

The REDseq user's guide

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October 27, 2023

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1 Introduction

Restriction Enzyme digestion (RED) followed by high throughput sequencing (REDseq) enables genome wide differentiation of highly accessible regions and inaccessible regions. Comparing the profiles of restriction enzyme (RE) digestion among different cell types, developmental stages, disease stages, or different tissues facilitates deciphering of complex regulation network of cell differentiation, developmental control, and disease etiology and progression. We have developed a Bioconductor package called *REDSeq* to address the fundamental upstream analysis tasks of REDseq dataset. We have implemented functions for building genomic map of restriction enzyme sites (`buildREmap`), assigning sequencing tags to

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RE sites (`assignSeq2REsite`), visualizing genome-wide distribution of differentially cut regions (`distanceHistSeq2RE`) and the distance distribution of sequence tags to corresponding RE sites (`distanceHistSeq2RE`), generating count table for identifying statistically significant RE sites (`summarizeByRE`). We have leveraged *BSgenome* on implementing function `buildREmap` for building genome-wide RE maps. The input data for `assignSeq2REsite` are represented as `GRanges`, for efficiently associating sequences with RE sites. It first identifies RE sites that have mapped sequence tags around the cut position taking consideration of user-defined offset, sequence length and strand in the aligned sequences. The user-defined offset guards against imperfect sticky end repair and primer addition process. These RE sites are used as seeds for assigning the remaining tags depending on which of five strategies the users select for partitioning sequences associated with multiple RE sites, i.e., unique, average, estimate, best and random. For experiment with at least two conditions with biological replicates, count summary generated from `summarizeByRE` can be easily used for identifying differentially cut RE sites using either *DESeq* or *edgeR*. Differentially cut RE sites can be annotated to the nearest gene using *ChIPpeakAnno*. In addition, for early stage experiments without replicates, `compareREDseq` outputs differentially cut RE sites between two experimental conditions using Fisher's Exact Test. For experiment with one experimental condition, `binom.test.REDseq` outputs differentially cut RE sites in the genome. Multiplicity adjustment functions from *multtest* package were integrated in both functions.

2 Examples of using REDseq

2.1 Task 1: Build a RE map for a genome

Given a fasta/fastq file containing the restriction enzyme recognition site and a *BSgenome* object, the function `buildREmap` builds a genome-wide RE map.

```
> library(REDseq)
> REpatternFilePath = system.file("extdata", "examplePattern.fa", package="REDseq")
> library(BSgenome.Celegans.UCSC.ce2)
> myMap = buildREmap( REpatternFilePath, BSgenomeName=Celegans, outfile="example.REmap")

>>> Finding all hits in sequences chrI ...
>>> DONE searching
>>> Finding all hits in sequences chrII ...
>>> DONE searching
>>> Finding all hits in sequences chrIII ...
>>> DONE searching
>>> Finding all hits in sequences chrIV ...
>>> DONE searching
>>> Finding all hits in sequences chrV ...
>>> DONE searching
>>> Finding all hits in sequences chrX ...
>>> DONE searching
>>> Finding all hits in sequences chrM ...
>>> DONE searching
```

2.2 Task 2: Assign mapped sequence tags to RE site

Given a mapped sequence tags as a GRanges and REmap as a Granges, `assignSeq2REsite` function assigns mapped sequence tags to RE site depending on the strategy users select. There are five strategies implemented, i.e., unique, average, estimate, best and random. For details, type `help(assignSeq2REsite)` in a R session.

```
> data(example.REDseq)
> data(example.map)
> r.unique = assignSeq2REsite(example.REDseq, example.map, cut.offset = 1,
+ seq.length = 36, allowed.offset = 5, min.FragmentLength = 60,
+ max.FragmentLength = 300, partitionMultipleRE = "unique")

Fri Oct 27 05:51:46 2023 Validating input ...
Fri Oct 27 05:51:46 2023 Prepare map data ...
Fri Oct 27 05:51:46 2023 Align to chromosome 2 ...
Fri Oct 27 05:51:47 2023 Finished 1st round of aligning! Start the 2nd round of aligning ...
Fri Oct 27 05:51:47 2023 Align to chromosome 2 ...
Fri Oct 27 05:51:47 2023 Start filtering ...

> r.best = assignSeq2REsite(example.REDseq, example.map,
+ cut.offset = 1, seq.length = 36, allowed.offset = 5,
+ min.FragmentLength = 60, max.FragmentLength = 300, partitionMultipleRE = "best")

Fri Oct 27 05:51:47 2023 Validating input ...
Fri Oct 27 05:51:47 2023 Prepare map data ...
Fri Oct 27 05:51:47 2023 Align to chromosome 2 ...
Fri Oct 27 05:51:47 2023 Finished 1st round of aligning! Start the 2nd round of aligning ...
Fri Oct 27 05:51:47 2023 Align to chromosome 2 ...
Fri Oct 27 05:51:47 2023 Start filtering ...
Fri Oct 27 05:51:47 2023 Partitioning reads over RE sites within 300 ...
Fri Oct 27 05:51:47 2023 get count for each RE ...

> r.random = assignSeq2REsite(example.REDseq, example.map, cut.offset = 1,
+ seq.length = 36, allowed.offset = 5, min.FragmentLength = 60,
+ max.FragmentLength = 300, partitionMultipleRE = "random")

Fri Oct 27 05:51:47 2023 Validating input ...
Fri Oct 27 05:51:47 2023 Prepare map data ...
Fri Oct 27 05:51:47 2023 Align to chromosome 2 ...
Fri Oct 27 05:51:47 2023 Finished 1st round of aligning! Start the 2nd round of aligning ...
Fri Oct 27 05:51:47 2023 Align to chromosome 2 ...
Fri Oct 27 05:51:47 2023 Start filtering ...
Fri Oct 27 05:51:47 2023 Partitioning reads over RE sites within 300 ...

> r.average = assignSeq2REsite(example.REDseq, example.map, cut.offset = 1,
+ seq.length = 36, allowed.offset = 5, min.FragmentLength = 60,
+ max.FragmentLength = 300, partitionMultipleRE = "average")

Fri Oct 27 05:51:47 2023 Validating input ...
Fri Oct 27 05:51:47 2023 Prepare map data ...
Fri Oct 27 05:51:47 2023 Align to chromosome 2 ...
Fri Oct 27 05:51:47 2023 Finished 1st round of aligning! Start the 2nd round of aligning ...
Fri Oct 27 05:51:47 2023 Align to chromosome 2 ...
Fri Oct 27 05:51:47 2023 Start filtering ...
Fri Oct 27 05:51:47 2023 Partitioning reads over RE sites within 300 ...

> r.estimate = assignSeq2REsite(example.REDseq, example.map, cut.offset = 1,
+ seq.length = 36, allowed.offset = 5, min.FragmentLength = 60,
+ max.FragmentLength = 300, partitionMultipleRE = "estimate")
```

```

Fri Oct 27 05:51:47 2023 Validating input ...
Fri Oct 27 05:51:47 2023 Prepare map data ...
Fri Oct 27 05:51:47 2023 Align to chromosome 2 ...
Fri Oct 27 05:51:47 2023 Finished 1st round of aligning! Start the 2nd round of aligning ...
Fri Oct 27 05:51:47 2023 Align to chromosome 2 ...
Fri Oct 27 05:51:47 2023 Start filtering ...
Fri Oct 27 05:51:47 2023 Partitioning reads over RE sites within 300 ...
Fri Oct 27 05:51:47 2023 get count for each RE ...

```

```
> head(r.estimate$passed.filter)
```

	SEQid	REid	Chr	strand	SEQstart	SEQend	REstart	REend
2	00000037	Sau96I.chr10.29	2	+	3012091	3012126	3012090	3012094
3	00000038	Sau96I.chr10.29	2	+	3012096	3012131	3012090	3012094
5	00000040	Sau96I.chr10.30	2	+	3012300	3012335	3012299	3012303
8	00000055	Sau96I.chr10.42	2	+	3018315	3018350	3018314	3018318
9	00000056	Sau96I.chr10.42	2	+	3018315	3018350	3018314	3018318
10	00000057	Sau96I.chr10.42	2	+	3018315	3018350	3018314	3018318

	Distance	Weight
2	1	1
3	6	1
5	1	1
8	1	1
9	1	1
10	1	1

The above examples are for single-end sequencing data. For paired-end sequencing data, please create inputS.RD and inputE.RD from input.RD first with start(input.RD) and end(input.RD), where inputS.RD contains the start of the input.RD and inputE.RD contains the end of the input.RD. Then call assignSeq2REsite twice with inputS.RD and inputE.RD respectively. Please set min.FragmentLength = 0, max.FragmentLength = 1, seq.length = 1 with both calls.

2.3 Task 3: Visualize the distribution of cut frequency in selected genomic regions and the distance distribution of sequence tags to corresponding RE sites

```
> data(example.assignedREDseq)
```

```
> plotCutDistribution(example.assignedREDseq,example.map, chr="2",  
+ xlim =c(3012000, 3020000))
```

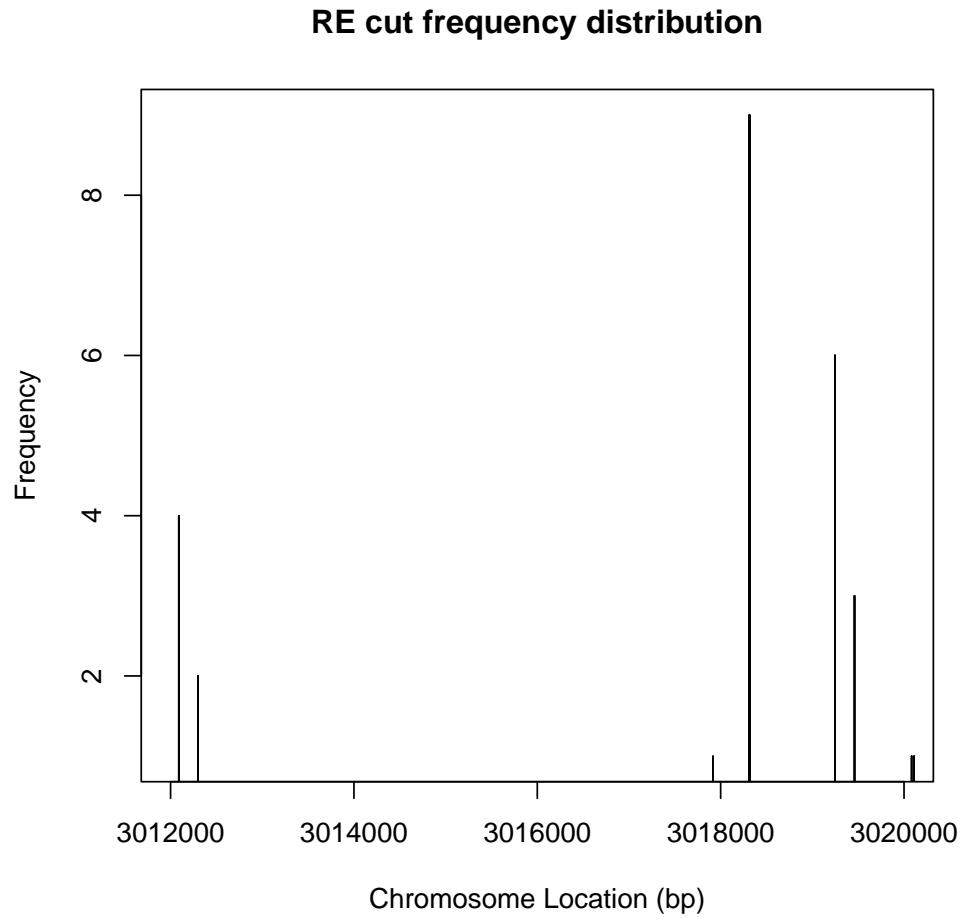


Figure 1: Plot to show the distribution of cut frequency in the selected genomic-regions with the function `plotCutDistribution`. The red triangle is the expected cut frequency for each RE site.

```
> distanceHistSeq2RE(example.assignedREDseq,ylim=c(0,25))
```

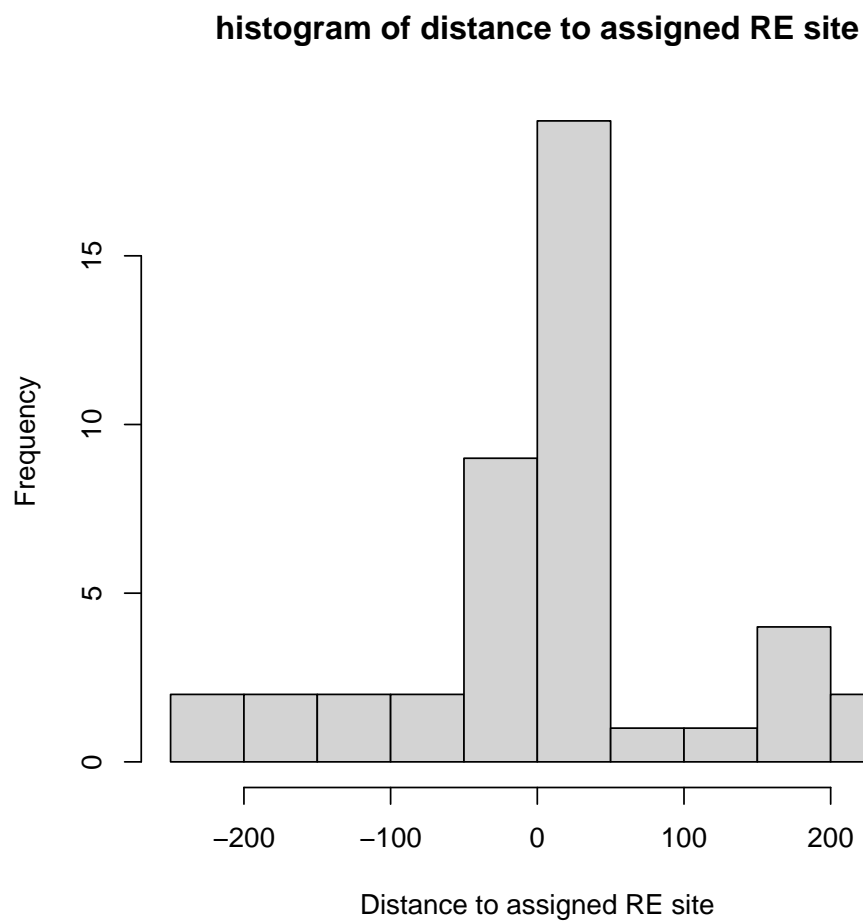


Figure 2: Plot to show the distribution of distance of sequence tags to associated RE sites with the function `distanceHistSeq2RE`.

2.4 Task 4: Generating count table for identifying statistically significant RE sites

Once you have obtained the assigned RE sites, you can use the function `summarizeByRE` to obtain a count table for identifying statistically significant RE sites using *DEseq* or *edgeR*.

```
> REsummary =summarizeByRE(example.assignedREDseq,by="Weight")
```

2.5 Task 5: Identifying differential cut RE sites for experiment with one experiment condition

```
> binom.test.REDseq(REsummary)
```

	p.value	total.weight.count	REid	cut.frequency
1	2.804822e-47	9	Sau96I.chr10.42	0.28125
2	9.061718e-31	6	Sau96I.chr10.43	0.18750
3	3.595919e-20	4	Sau96I.chr10.29	0.12500
4	4.959892e-15	3	Sau96I.chr10.50	0.09375
5	4.959892e-15	3	Sau96I.chr10.45	0.09375
6	4.959901e-10	2	Sau96I.chr10.30	0.06250
7	4.959901e-10	2	Sau96I.chr10.51	0.06250
8	3.199950e-05	1	Sau96I.chr10.40	0.03125
9	3.199950e-05	1	Sau96I.chr10.49	0.03125
10	3.199950e-05	1	Sau96I.chr10.47	0.03125

	BH.adjusted.p.value
1	2.804822e-46
2	4.530859e-30
3	1.198640e-19
4	9.919784e-15
5	9.919784e-15
6	7.085573e-10
7	7.085573e-10
8	3.199950e-05
9	3.199950e-05
10	3.199950e-05

2.6 Task 6: Identifying differential cut RE sites for early stage experiment without replicates

```
> x= cbind(c("RE1", "RE2", "RE3", "RE4"), c(10,1,100, 0),c(5,5,50, 40))
> colnames(x) = c("REid", "control", "treated")
> compareREDseq(x)
```

	p.value	control.count	treated.count	REid	control.total	treated.total
1	6.233642e-16	0	40	RE4	111	100
2	1.159388e-10	100	50	RE3	111	100
3	1.035503e-01	1	5	RE2	111	100
4	2.943364e-01	10	5	RE1	111	100

	odds.ratio	BH.adjusted.p.value
1	Inf	2.493457e-15
2	0.1112945	2.318777e-10
3	5.7478720	1.380671e-01
4	0.5331227	2.943364e-01

3 References

1. Roberts, R.J., Restriction endonucleases. CRC Crit Rev Biochem, 1976. 4(2): p. 123-64.
2. Kessler, C. and V. Manta, Specificity of restriction endonucleases and DNA modification methyltransferases a review (Edition 3). Gene, 1990. 92(1-2): p. 1-248.
3. Pingoud, A., J. Alves, and R. Geiger, Restriction enzymes. Methods Mol Biol, 1993. 16: p. 107-200.
4. bibitemAnders10 Anders, S. and W. Huber, Differential expression analysis for sequence count data. Genome Biol, 2010. 11(10): p. R106.
5. Robinson, M.D., D.J. McCarthy, and G.K. Smyth, edgeR: a Bioconductor package for differential expression analysis of digital gene expression data. Bioinformatics, 2010. 26(1): p. 139-40.
6. Zhu, L.J., et al., ChIPpeakAnno: a Bioconductor package to annotate ChIP-seq and ChIP-chip data. BMC Bioinformatics, 2010. 11: p. 237.
7. Pages, H., BSgenome package. <http://bioconductor.org/packages/2.8/bioc/vignettes/BSgenome/inst/doc/GenomeSearching.pdf>
8. Zhu, L.J., et al., REDseq: A Bioconductor package for Analyzing High Throughput Sequencing Data from Restriction Enzyme Digestion. (In preparation)

4 Session Info

```
> sessionInfo()
```

```
R version 4.3.1 (2023-06-16)  
Platform: aarch64-apple-darwin20 (64-bit)  
Running under: macOS Ventura 13.6.1
```

```
Matrix products: default
```

```
BLAS: /Library/Frameworks/R.framework/Versions/4.3-arm64/Resources/lib/libRblas.0.dylib  
LAPACK: /Library/Frameworks/R.framework/Versions/4.3-arm64/Resources/lib/libRlapack.dylib
```

```
locale:
```

```
[1] C/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
```

```
time zone: America/New_York
```

```
tzcode source: internal
```


attached base packages:

```
[1] stats4      stats      graphics  grDevices  utils      datasets  methods
[8] base
```

other attached packages:

```
[1] REDseq_1.48.0           ChIPpeakAnno_3.36.0
[3] multtest_2.58.0         Biobase_2.62.0
[5] BSgenome.Celegans.UCSC.ce2_1.4.0 BSgenome_1.70.0
[7] rtracklayer_1.62.0      BiocIO_1.12.0
[9] Biostrings_2.70.1       XVector_0.42.0
[11] GenomicRanges_1.54.0    GenomeInfoDb_1.38.0
[13] IRanges_2.36.0          S4Vectors_0.40.1
[15] BiocGenerics_0.48.0
```

loaded via a namespace (and not attached):

```
[1] tidyselect_1.2.0        dplyr_1.1.2
[3] blob_1.2.4              filelock_1.0.2
[5] bitops_1.0-7            fastmap_1.1.1
[7] RCurl_1.98-1.12         BiocFileCache_2.10.1
[9] GenomicAlignments_1.38.0 XML_3.99-0.14
[11] digest_0.6.33           lifecycle_1.0.3
[13] survival_3.5-5          KEGGREST_1.42.0
[15] VennDiagram_1.7.3       RSQLite_2.3.1
[17] magrittr_2.0.3          compiler_4.3.1
[19] rlang_1.1.1             progress_1.2.2
[21] tools_4.3.1             utf8_1.2.3
[23] yaml_2.3.7              lambda.r_1.2.4
[25] prettyunits_1.1.1       S4Arrays_1.2.0
[27] bit_4.0.5               curl_5.0.1
[29] DelayedArray_0.28.0     regioneR_1.34.0
[31] xml2_1.3.5              abind_1.4-5
[33] BiocParallel_1.36.0     grid_4.3.1
[35] fansi_1.0.4             colorspace_2.1-0
[37] ggplot2_3.4.2           scales_1.2.1
[39] MASS_7.3-60             biomaRt_2.58.0
[41] SummarizedExperiment_1.32.0 cli_3.6.1
[43] crayon_1.5.2            generics_0.1.3
[45] httr_1.4.6              rjson_0.2.21
[47] DBI_1.1.3               cachem_1.0.8
[49] stringr_1.5.0           zlibbioc_1.48.0
[51] splines_4.3.1           parallel_4.3.1
[53] AnnotationDbi_1.64.0    formatR_1.14
```

[55]	restfulr_0.0.15	matrixStats_1.0.0
[57]	vctrs_0.6.3	Matrix_1.6-0
[59]	hms_1.1.3	bit64_4.0.5
[61]	RBGL_1.78.0	GenomicFeatures_1.54.0
[63]	glue_1.6.2	codetools_0.2-19
[65]	gtable_0.3.3	stringi_1.7.12
[67]	futile.logger_1.4.3	munsell_0.5.0
[69]	tibble_3.2.1	pillar_1.9.0
[71]	rappdirs_0.3.3	graph_1.80.0
[73]	GenomeInfoDbData_1.2.10	R6_2.5.1
[75]	dbplyr_2.3.3	lattice_0.21-8
[77]	futile.options_1.0.1	png_0.1-8
[79]	Rsamtools_2.18.0	memoise_2.0.1
[81]	Rcpp_1.0.11	InteractionSet_1.30.0
[83]	SparseArray_1.2.0	MatrixGenerics_1.14.0
[85]	pkgconfig_2.0.3	