathways in sets of genes.

**Usage**

```

pathwayEnrichment(
  object,
  features,
  species,
  feature_column = "feature",
  universe = NULL,
  clustered = TRUE,
  kegg = FALSE,
  ontology = c("BP", "CC", "MF"),
  fltr_DE = 0.1,
  fltr_N = 500,
  fltr_P.DE = 0.05,
  ...
)

```

**Arguments**

<code>object</code>	A TimeSeriesExperiment object.
<code>features</code>	A vector of ENTREZID for enrichment testing.
<code>species</code>	A character string specifying the species. See <a href="#">goana</a> for details.
<code>feature_column</code>	the feature column in 'feature.data' slot holding ENTREZIDs, by default the 'feature' column.
<code>universe</code>	A vector of genes in the universe. By default all the genes in the 'raw.data' slot.
<code>clustered</code>	Whether features should be grouped based on cluster assignment (stored in <code>object@cluster.features\$cluster_map</code> ) and tested as separate sets.
<code>kegg</code>	Whether KEGG pathways should be used instead of gene ontology (GO)
<code>ontology</code>	character vector of ontologies to be included in output. Elements should be one or more of "BP", "CC" or "MF".
<code>fltr_DE</code>	A scalar fraction of the number of genes in tested set to use as a threshold for filtering genes based on "DE" column (the number of genes in the DE set). Default is 0.1, i.e. at least 0.1 genes in the set must be present in the pathway.
<code>fltr_N</code>	A number of genes used as a threshold to filter out all pathway terms of size greater than the threshold. Default is 500.
<code>fltr_P.DE</code>	A p-value threshold to filter out terms in the enrichment results. Default is 0.05.
<code>...</code>	other parameters for <a href="#">goana</a> or <code>limma::kegga()</code> .

**Value**

a data.frame or list of data.frames with enrichment results.

**Examples**

```

data("endoderm_small")
selected_genes <- c('114299', '2825', '3855', '221400', '7941',

```

```
      '6164', '1292', '6161', '6144', '23521')
enrich_res <- pathwayEnrichment(
  object = endoderm_small, clustered = FALSE,
  features = selected_genes,
  species = "Hs", ontology = "BP", fltr_DE = 0,
  fltr_N = Inf, fltr_P.DE = 0.05)
head(enrich_res)
```

---

plotEnrichment	<i>Plot enrichment results.</i>
----------------	---------------------------------

---

### Description

Plotting top most enriched terms found with DE methods and tested for overrepresentation in GO/KEGG db using goana/kegga from limma package.

### Usage

```
plotEnrichment(enrich, n_max = 15)
```

### Arguments

enrich	a data matrix or data.frame with enrichment result - outputs from <a href="#">pathwayEnrichment</a> function or <a href="#">goana</a> , <a href="#">limma::kegga()</a> . Must contain columns Term, DE, and P.DE.
n_max	max number of terms to show

### Value

a ggplot object

### Examples

```
data("endoderm_small")
selected_genes <- c('114299', '2825', '3855', '221400', '7941',
  '6164', '1292', '6161', '6144', '23521')
enrich_res <- pathwayEnrichment(
  object = endoderm_small, clustered = FALSE,
  features = selected_genes,
  species = "Hs", ontology = "BP", fltr_DE = 0,
  fltr_N = Inf, fltr_P.DE = 0.05)
plotEnrichment(enrich = enrich_res, n_max = 15)
```

---

plotHeatmap	<i>Plot a heatmap</i>
-------------	-----------------------

---

### Description

Generates a heatmap of expression data.

### Usage

```
plotHeatmap(  
  object,  
  num.feats = 200,  
  scale = TRUE,  
  feat_desc = "feature",  
  sample_desc = "sample"  
)
```

### Arguments

object	A TimeSeriesExperiment object
num.feats	Number of top most features to use.
scale	Whether to scale the data (by features) before plotting.
feat_desc	One of the column names from feature_data(object) to describe the features.
sample_desc	One of the column names from sample_data(object) to describe the samples.

### Value

Returns a ggplot2 object.

### Examples

```
data("endoderm_small")  
endoderm_small <- normalizeData(endoderm_small)  
## Not run:  
  plotHeatmap(endoderm_small)  
  
## End(Not run)
```



---

plotSamplePCA	<i>Plot a standard PCA</i>
---------------	----------------------------

---

**Description**

Generates a standard PCA plot of observations in the dataset.

**Usage**

```
plotSamplePCA(object, axis = c(1, 2), col.var = NULL, ...)
```

**Arguments**

object	A TimeSeriesExperiment object
axis	An integer vector indicating principal components to use for plotting, by default 1:2.
col.var	A character string indicating a column from sample_data(object) which should be used for coloring the points. By default NULL.
...	other parameters ggplot.

**Value**

Returns a ggplot2 objet.

**Examples**

```
data("endoderm_small")
endoderm_small <- runPCA(endoderm_small)
plotSamplePCA(endoderm_small, col.var = "group")
```

---

plotTimeSeries	<i>Plot selected time series.</i>
----------------	-----------------------------------

---

**Description**

Plotting expression over time for selected genes curve and colors correspond to distinct groups.

**Usage**

```
plotTimeSeries(
  object,
  features = rownames(object),
  trans = FALSE,
  smooth = TRUE,
  ncol = 5,
  scales = "free"
)
```

**Arguments**

object	A TimeSeriesExperiment object.
features	A vector of names of selected features to plot.
trans	A boolean indicating whether (TRUE) transformed, variance stabilized, assay values should be printed or (FALSE) just normalized by sample depth.
smooth	If TRUE a smoothed line is plotted for each gene and each group, else a piecewise linear average (over replicates) curve is plotted.
ncol	An integer indicating the number of columns for faceting. Default is 5.
scales	character scalar indecating facet scales, by default "free".

**Value**

list of ggplot objects

**Examples**

```
data("endoderm_small")
feat_to_plot <- rownames(endoderm_small)[1:10]
plotTimeSeries(endoderm_small, features = feat_to_plot, smooth = FALSE)
```

---

plotTimeSeriesClusters

*Plot (time) series over clusters.*

---

**Description**

Plots timecourse (aggregated over replicates) feature data faceted by computed clusters and experimental group.

**Usage**

```
plotTimeSeriesClusters(
  object,
  features = NULL,
  transparency = 0.5,
  ncol = 4,
  scales = "free"
)
```

**Arguments**

object	A TimeSeriesExperiment object.
features	A vector of names of selected features to plot.
transparency	transparency of trajectory lines.
ncol	number of columns in the facet plot.
scales	character scalar indecating facet scales, by default "free".

**Value**

ggplot object

**Examples**

```
data("endoderm_small")
endoderm_small <- clusterTimeSeries(endoderm_small)
plotTimeSeriesClusters(endoderm_small)
```

---

plotTimeSeriesPCA      *Overlay (time) series over PCA grid*

---

**Description**

PCA plot for data features, with time-series levels overlaid on top.

**Usage**

```
plotTimeSeriesPCA(
  object,
  axis = c(1, 2),
  m = 20,
  n = 20,
  group.highlight = NULL,
  linecol = NULL,
  ...
)
```

**Arguments**

object	A TimeSeriesExperiment object.
axis	An integer vector indicating principal components to use for plotting, by default 1:2.
m	a number of tiles in a grid in the horizontal direction.
n	a number of tiles in a grid in the vertical direction.
group.highlight	An optional character string indicating the group subset for which the time-course trends should be plotted. By default all time-course trends are plotted for all groups.
linecol	a vector indicating the color of the gene profile trend line, different for each group.
...	other parameters for the line plots.

**Value**

None

**Examples**

```
data("endoderm_small")
endoderm_small <- runPCA(endoderm_small)
plotTimeSeriesPCA(endoderm_small)
```

---

replicates	<i>Replicate information</i>
------------	------------------------------

---

**Description**

Replicate information

**Usage**

```
replicates(object, ...)
```

## S4 method for signature 'TimeSeriesExperiment'

```
replicates(object)
```

replicates(object, ...) <- value

## S4 replacement method for signature 'TimeSeriesExperiment'

```
replicates(object) <- value
```

**Arguments**

object	a TimeSeriesExperiment object.
...	arguments to other functions.
value	a character vector with new replicate ids.

**Value**

a character vector

**Examples**

```
data("endoderm_small")
head(replicates(endoderm_small))
replicates(endoderm_small) <- sample(c("R1", "R2", "R3"),
  ncol(endoderm_small), replace = TRUE)
head(replicates(endoderm_small))
```

---

```
rowData<-, TimeSeriesExperiment-method
      Row data for TimeSeriesExperiment
```

---

**Description**

rowData() holds information on individual features including corresponding feature name in column 'feature'.

**Usage**

```
## S4 replacement method for signature 'TimeSeriesExperiment'
rowData(x, ...) <- value
```

**Arguments**

x	a TimeSeriesExperiment object.
...	arguments to other functions.
value	a DataFrame with new feature data

**Value**

a [S4Vectors::DataFrame](#)

**Examples**

```
data("endoderm_small")
head(rowData(endoderm_small))
rowData(endoderm_small) <- data.frame(
  feature = rowData(endoderm_small)$feature,
  random = sample(nrow(endoderm_small), nrow(endoderm_small)))
head(rowData(endoderm_small))
```

---

runPCA	<i>Run PCA scores.</i>
--------	------------------------

---

**Description**

Compute the projection of either samples or time series features onto a PCA space. In case, of PCA for features, the PCA can be computed for individual sample group as indicated or for samples from all groups. In either, case the data is first collapsed over replicates, so that each gene is represented as a vector of a single time course.

**Usage**

```
runPCA(
  object,
  collapse.replicates = FALSE,
  groups.selected = NULL,
  var.stabilize.method = "log1p"
)
```

**Arguments**

`object` A TimeSeriesExperiment object

`collapse.replicates` Whether PCA should be computed on the data with replicates aggregated.

`groups.selected` An optional character string indicating a particular group of samples PCA should be applied to. By default set to NULL and all groups are included.

`var.stabilize.method` Method for variance stabilization (VST). Currently, supports "none" (no VST), "log1p" (log plus one), "asinh" (inverse hyperbolic sine) or "deseq" ([varianceStabilizingTransform](#) function from DESeq2 package). Default is "log1p".

**Value**

Returns TimeSeriesExperiment object with PCA results in `dim.red` slot, a lists containing matrices of coordinates `'pca_sample'`, and `'pca_features'`, as well as a vector `'pca_eigs'`.

**Examples**

```
data("endoderm_small")
endoderm_small <- runPCA(endoderm_small)
head(dimensionReduction(endoderm_small, "pca_sample")[, 1:5])
head(dimensionReduction(endoderm_small, "pca_eigs"))
```

---

```
show, TimeSeriesExperiment-method
  show method for TimeSeriesExperiment
```

---

**Description**

show method for TimeSeriesExperiment

**Usage**

```
## S4 method for signature 'TimeSeriesExperiment'
show(object)
```

**Arguments**

object            A TimeSeriesExperiment object

**Value**

nothing, just prints to console

---

timepointDE            *Differential timepoint expression testing.*

---

**Description**

This is a wrapper around `lmFit` and `voom` functions from `limma` package for testing differential expression at specified timepoints.

**Usage**

```
timepointDE(object, timepoints = "all", min_gene_sum = 1, alpha = 0.05)
```

**Arguments**

object            A TimeSeriesExperiment object.  
timepoints        Vector of timepoints to test at.  
min\_gene\_sum     A scalar for filtering sparse genes before DE testing. Default is 1.  
alpha            A scalar for level of significance. Default is 0.05.

**Value**

a TimeSeriesExperiment object with timepoint differential expression testing results stored in 'timepoint\_de' element in `diff.expr` slot.

**Examples**

```
data("endoderm_small")  
endoderm_small <- timepointDE(endoderm_small, timepoint = 1.0)  
head(differentialExpression(endoderm_small, "timepoint_de")$`1`)
```

---

timepoints	<i>Timepoint information</i>
------------	------------------------------

---

**Description**

Timepoint information

**Usage**

```
timepoints(object, ...)

## S4 method for signature 'TimeSeriesExperiment'
timepoints(object)

timepoints(object, ...) <- value

## S4 replacement method for signature 'TimeSeriesExperiment'
timepoints(object) <- value
```

**Arguments**

object	a TimeSeriesExperiment object.
...	arguments to other functions.
value	a numeric vector with new time information.

**Value**

a numeric vector

**Examples**

```
data("endoderm_small")
head(timepoints(endoderm_small))
timepoints(endoderm_small) <- sample(1:ncol(endoderm_small))
head(timepoints(endoderm_small))
```

---

timeSeries	<i>Time series formatted data.</i>
------------	------------------------------------

---

**Description**

Getter and setter methods for timeSeries slot of a TimeSeriesExperiment object.



**Usage**

```
timeSeries(object, ...)

## S4 method for signature 'TimeSeriesExperiment'
timeSeries(object, name = NULL)

timeSeries(object, ...) <- value

## S4 replacement method for signature 'TimeSeriesExperiment'
timeSeries(object) <- value
```

**Arguments**

object	a TimeSeriesExperiment object.
...	arguments to other functions.
name	a character string, one of 'ts', 'ts_with_lags', 'ts_collapsed' and 'ts_collapsed_with_lags'. If NULL, all elements are returned.
value	replacement list

**Details**

timeSeries slot is a list with 'ts' and (optionally) 'ts\_collapsed' storing data formatted as time-series/time-courses.

**Value**

a data.frame

**Examples**

```
data("endoderm_small")
endoderm_small <- makeTimeSeries(endoderm_small)
head(timeSeries(endoderm_small))
head(timeSeries(endoderm_small, name = 'ts'))
```

---

TimeSeriesExperiment-class

*Time Series Experiment Class*

---

**Description**

TimeSeriesExperiment is an extension of SummarizedExperiment class with the following new slots (in addition to SummarizedExperiment slots).

Constructor for 'TimeSeriesExperiment' object which stores same data as SummarizedExperiment but also slots for time, group, replicate and other time-series formatted data useful for applying data-analysis.

**Usage**

```
TimeSeriesExperiment(
  ...,
  timepoint = numeric(0),
  group = character(0),
  replicate = character(0)
)
```

**Arguments**

...	For <a href="#">SummarizedExperiment:: SummarizedExperiment</a> , S4 methods list and matrix, arguments identical to those of the SimpleList method.
timepoint	a vector indicating timepoint at which each sample was collected or a character string equal to one of the column names of a supplied colData.
group	a vector indicating a group membership for each sample or a character string equal to one of the column names of a supplied colData. If not specified, the group is set to 'G1' for each sample.
replicate	a vector indicating a replicate id of each sample or a character string equal to one of the column names of a supplied colData. If not specified, the replicate is set to 'R1' for each sample.

**Details**

The TimeSeriesExperiment class is an object (the main data container) used for in the time series/course experiment analysis. It stores all relevant information associated with the dataset, including the raw data, group, replicate and time associated with each sample (column of the data). The object includes also slots for results from some class-specific methods.

TimeSeriesExperiment constructor initializes the TimeSeriesExperiment TimeSeriesExperiment object and populates the time, replicate, and group slots.

**Value**

Returns an initialized TimeSeriesExperiment object.

**Slots**

timepoint	A vector indicating the time-point of each sample collection.
group	A vector indicating the group membership of each sample.
replicate	A vector indicating the replicate id of each sample.
assayCollapsed	A matrix with assay data aggregated over replicates.
colDataCollapsed	A <a href="#">DataFrame</a> where rows correspond to samples aggregated over replicates and columns indicate group membership and time-point.
timeSeries	A list of time-course formatted data. Each element of the list is a <a href="#">DataFrame</a> with the first three columns indicating feature, group, replicate, and the remaining ones storing the assay data at consecutive time points. TimeSeriesExperiment methods will typically generate elements of the list named: 'ts', 'ts_collapsed'.

**dimensionReduction** A list of results from applying dimensionality reduction methods; elements named by technique used.

**clusterAssignment** A list of results from clustering of the time-series features (rows) containing elements: 'settings', 'hclust', 'clust\_map' and 'clust\_centroids'.

**differentialExpression** A list of results from differential expression analysis. Either from point-wise ('timepoint\_de') or trajectory ('trajectory\_de') differential expression analysis.

## Examples

```
raw <- matrix(runif(3000), ncol = 30)
timepoint <- rep(rep(1:5, each = 3), 2)
replicate <- rep(1:3, 10)
group <- rep(1:2, each = 15)
test_TimeSeriesExperiment <- TimeSeriesExperiment(
  assays = list(raw),
  timepoint = timepoint,
  replicate = replicate,
  group = group)
test_TimeSeriesExperiment
```

---

trajectoryDE

*Differential trajectory testing.*

---

## Description

Performs differential trajectory testing for timecourse data using [adonis](#) method.

## Usage

```
trajectoryDE(
  object,
  dist_method = "euclidean",
  p_adj_method = "BH",
  lambda = c(0.5, 0.25),
  verbose = TRUE,
  ...
)
```

## Arguments

<b>object</b>	A TimeSeriesExperiment object.
<b>dist_method</b>	the name of any method used in <code>vegdist</code> to calculate pairwise distances, "euclidean" by defaults.
<b>p_adj_method</b>	a correction method. See details in <a href="#">p.adjust</a> . Default is "BH".

lambda	Weights for each lag difference, for time-course data. Length of lambda specifies number of lags to include. Default is c(0.5, 0.25) for lag 1 and 2. Used only if 'timecourse.data' slot not initialized.
verbose	whether code comments should be printed. Default is TRUE.
...	other options to <code>adonis</code> function from <code>vegan</code> .

**Value**

a data.frame with adonis results for all features.

**Examples**

```
data("endoderm_small")
endoderm_small <- makeTimeSeries(endoderm_small)
## Not run:
  endoderm_small <- trajectoryDE(endoderm_small)
  head(differentialExpression(endoderm_small, "trajectory_de"))

## End(Not run)
```

---

varianceStabilization *Variance stabilization.*

---

**Description**

This function performs variance stabilization on assay data.

**Usage**

```
varianceStabilization(X, method = "asinh")
```

**Arguments**

X	an assay data matrix or data.frame where columns correspond
method	Method for variance stabilization (VST). Currently, supports "none" (no VST), "log1p" (log plus one), "asinh" (inverse hyperbolic sine) or "deseq" ( <a href="#">varianceStabilizingTransform</a> function from DESeq2 package). Default is "log1p".

**Value**

Returns a varianced stabilized data matrix.

**Examples**

```
X <- sapply(exp(rlnorm(10)), function(m) rbinom(20, size = 1, mu = m))
head(X)
Y <- varianceStabilization(X, method = "asinh")
head(Y)
```

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