

# Introduction to RBM package

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## 1 Overview

This document provides an introduction to the `RBM` package. The `RBM` package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the `RBM` package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

## 2 Getting started

The RBM package can be installed and loaded through the following R code.  
Install the RBM package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

## 3 RBM\_T and RBM\_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The  $p$ -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(myresult$permutation_p<=0.05)
```

```

[1] 24

> which(myresult$permutation_p<=0.05)

[1] 24 33 47 126 141 149 202 209 230 379 398 400 417 427 444 651 690 750 792
[20] 909 915 918 949 967

> sum(myresult$bootstrap_p<=0.05)

[1] 2

> which(myresult$bootstrap_p<=0.05)

[1] 202 445

> permutation_adj_p <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adj_p<=0.05)

[1] 1

> bootstrap_adj_p <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adj_p<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 21

> which(myresult2$bootstrap_p<=0.05)

[1] 27 70 77 123 240 306 370 406 464 521 665 679 702 704 727 802 829 836 873
[20] 922 942

> bootstrap2_adj_p <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adj_p<=0.05)

[1] 0

```

- Examples using the RBM\_F function: normdata\_F simulates a standardized gene expression data and unifdata\_F simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```

> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)

              Length Class  Mode
ordfit_t      3000   -none-  numeric
ordfit_pvalue 3000   -none-  numeric
ordfit_beta1   3000   -none-  numeric
permutation_p 3000   -none-  numeric
bootstrap_p    3000   -none-  numeric

> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 70

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 61

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 63

> which(myresult_F$permutation_p[, 1]<=0.05)

 [1]  12  25  30  34  44  54  88 103 104 125 128 171 178 197 207
[16] 231 253 255 282 294 314 317 323 325 326 340 349 360 364 370
[31] 372 395 414 418 422 435 448 460 462 468 479 498 510 532 557
[46] 593 612 621 630 632 667 669 696 766 793 847 854 855 856 922
[61] 929 939 949 953 966 976 982 987 992 1000

> which(myresult_F$permutation_p[, 2]<=0.05)

 [1]  12  25  30  34  44  54  93 103 104 125 128 171 207 209 231
[16] 253 255 269 282 294 314 317 323 326 349 360 364 370 372 395
[31] 414 418 422 439 448 460 462 468 479 498 518 532 557 612 630
[46] 667 766 793 847 854 856 858 939 949 953 976 982 983 987 992
[61] 1000

> which(myresult_F$permutation_p[, 3]<=0.05)

 [1]  12  25  30  34  44  54  61  79 103 104 125 128 171 175 207
[16] 231 253 255 282 294 314 317 323 325 326 349 360 364 370 372
[31] 395 414 422 435 447 448 460 462 468 479 498 510 532 557 612
[46] 667 766 793 814 819 847 854 855 856 929 939 949 953 976 982
[61] 987 992 1000

```

```

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

[1] 15

> con2_adjp <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adjp<=0.05/3)

[1] 17

> con3_adjp <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adjp<=0.05/3)

[1] 14

> which(con2_adjp<=0.05/3)

[1] 25 54 103 104 372 395 422 479 557 793 847 854 856 953 982
[16] 992 1000

> which(con3_adjp<=0.05/3)

[1] 12 54 103 104 231 253 282 370 372 422 462 793 953 982

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

              Length Class  Mode
ordfit_t      3000   -none- numeric
ordfit_pvalue 3000   -none- numeric
ordfit_beta1  3000   -none- numeric
permutation_p 3000   -none- numeric
bootstrap_p   3000   -none- numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 60

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 50

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 48

```

```

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 35 37 41 80 87 94 97 127 162 176 194 199 200 201 245 260 267 312 315
[20] 316 317 348 385 417 433 436 449 486 538 543 548 583 584 585 606 619 623 642
[39] 687 717 741 773 774 798 800 802 808 809 833 842 856 861 891 914 919 926 930
[58] 951 957 993

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 5 12 41 87 90 94 127 162 176 194 199 201 218 260 267 312 315 317 385
[20] 411 417 436 449 474 486 538 543 548 584 585 596 606 619 687 717 773 774 798
[39] 800 802 808 809 833 842 890 919 926 930 951 983

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 4 5 41 87 94 97 127 162 176 194 200 277 312 315 316 317 385 417 433
[20] 449 474 486 538 543 548 583 584 585 623 640 687 734 773 774 798 800 802 808
[39] 809 833 842 861 890 919 926 930 951 957

> con21_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adj_p<=0.05/3)

[1] 4

> con22_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adj_p<=0.05/3)

[1] 3

> con23_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adj_p<=0.05/3)

[1] 2

```

## 4 Ovarian cancer methylation example using the RBM\_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM\_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the RBM\_T function and presenting the results for further validation and investigations.

```

> system.file("data", package = "RBM")

[1] "/private/var/folders/r0/l4fjk6cj5xj0j3brt4bplpl40000gt/T/RtmphYntGX/Rinst2d7129819cf0/RBM/d

> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)

      IlmnID      Beta      exmdata2[, 2]      exmdata3[, 2]
cg00000292: 1  Min.   :0.01058  Min.   :0.01187  Min.   :0.009103
cg00002426: 1  1st Qu.:0.04111  1st Qu.:0.04407  1st Qu.:0.041543
cg00003994: 1  Median :0.08284  Median :0.09531  Median :0.087042
cg00005847: 1  Mean    :0.27397  Mean    :0.28872  Mean    :0.283729
cg00006414: 1  3rd Qu.:0.52135  3rd Qu.:0.59032  3rd Qu.:0.558575
cg00007981: 1  Max.    :0.97069  Max.    :0.96937  Max.    :0.970155
(Other)      :994              NA's     :4
exmdata4[, 2]      exmdata5[, 2]      exmdata6[, 2]      exmdata7[, 2]
Min.   :0.01019  Min.   :0.01108  Min.   :0.01937  Min.   :0.01278
1st Qu.:0.04092  1st Qu.:0.04059  1st Qu.:0.05060  1st Qu.:0.04260
Median :0.09042  Median :0.08527  Median :0.09502  Median :0.09362
Mean    :0.28508  Mean    :0.28482  Mean    :0.27348  Mean    :0.27563
3rd Qu.:0.57502  3rd Qu.:0.57300  3rd Qu.:0.52099  3rd Qu.:0.52240
Max.    :0.96658  Max.    :0.97516  Max.    :0.96681  Max.    :0.95974
              NA's     :1
exmdata8[, 2]
Min.   :0.01357
1st Qu.:0.04387
Median :0.09282
Mean    :0.28679
3rd Qu.:0.57217
Max.    :0.96268

> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)

      Length Class  Mode
ordfit_t      1000  -none- numeric
ordfit_pvalue 1000  -none- numeric
ordfit_beta0   1000  -none- numeric
ordfit_beta1   1000  -none- numeric
permutation_p  1000  -none- numeric
bootstrap_p    1000  -none- numeric

> sum(diff_results$ordfit_pvalue<=0.05)

[1] 47

```

```

> sum(diff_results$permutation_p<=0.05)

[1] 63

> sum(diff_results$bootstrap_p<=0.05)

[1] 68

> ordfit_adj_p <- p.adjust(diff_results$ordfit_pvalue, "BH")
> sum(ordfit_adj_p<=0.05)

[1] 0

> perm_adj_p <- p.adjust(diff_results$permutation_p, "BH")
> sum(perm_adj_p<=0.05)

[1] 3

> boot_adj_p <- p.adjust(diff_results$bootstrap_p, "BH")
> sum(boot_adj_p<=0.05)

[1] 7

> diff_list_perm <- which(perm_adj_p<=0.05)
> diff_list_boot <- which(boot_adj_p<=0.05)
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t[
> print(sig_results_perm)

      IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
848 cg00826384 0.05721674    0.05612171    0.06644259    0.06358381
851 cg00830029 0.58362500    0.59397870    0.64739610    0.67269640
911 cg00888479 0.07388961    0.07361080    0.10149800    0.09985076
      exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
848    0.05230160    0.06119713    0.06542751    0.06240686
851    0.50820240    0.34657470    0.66276570    0.64634510
911    0.08633986    0.06765189    0.09070268    0.12417730
      diff_results$ordfit_t[diff_list_perm]
848                                -1.687144
851                                -2.986319
911                                -3.490240
      diff_results$permutation_p[diff_list_perm]
848                                0
851                                0
911                                0

> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t[
> print(sig_results_boot)

```



	IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]	exmdata4[, 2]
95	cg00081975	0.03633894	0.04975194	0.06024723	0.05598723
146	cg00134539	0.61101320	0.53321780	0.45999340	0.46787420
280	cg00260778	0.64319890	0.60488960	0.56735060	0.53150910
632	cg00615377	0.11265030	0.16140570	0.19404450	0.17468600
677	cg00651216	0.06825629	0.12529090	0.14409190	0.13907250
833	cg00814580	0.09348613	0.09619816	0.12010440	0.11534240
979	cg00945507	0.13432250	0.23854600	0.34749760	0.28903340
	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]	exmdata8[, 2]	
95	0.04561792	0.05115624	0.06068253	0.06168212	
146	0.67191510	0.63137380	0.47929610	0.45428300	
280	0.61920530	0.61925200	0.46753250	0.55632410	
632	0.12573100	0.14483660	0.16338240	0.20130510	
677	0.07669587	0.09597587	0.11690440	0.15194540	
833	0.09577040	0.11598850	0.12860890	0.14111200	
979	0.11848510	0.16653850	0.30718420	0.26624740	
	diff_results\$ordfit_t[diff_list_boot]				
95	-2.654324				
146	5.636263				
280	4.337628				
632	-3.722206				
677	-3.457874				
833	-3.278186				
979	-4.968792				
	diff_results\$bootstrap_p[diff_list_boot]				
95	0				
146	0				
280	0				
632	0				
677	0				
833	0				
979	0				