

# Package ‘openCyto’

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**Description** This package is designed to facilitate the automated gating methods in sequential way to mimic the manual gating strategy.

**License** Artistic-2.0

**LazyLoad** yes

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as.data.table                      *convert a gatingTemplate object to a data.table*

**Description**

It is the inverse function of gatingTemplate constructor.

**Usage**

```
## S3 method for class 'gatingTemplate'
as.data.table(x, keep.rownames = FALSE)
```

**Arguments**

x                      gatingTemplate object  
 keep.rownames      not used

**Value**

a data.table

boolMethod-class                      *A class to represent a boolean gating method.*

**Description**

It extends refGate class.

---

dims,gtMethod-method    *get gating method dimensions*

---

### Description

get gating method dimensions

### Usage

```
## S4 method for signature 'gtMethod'
dims(x)
```

### Arguments

x                    gtMethod

---

dummyMethod-class    *A class to represent a dummy gating method that does nothing but serves as reference to be referred by other population*

---

### Description

It is generated automatically by the csv template preprocessing to handle the gating function that returns multiple gates.

---

fcEllipsoidGate        *constuctor for fcEllipsoidGate*

---

### Description

constuctor for fcEllipsoidGate

### Usage

```
fcEllipsoidGate(x, priors, posts)
```

### Arguments

x                    a ellipsoidGate object  
 priors              a list storing priors  
 posts                a list storing posteriors

---

fcEllipsoidGate-class *a concrete class that represents the ellipsoidGate generated by flowClust*

---

**Description**

It stores priors and posteriors as well as the actual ellipsoidGate.

---

fcFilter-class *a virtual class that represents the gating result generated by flowClust gating function*

---

**Description**

Basically it extends flowCore 'filter classes to have extra slot to store priors and posteriors

---

fcFilterList *constructor for fcFilterList*

---

**Description**

constructor for fcFilterList

**Usage**

fcFilterList(x)

**Arguments**

x *list of fcFilter (i.e. fcPolygonGate or fcRectangleGate)*

---

fcFilterList-class *a class that extends filterList class.*

---

**Description**

Each filter in the filterList must extends the fcFilter class

---

fcPolygonGate      *constructor for fcPolygonGate*

---

### Description

constructor for fcPolygonGate

### Usage

fcPolygonGate(x, priors, posts)

### Arguments

x	a polygonGate object
priors	a list storing priors
posts	a list storing posteriors

---

fcPolygonGate-class      *a concrete class that reprints the polygonGate generated by flowClust*

---

### Description

It stores priors and posteriors as well as the actual polygonGate.

---

fcRectangleGate      *constructor for fcRectangleGate*

---

### Description

constructor for fcRectangleGate

### Usage

fcRectangleGate(x, priors, posts)

### Arguments

x	a rectangleGate object
priors	a list storing priors
posts	a list storing posteriors

---

fcRectangleGate-class *a concrete class that represents the rectangleGate generated by flowClust*

---

### Description

It stores priors and posteriors as well as the actual rectangleGate.

---

fcTree *constructor of fcTree*

---

### Description

It adds an extra node data slot "fList"(which is a filterList object) to the gatingTemplate

### Usage

fcTree(gt)

### Arguments

gt *a gatingTemplate object*

---

fcTree-class *A class to represent a flowClust tree.*

---

### Description

It is a graphNEL used as a container to store priors and posteriors for each flowClust gate that can be visualized for the purpose of fine-tuning parameters for flowClust algorithm

---

gate\_flowclust\_1d *Applies flowClust to 1 feature to determine a cutpoint between the minimum cluster and all other clusters.*

---

### Description

We cluster the observations in fr into K clusters.

**Usage**

```

gate_flowclust_1d(
  fr,
  params,
  filterId = "",
  K = NULL,
  trans = 0,
  min.count = -1,
  max.count = -1,
  nstart = 1,
  prior = NULL,
  criterion = c("BIC", "ICL"),
  cutpoint_method = c("boundary", "min_density", "quantile", "posterior_mean",
    "prior_density"),
  neg_cluster = 1,
  cutpoint_min = NULL,
  cutpoint_max = NULL,
  min = NULL,
  max = NULL,
  quantile = 0.99,
  quantile_interval = c(0, 10),
  plot = FALSE,
  debug = FALSE,
  ...
)

```

**Arguments**

fr	a flowFrame object
params	character channel to be gated on
filterId	A character string that identifies the filter created.
K	the number of clusters to find
trans, min.count, max.count, nstart	some flowClust parameters. see <a href="#">flowClust</a>
prior	list of prior parameters for the Bayesian <a href="#">flowClust</a> . If NULL, no prior is used.
criterion	a character string stating the criterion used to choose the best model. May take either "BIC" or "ICL". This argument is only relevant when K is NULL or if length(K) > 1. The value selected is passed to <a href="#">flowClust</a> .
cutpoint_method	How should the cutpoint be chosen from the fitted <a href="#">flowClust</a> model? See Details.
neg_cluster	integer. The index of the negative cluster. The cutpoint is computed between clusters neg_cluster and neg_cluster + 1.
cutpoint_min	numeric value that sets a minimum threshold for the cutpoint. If a value is provided, any cutpoint below this value will be set to the given minimum value. If NULL (default), there is no minimum cutpoint value.
cutpoint_max	numeric value that sets a maximum threshold for the cutpoint. If a value is provided, any cutpoint above this value will be set to the given maximum value. If NULL (default), there is no maximum cutpoint value.



min	a numeric value that sets the lower bound for data filtering. If NULL (default), no truncation is applied.
max	a numeric value that sets the upper bound for data filtering. If NULL (default), no truncation is applied.
quantile	the quantile for which we will find the cutpoint using the quantile cutpoint_method. If the cutpoint_method is not set to quantile, this argument is ignored.
quantile_interval	a vector of length 2 containing the end-points of the interval of values to find the quantile cutpoint. If the cutpoint_method is not set to quantile, this argument is ignored.
plot	logical value indicating that the fitted <code>flowClust</code> model should be plotted along with the cutpoint
debug	logical indicating whether to carry the prior and posteriors with the gate for debugging purpose. Default is FALSE.
...	additional arguments that are passed to <code>flowClust</code>

### Details

By default, the cutpoint is chosen to be the boundary of the first two clusters. That is, between the first two cluster centroids, we find the midpoint between the largest observation from the first cluster and the smallest observations from the second cluster. Alternatively, if the `cutpoint_method` is `min_density`, then the cutpoint is the point at which the density between the first and second smallest cluster centroids is minimum.

### Value

a `rectangleGate` object consisting of all values beyond the cutpoint calculated

### Examples

```
## Not run:
gate <- gate_flowclust_1d(fr, params = "APC-A", K = 2) # fr is a flowFrame

## End(Not run)
```

---

gate_flowclust_2d	<i>Automatic identification of a population of interest via flowClust based on two markers</i>
-------------------	--

---

### Description

We cluster the observations in `fr` into `K` clusters. We set the cutpoint to be the point at which the density between the first and second smallest cluster centroids is minimum.

**Usage**

```

gate_flowclust_2d(
  fr,
  xChannel,
  yChannel,
  filterId = "",
  K = 2,
  usePrior = "no",
  prior = list(NA),
  trans = 0,
  min.count = -1,
  max.count = -1,
  nstart = 1,
  plot = FALSE,
  target = NULL,
  transitional = FALSE,
  quantile = 0.9,
  translation = 0.25,
  transitional_angle = NULL,
  min = NULL,
  max = NULL,
  ...
)

```

**Arguments**

fr	a flowFrame object
xChannel, yChannel	character specifying channels to be gated on
filterId	A character string that identifies the filter created.
K	the number of clusters to find
usePrior	Should we use the Bayesian version of <a href="#">flowClust</a> ? Answers are "yes", "no", or "vague". The answer is passed along to <a href="#">flowClust</a> .
prior	list of prior parameters for the Bayesian version of <a href="#">flowClust</a> . If usePrior is set to no, then the list is unused.
trans, min.count, max.count, nstart	some flowClust parameters. see <a href="#">flowClust</a>
plot	a logical value indicating if the fitted mixture model should be plotted. By default, no.
target	a numeric vector of length 2 (number of dimensions) containing the location of the cluster of interest. See details.
transitional	logical value indicating if a transitional gate should be constructed from the target <a href="#">flowClust</a> cluster. By default, no.
quantile	the contour level of the target cluster from the <a href="#">flowClust</a> fit to construct the gate
translation	a numeric value between 0 and 1 used to position a transitional gate if transitional = TRUE. This argument is ignored if transitional = FALSE. See details

transitional_angle	the angle (in radians) of the transitional gate. It is also used to determine which quadrant the final gate resides in. See details. Ignored if transitional = FALSE.
min	A vector of length 2. Truncate observations less than this minimum value. The first value truncates the xChannel, and the second value truncates the yChannel. By default, this vector is NULL and is ignored.
max	A vector of length 2. Truncate observations greater than this maximum value. The first value truncates the xChannel, and the second value truncates the yChannel. By default, this vector is NULL and is ignored.
...	additional arguments that are passed to <code>flowClust</code>

### Details

The cluster for the population of interest is selected as the one with cluster centroid nearest the target in Euclidean distance. By default, the largest cluster (i.e., the cluster with the largest proportion of observations) is selected as the population of interest.

We also provide the option of constructing a transitional gate from the selected population of interest. The location of the gate can be controlled with the `translation` argument, which translates the gate along the major axis of the target cluster as a function of the appropriate chi-squared coefficient. The larger translation is, the more gate is shifted in a positive direction. Furthermore, the width of the transitional gate can be controlled with the `quantile` argument.

The direction of the transitional gate can be controlled with the `transitional_angle` argument. By default, it is NULL, and we use the eigenvector of the target cluster that points towards the first quadrant (has positive slope). If `transitional_angle` is specified, we rotate the eigenvectors so that the angle between the x-axis (with the cluster centroid as the origin) and the major eigenvector (i.e., the eigenvector with the larger eigenvalue) is `transitional_angle`. So based on range that the angle falls in, the final `rectangleGate` will be constructed at the corresponding quadrant. i.e. Clockwise,  $[0, \pi/2]$  UR,  $(\pi/2, \pi]$  LR,  $(\pi, 3/2 * \pi]$  LL,  $(3/2 * \pi, 2 * \pi]$  UL

### Value

a `polygonGate` object containing the contour (ellipse) for 2D gating.

### Examples

```
## Not run:
gate <- gate_flowclust_2d(fr, xChannel = "FSC-A", yChannel = "SSC-A", K = 3) # fr is a flowFrame

## End(Not run)
```

---

gate_mindensity	<i>Determines a cutpoint as the minimum point of a kernel density estimate between two peaks</i>
-----------------	--

---

### Description

We fit a kernel density estimator to the cells in the `flowFrame` and identify the two largest peaks. We then select as the cutpoint the value at which the minimum density is attained between the two peaks of interest.

**Usage**

```

gate_mindensity(
  fr,
  channel,
  filterId = "",
  positive = TRUE,
  pivot = FALSE,
  gate_range = NULL,
  min = NULL,
  max = NULL,
  peaks = NULL,
  ...
)

```

**Arguments**

fr	a flowFrame object
channel	TODO
filterId	TODO
positive	If TRUE, then the gate consists of the entire real line to the right of the cutpoint. Otherwise, the gate is the entire real line to the left of the cutpoint. (Default: TRUE)
pivot	logical value. If TRUE, we choose as the two peaks the largest peak and its neighboring peak. See details.
gate_range	numeric vector of length 2. If given, this sets the bounds on the gate applied. If no gate is found within this range, we set the gate to the minimum value within this range if positive is TRUE and the maximum value of the range otherwise.
min	a numeric value that sets the lower boundary for data filtering
max	a numeric value that sets the upper boundary for data filtering
peaks	numeric vector. If not given, then perform peak detection first by .find_peaks
...	Additional arguments for peak detection.

**Details**

In the default case, the two peaks of interest are the two largest peaks obtained from the `link{density}` function. However, if `pivot` is TRUE, we choose the largest peak and its neighboring peak as the two peaks of interest. In this case, the neighboring peak is the peak immediately to the left of the largest peak if `positive` is TRUE. Otherwise, the neighboring peak is selected as the peak to the right.

In the special case that there is only one peak, we are conservative and set the cutpoint as the `min(x)` if `positive` is TRUE, and the `max(x)` otherwise.

**Value**

a `rectangleGate` object based on the minimum density cutpoint

**Examples**

```

## Not run:
gate <- gate_mindensity(fr, channel = "APC-A") # fr is a flowFrame

## End(Not run)

```

---

gate_mindensity2	<i>An improved version of mindensity used to determines a cutpoint as the minimum point of a kernel density estimate between two peaks.</i>
------------------	---

---

### Description

Analogous to the original `openCyto::mindensity()`, `mindensity2` operates on a standard `flowFrame`. Its behavior is closely modeled on the original `mindensity()` whenever possible. However, the underlying peak-finding algorithm (`improvedMindensity`) behaves significantly differently.

### Usage

```
gate_mindensity2(
  fr,
  channel,
  filterId = "",
  pivot = FALSE,
  gate_range = NULL,
  min = NULL,
  max = NULL,
  peaks = NULL,
  ...
)
```

### Arguments

<code>fr</code>	a <code>flowFrame</code> object
<code>channel</code>	the channel to operate on
<code>filterId</code>	a name to refer to this filter
<code>pivot</code>	logical value. If <code>TRUE</code> , we choose as the two peaks the largest peak and its neighboring peak. See details.
<code>gate_range</code>	numeric vector of length 2. If given, this sets the bounds on the gate applied.
<code>min</code>	a numeric value that sets the lower boundary for data filtering
<code>max</code>	a numeric value that sets the upper boundary for data filtering
<code>peaks</code>	numeric vector. If not given, then perform peak detection first by <code>.find_peaks</code>
<code>...</code>	Additional arguments for peak detection.

### Value

a `rectangleGate` object based on the minimum density cutpoint

### Author(s)

Greg Finak, Phu T. Van

### Examples

```
## Not run:
gate <- gate_mindensity2(fr, channel = "APC-A") # fr is a flowFrame

## End(Not run)
```

---

gate\_quad\_sequential    *sequential quadrant gating function*

---

### Description

The order of 1d-gating is determined so that the gates better capture the distributions of flow data.

### Usage

```
gate_quad_sequential(fr, channels, gFunc, min = NULL, max = NULL, ...)
```

### Arguments

fr	flowFrame
channels	character two channels used for gating
gFunc	the name of the 1d-gating function to be used for either dimension
min	a numeric vector that sets the lower bounds for data filtering
max	a numeric vector that sets the upper bounds for data filtering
...	other arguments passed to <code>.find_peak</code> (e.g. 'num_peaks' and 'adjust'). see <a href="#">tailgate</a>

### Value

a filters that contains four rectangleGates

---

gate\_quad\_tmix    *quadGate based on flowClust::tmixFiler*

---

### Description

This gating method identifies two quadrants (first, and third quadrants) by fitting the data with tmixture model. It is particularly useful when the two markers are not well resolved thus the regular quadGate method based on 1d gating will not find the perfect cut points on both dimensions.

### Usage

```
gate_quad_tmix(
  fr,
  channels,
  K,
  usePrior = "no",
  prior = list(NA),
  quantile1 = 0.8,
  quantile3 = 0.8,
  trans = 0,
  plot = FALSE,
  ...
)
```

**Arguments**

fr	flowFrame
channels	character vector specifies two channels
K	see <a href="#">gate_flowclust_2d</a>
usePrior	see <a href="#">gate_flowclust_2d</a>
prior	see <a href="#">gate_flowclust_2d</a>
quantile1	numeric specifies the quantile level(see 'level' in <a href="#">flowClust</a> ) for the first quadrant (x-y+)
quantile3	numeric specifies the quantile level see 'level' in <a href="#">flowClust</a> for third quadrant (x+y-)
trans	see <a href="#">gate_flowclust_2d</a>
plot	logical whether to plot flowClust clustering results
...	other arguments passed to <a href="#">flowClust</a>

**Value**

a filters object that contains four polygonGates following the order of (-+,++,+,-)

---

gate_quantile	<i>Determine the cutpoint by the events quantile.</i>
---------------	---

---

**Description**

It is possible that the cutpoint calculated by quantile function may not produce the exact the probability set by 'probs' argument if there are not enough cell events to reach that precision. Sometime the difference could be significant.

**Usage**

```
gate_quantile(
  fr,
  channel,
  probs = 0.999,
  plot = FALSE,
  filterId = "",
  min = NULL,
  max = NULL,
  ...
)
```

**Arguments**

fr	a flowFrame object
channel	the channel from which the cytokine gate is constructed
probs	probabilities passed to 'stats::quantile' function.
plot	whether to plot the gate result
filterId	the name of the filter
min	a numeric value that sets the lower boundary for data filtering
max	a numeric value that sets the upper boundary for data filtering
...	additional arguments passed to 'stats::quantile' function.

**Value**

a rectangleGate

**Examples**

```
## Not run:
gate <- gate_quantile(fr, Channel = "APC-A", probs = 0.995) # fr is a flowFrame

## End(Not run)
```

---

gate_tail	<i>Gates the tail of a density using the derivative of a kernel density estimate</i>
-----------	--

---

**Description**

These methods aim to set a one-dimensional gate (cutpoint) near the edge of a peak in the density specified by a channel of a [flowFrame](#) to isolate the tail population. They allow two approaches to do this, both beginning by obtaining a smoothed kernel density estimate (KDE) of the original density and then utilizing either its first or second derivative.

We determine a gating cutpoint using either the first or second derivative of the kernel density estimate (KDE) of the  $x$ .

**Usage**

```
gate_tail(
  fr,
  channel,
  filterId = "",
  num_peaks = 1,
  ref_peak = 1,
  strict = TRUE,
  tol = 0.01,
  side = "right",
  min = NULL,
  max = NULL,
  bias = 0,
  positive = TRUE,
  ...
)

.cytokine_cutpoint(
  x,
  num_peaks = 1,
  ref_peak = 1,
  method = c("first_deriv", "second_deriv"),
  tol = 0.01,
  adjust = 1,
  side = "right",
  strict = TRUE,
  plot = FALSE,
```



```

    auto_tol = FALSE,
    ...
)

```

### Arguments

fr	a flowFrame object
channel	the channel from which the cytokine gate is constructed
filterId	the name of the filter
num_peaks	the number of peaks expected to see. This effectively removes any peaks that are artifacts of smoothing
ref_peak	After num_peaks are found, this argument provides the index of the reference population from which a gate will be obtained. By default, the peak farthest to the left is used.
strict	logical when the actual number of peaks detected is less than ref_peak. an error is reported by default. But if strict is set to FALSE, then the reference peak will be reset to the peak of the far right.
tol	the tolerance value
side	On which side of the density do we want to gate the tail. Valid options are "left" or "right".
min	a numeric value that sets the lower boundary for data filtering
max	a numeric value that sets the upper boundary for data filtering
bias	a numeric value that adds a constant to the calculated cutpoint(threshold). Default is 0.
positive	If FALSE, after finding the cutpoint, the rectangleGate returned will have bounds $(-\text{Inf}, a]$ as opposed to $[a, \text{Inf})$ . This argument will be ignored if gate_tail is used in a gatingTemplate with gt_gating or via gs_add_gating_method. In those cases, pop = "-" should be used instead.
...	additional arguments passed to .deriv_density
x	a numeric vector used as input data
method	the method used to select the cutpoint. See details.
adjust	the scaling adjustment applied to the bandwidth used in the first derivative of the kernel density estimate
plot	logical specifying whether to plot the peaks found 'right' (default) or 'left'?
auto_tol	when TRUE, it tries to set the tolerance automatically.

### Details

The default behavior of the first approach, specified by method = "first\_deriv", finds valleys in the first derivative of the KDE and uses the lowest such valley to place the cutpoint on the steep right shoulder of the largest peak in the original density.

The default behavior of the second approach, specified by method = "second\_deriv", is to find peaks in the second derivative of the KDE and use the largest such peak to place the cutpoint at the point on the right shoulder of the largest peak in the original density where it is most rapidly flattening (the first derivative is rapidly growing less negative).

Both approaches can be significantly modified from defaults with a number of optional arguments. The num\_peaks argument specifies how many peaks should be found in the smoothed KDE and

ref\_peak specifies around which peak the gate's cutpoint should be placed (starting from the left-most peak). Setting the side argument to "left" modifies the procedure to put the cutpoint on the left side of the reference peak to isolate a left tail. The max and min arguments allow for pre-filtering extreme values in the channel of interest (keeping only observations with channel values less than max and/or more than min). The bandwidth used for kernel density estimation can be proportionally scaled using adjust (e.g. adjust = 0.5 will use a bandwidth that is half of the default). This allows for tuning the level of smoothing applied in the estimation.

Lastly, the tol, auto\_tol, and bias arguments allow for adjustments to be made to the cutpoint that would otherwise be returned. tol provides a tolerance value that the absolute value of the KDE derivative at the cutpoint must be under. If the derivative at the original cutpoint is greater than tol in magnitude, the returned cutpoint will be the first point to the right of the original cutpoint (or to the left in the case of side = "left") with corresponding derivative within tol. Thus in practice, a smaller value for tol effectively pushes the cutpoint further down the shoulder of the peak towards the flat tail. tol is set to 0.01 by default but setting auto\_tol = TRUE will set the tolerance to a reasonable estimate of 1% of the maximum absolute value of the first derivative of the KDE. tol and auto\_tol are only used for method = "first\_deriv". Additionally, the bias argument allows for directly shifting the returned cutpoint left or right.

It is also possible to pass additional arguments to control the calculation of the derivative, which will have some effect on the resulting cutpoint determination, but this should usually not be needed. By default the number of grid points for the derivative calculation will be 10,000, but this can be changed with num\_points. The default bandwidth can also be directly adjusted with bandwidth, where the final value used will be given by adjust\*bandwidth

By default, we compute the first derivative of the kernel density estimate. Next, we determine the lowest valley from the derivative, which corresponds to the density's mode for cytokines. We then construct a gating cutpoint as the value less than the tolerance value tol in magnitude and is also greater than the lowest valley.

Alternatively, if the method is selected as second\_deriv, we select a cutpoint from the second derivative of the KDE. Specifically, we choose the cutpoint as the largest peak of the second derivative of the KDE density which is greater than the reference peak.

## Value

a filterList containing the gates (cutpoints) for each sample with the corresponding [rectangleGate](#) objects defining the tail as the positive population.

the cutpoint along the x-axis

## Examples

```
## Not run:
gate <- gate_tail(fr, Channel = "APC-A") # fr is a flowFrame

## End(Not run)
```

---

gatingTemplate-class    *a class storing the gating method and population information in a graphNEL object*

---

**Description**

Each cell population is stored in graph node and is connected with its parent population or its reference node for boolGate or refGate.

It parses the csv file that specifies the gating scheme for a particular staining pannel.

**Usage**

```
gatingTemplate(x, ...)

## S4 method for signature 'character'
gatingTemplate(
  x,
  name = "default",
  strict = TRUE,
  strip_extra_quotes = FALSE,
  ...
)
```

**Arguments**

x	character csv file name
...	other arguments passed to <code>data.table::fread</code>
name	character the label of the gating template
strict	logical whether to perform validity check(special characters) on the alias column. By default it is(and should be) turned on for the regular template parsing. But sometime it is useful to turned it off to bypass the check for the dummy nodes(e.g. the csv template generated by 'gh_generate_template' with some existing boolean gates that has '!' or ':' symbol).
strip_extra_quotes	logical Extra quotes are added to strings by fread. This causes problems with parsing R strings to expressions in some cases. Default FALSE for usual behaviour. TRUE should be passed if parsing gating_args fails.

**Details**

This csv must have the following columns:

'alias': a name used label the cell population, the path composed by the alias and its precedent nodes (e.g. /root/A/B/alias) has to be uniquely identifiable. So alias can not contain '/' character, which is reserved as path delimiter.

'pop': population patterns of '+/-' or '+/-/-' , which tells the algorithm which side (postive or negative) of 1d gate or which quadrant of 2d gate to be kept.

'parent': the parent population alias, its path has to be uniquely identifiable.

'dims': characters seperated by comma specifying the dimensions(1d or 2d) used for gating. It can be either channel name or stained marker name (or the substrings of channel/marker names as long as they are uniquely identifiable.).

'gating\_method': the name of the gating function (e.g. 'flowClust'). It is invoked by a wrapper function that has the identical function name prefixed with a dot.(e.g. '.flowClust')

'gating\_args': the named arguments passed to gating function (Note that double quotes are often used as text delimiter by some csv editors. So try to use single quote instead if needed.)

'collapseDataForGating': When TRUE, data is collapsed (within groups if 'groupBy' specified) before gating and the gate is replicated across collapsed samples. When set FALSE (or blank), then 'groupBy' argument is only used by 'preprocessing' and ignored by gating.

'groupBy': If given, samples are split into groups by the unique combinations of study variable (i.e. column names of pData, e.g. "PTID:VISITNO"). when split is numeric, then samples are grouped by every N samples

'preprocessing\_method': the name of the preprocessing function (e.g. 'prior\_flowclust'). It is invoked by a wrapper function that has the identical function name prefixed with a dot (e.g. '.prior\_flowclust') the preprocessing results are then passed to gating wrapper function through 'pps\_res' argument.

'preprocessing\_args': the named arguments passed to preprocessing function.

## Examples

```
## Not run:
gt <- gatingTemplate(system.file("extdata/gating_template/tcell.csv", package = "openCyto"))
plot(gt)

## End(Not run)
```

---

getGate, fcTree, character-method  
*get gates saved in fcTree*

---

## Description

get gates saved in fcTree

## Usage

```
## S4 method for signature 'fcTree,character'
getGate(obj, y, ...)
```

## Arguments

obj	fcTree
y	character node name
...	other arguments (not used)

---

```
getNodes,fcTree-method
      get nodes from fcTree
```

---

**Description**

get nodes from fcTree

**Usage**

```
## S4 method for signature 'fcTree'
getNodes(x, y)
```

**Arguments**

x	fcTree
y	character node name

---

```
gh_generate_template generate a partially complete csv template from the existing gating
hierarchy
```

---

**Description**

To ease the process of replicating the existing (usually a manual one) gating schemes, this function populate an empty gating template with the 'alias', 'pop', 'parent' and 'dims' columns that exacted from an GatingHierarchy, and leave the other columns (e.g. 'gating\_method') blank. So users can make changes to that template instead of writing from scratch.

**Usage**

```
gh_generate_template(gh)
```

**Arguments**

gh	a GatingHierarchy likely parsed from a xml workspace
----	--

**Value**

a gating template in data.frame format that requires further edition after output to csv

**Examples**

```
library(flowWorkspace)
dataDir <- system.file("extdata",package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual",full = TRUE))
gh_generate_template(gs[[1]])
```

---

```
groupBy,gtMethod-method
```

*get the grouping variable for the gating method*

---

### Description

When specified, the flow data is grouped by the grouping variable (column names in pData). Within each group, when isCollapse is set to TRUE, the gating method is applied to the collapsed data. Otherwise, it is done independently for each individual sample(flowFrame). Grouping variable is also used by preprocessing method.

### Usage

```
## S4 method for signature 'gtMethod'
groupBy(object)
```

### Arguments

```
object          gtMethod
```

---

```
gs_add_gating_method  apply a gating method to the GatingSet
```

---

### Description

When interacting with the existing gated data, this function provides an alternative way to interact with the GatingSet by supplying the gating description directly through arguments without the need to write the complete csv gating template.

### Usage

```
gs_add_gating_method(
  gs,
  alias = "*",
  pop = "+",
  parent,
  dims = NA,
  gating_method,
  gating_args = NA,
  collapseDataForGating = NA,
  groupBy = NA,
  preprocessing_method = NA,
  preprocessing_args = NA,
  strip_extra_quotes = FALSE,
  ...
)
```

**Arguments**

gs                    GatingSet or GatingSetList  
 alias, pop, parent, dims, gating\_method, gating\_args, collapseDataForGating, groupBy, preprocessing\_...  
                           see details in [gatingTemplate](#)  
 strip\_extra\_quotes    logical Extra quotes are added to strings by fread. This causes problems with parsing R strings to expressions in some cases. Default FALSE for usual behaviour. TRUE should be passed if parsing gating\_args fails.  
 ...                    other arguments

- mc.cores passed to multicore package for parallel computing
- parallel\_type character specifying the parallel type. The valid options are "none", "multicore", "cluster".
- cl cluster object passed to parallel package (when parallel\_type is "cluster")

**Details**

Calls to `gs_add_gating_method` can also be easily reversed with `gs_remove_gating_method`. Note, however, that it is not possible to differentiate between different `GatingSet` objects loaded from the same directory with `load_gs` within a session. Thus, to guarantee a clean history for `gs_remove_gating_method`, it is necessary to call `gs_add_gating_method_init` on the loaded `GatingSet` immediately after re-loading it. See the documentation for `gs_add_gating_method_init` for more details. This will not be an issue for `GatingSet` objects created directly using the constructor.

**See Also**

[gs\\_remove\\_gating\\_method](#) [gs\\_add\\_gating\\_method\\_init](#)

**Examples**

```
## Not run:
# add quad gates
gs_add_gating_method(gs, gating_method = "mindensity", dims = "CCR7,CD45RA", parent = "cd4-cd8+", pop = "CCR7+,
# polyfunctional gates (boolean combinations of existing marginal gates)
gs_add_gating_method(gs, gating_method = "polyFunctions", parent = "cd8", gating_args = "cd8/IFNg:cd8/IL2:cd8/
#boolGate method
gs_add_gating_method(gs, alias = "IL2orIFNg", gating_method = "boolGate", parent = "cd4", gating_args = "cd4/IL
## End(Not run)
```

---

`gs_add_gating_method_init`

*Clear history of `gs_add_gating_method` calls for a given `GatingSet` or `GatingSetList`*

---





**Usage**

```
gs_remove_gating_method(gs)
```

**Arguments**

**gs** The GatingSet or GatingSetList for which the most recent gs\_add\_gating\_method call should be reversed.

**See Also**

[gs\\_add\\_gating\\_method](#) [gs\\_add\\_gating\\_method\\_init](#)

**Examples**

```
## Not run:
# add quad gates
gs_add_gating_method(gs, gating_method = "mindensity", dims = "CCR7,CD45RA", parent = "cd4-cd8+", pop = "CCR7+
# Remove the gates and nodes resulting from that gs_add_gating_method call
gs_remove_gating_method(gs)

## End(Not run)
```

---

gtMethod-class

*A class to represent a gating method.*

---

**Description**

A gating method object contains the specifics for generating the gates.

**Slots**

**name** a character specifying the name of the gating method

**dims** a character vector specifying the dimensions (channels or markers) of the gate

**args** a list specifying the arguments passed to gating function

**groupBy** a character or integer specifying how to group the data. If character, group the data by the study variables (columns in pData). If integer, group the data by every N samples.

**collapse** a logical specifying whether to collapse the data within group before gating. it is only valid when groupBy is specified

**Examples**

```
## Not run:
gt <- gatingTemplate(system.file("extdata/gating_template/tcell.csv", package = "openCyto"))
gh_pop_get_gate(gt, '2', '3')

## End(Not run)
```

---

gtPopulation-class	<i>A class to represent a cell population that will be generated by a gating method.</i>
--------------------	--

---

### Description

A class to represent a cell population that will be generated by a gating method.

### Slots

**id** numeric unique ID that is consistent with node label of graphNEL in gating template

**name** character the name of population

**alias** character the more user friendly name of population

### Examples

```
## Not run:
gt <- gatingTemplate(system.file("extdata/gating_template/tcell.csv", package = "openCyto"))

gt_get_nodes(gt, '2')

## End(Not run)
```

---

gtSubsets-class	<i>A class representing a group of cell populations.</i>
-----------------	--

---

### Description

It extends gtPopulation class.

---

gt_gating	<i>Applies a gatingTemplate to a GatingSet.</i>
-----------	---

---

### Description

It loads the gating methods by topological order and applies them to GatingSet.

### Usage

```
gt_gating(x, y, ...)
```

**Arguments**

- |     |  |
|-----|--|
| x   | a gatingTemplate object  |
| y   | a GatingSet object   |
| ... | <ul style="list-style-type: none"> <li>• start a character that specifies the population (corresponding to 'alias' column in csv template) where the gating process will start from. It is useful to quickly skip some gates and go directly to the target population in the testing run. Default is "root".</li> <li>• stop.at a character that specifies the population (corresponding to 'alias' column in csv template) where the gating process will stop at. Default is NULL, indicating the end of gating tree.</li> <li>• keep.helperGates a logical flag indicating whether to keep the intermediate helper gates that are automatically generated by openCyto. Default is TRUE.</li> <li>• mc.cores passed to multicore package for parallel computing</li> <li>• parallel_type character specifying the parallel type. The valid options are "none", "multicore", "cluster".</li> <li>• cl cluster object passed to parallel package (when parallel_type is "cluster")</li> </ul> |

**Value**

Nothing. As the side effect, gates generated by gating methods are saved in GatingSet.

**Examples**

```
## Not run:
gt <- gatingTemplate(file.path(path, "data/ICStemplate.csv"), "ICS")
gs <- GatingSet(fs) #fs is a flowSet/ncdfFlowSet
gt_gating(gt, gs)
gt_gating(gt, gs, stop.at = "v") #proceed the gating until population 'v'
gt_gating(gt, gs, start = "v") # start from 'v'
gt_gating(gt, gs, parallel_type = "multicore", mc.cores = 8) #parallel gating using multicore
#parallel gating by using cluster
cl1 <- makeCluster (8, type = "MPI")
gt_gating(gt, gs, parallel_type = "cluster", cl = cl1)
stopCluster ( cl1 )

## End(Not run)
```

---

gt\_get\_children

*get children nodes*

---

**Description**

get children nodes

**Usage**

```
gt_get_children(obj, y)
```

**Arguments**

obj	gatingTemplate
y	character parent node path

**Examples**

```
## Not run:
gt <- gatingTemplate(system.file("extdata/gating_template/tcell.csv", package = "openCyto"))

gt_get_nodes(gt, "/nonDebris")
gt_get_children(gt, "/nonDebris")

## End(Not run)
```

---

gt_get_gate	<i>get gating method from the node</i>
-------------	--

---

**Description**

get gating method from the node

**Usage**

```
gt_get_gate(obj, y, z)
```

**Arguments**

obj	gatingTemplate
y	character parent node path
z	character child node path

**Examples**

```
## Not run:
gt <- gatingTemplate(system.file("extdata/gating_template/tcell.csv", package = "openCyto"))
gt_get_nodes(gt, only.names = TRUE)
gt_get_nodes(gt, "/nonDebris")
gt_get_children(gt, "/nonDebris")
gt_get_gate(gt, "/nonDebris", "/nonDebris/singlets")

## End(Not run)
```

---

gt_get_nodes	<i>get nodes from <a href="#">gatingTemplate</a> object</i>
--------------	---

---

**Description**

get nodes from [gatingTemplate](#) object

**Usage**

```
gt_get_nodes(  
  x,  
  y,  
  order = c("default", "bfs", "dfs", "tsort"),  
  only.names = FALSE  
)
```

**Arguments**

x	<a href="#">gatingTemplate</a>
y	character node index. When missing, return all the nodes
order	character specifying the order of nodes. options are "default", "bfs", "dfs", "tsort"
only.names	logical specifying whether user wants to get the entire <a href="#">gtPopulation</a> object or just the name of the population node

**Examples**

```
## Not run:  
gt <- gatingTemplate(system.file("extdata/gating_template/tcell.csv", package = "openCyto"))  
gt_get_nodes(gt)[1:2]  
gt_get_nodes(gt, only.names = TRUE)  
gt_get_nodes(gt, "/nonDebris")  
  
## End(Not run)
```

---

gt_get_parent	<i>get parent nodes</i>
---------------	-------------------------

---

**Description**

get parent nodes

**Usage**

```
gt_get_parent(obj, y, isRef = FALSE)
```

**Arguments**

obj	gatingTemplate
y	character child node path
isRef	logical whether show the reference node besides the parent node

**Examples**

```
## Not run:
gt <- gatingTemplate(system.file("extdata/gating_template/tcell.csv", package = "openCyto"))

gt_get_nodes(gt, "/nonDebris")
gt_get_parent(gt, "/nonDebris/singlets")

## End(Not run)
```

---

gt_list_methods	<i>Print a list of the registered gating methods</i>
-----------------	--

---

**Description**

Print a list of the registered gating methods

**Usage**

```
gt_list_methods()
```

**Value**

Does not return anything. Prints a list of the available gating methods.

---

gt_toggle_helpergates	<i>toggle/delete the hidden flag of the helper gates</i>
-----------------------	--

---

**Description**

The helper gates are defined as the referred gates in csv template. And all the children of referred gates are also referred gates thus they are considered the helper gates and can usually be hidden to simply the final gating tree.

**Usage**

```
gt_toggle_helpergates(gt, gs)
```

```
gt_get_helpergates(gt, gs)
```

```
gt_delete_helpergates(gt, gs)
```

**Arguments**

gt	gatingTemplate object
gs	GatingSet

**Details**

Note that delete action is NOT reversible.

**Examples**

```
## Not run:
gt <- gatingTemplate(gtFile)
#run the gating
gt_gating(gt, gs)
#hide the gates that are not of interest
gt_toggle_helper_gates(gt, gs)
#or simply remove them if you are sure they will not be useful in future
gt_delete_helper_gates(gt, gs)

## End(Not run)
```

---

isCollapse.gtMethod-method

*get the flag that determines whether gating method is applied on collapsed data*

---

**Description**

When TRUE, the flow data(multiple flowFrames) is collapsed into one and the gating method is applied on the collapsed data. Once the gate is generated, it is then replicated and applied to the each single flowFrame.

**Usage**

```
## S4 method for signature 'gtMethod'
isCollapse(object)
```

**Arguments**

object	gtMethod
--------	----------

**Value**

logical

---

names,gtMethod-method *get gating method name*

---

### Description

get gating method name

### Usage

```
## S4 method for signature 'gtMethod'
names(x)
```

### Arguments

x                   gtMethod

### Examples

```
## Not run:
gt <- gatingTemplate(system.file("extdata/gating_template/tcell.csv", package = "openCyto"))

gtMthd <- gt_get_gate(gt, "/nonDebris/singlets", "/nonDebris/singlets/lymph")
names(gtMthd)
dims(gtMthd)
parameters(gtMthd)
isCollapse(gtMthd)
groupBy(gtMthd)

gtPop <- gt_get_nodes(gt, "/nonDebris/singlets/lymph/cd3/cd4+cd8-/CD38+")
names(gtPop)
alias(gtPop)

## End(Not run)
```

---

names,gtPopulation-method  
*get population name*

---

### Description

get population name

### Usage

```
## S4 method for signature 'gtPopulation'
names(x)
```

### Arguments

x                   gtPopulation object



---

ocRectangleGate-class *the class that carries event indices as well*

---

**Description**

the class that carries event indices as well

---

ocRectRefGate *constructor for ocRectRefGate*

---

**Description**

constructor for ocRectRefGate

**Usage**

ocRectRefGate(rectGate, boolExprs)

**Arguments**

rectGate	rectangleGate
boolExprs	character boolean expression of reference nodes

---

ocRectRefGate-class *special gate type that mix the rectangleGate with boolean gate*

---

**Description**

special gate type that mix the rectangleGate with boolean gate

---

openCyto *Hierarchical Gating Pipeline for flow cytometry data*

---

**Description**

Hierarchical Gating Pipeline for flow cytometry data.

**Details**

openCyto is a package designed to facilitate the automated gating methods in sequential way to mimic the manual gating strategy.

Package:	openCyto
Type:	Package
Version:	1.2.8
Date:	2014-04-10
License:	GPL (>= 2)
LazyLoad:	yes

**Author(s)**

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Maintainer: Mike Jiang <wjiang2@fhcrc.org>

**See Also**

See [gt\\_gating](#), [gate\\_flowclust\\_1d](#), for an overview of gating functions.

**Examples**

```
## Not run: gatingTemplate('test.csv')
```

---

openCyto-deprecated     *Deprecated functions in package openCyto.*

---

**Description**

`add_pop` → [gs\\_add\\_gating\\_method](#)  
`add_pop_init` → [gs\\_add\\_gating\\_method\\_init](#)  
`prior_flowClust` → [prior\\_flowclust](#)  
`templateGen` → [gh\\_generate\\_template](#)  
`gate_flowClust_1d` → [gate\\_flowclust\\_1d](#)  
`gate_flowClust_2d` → [gate\\_flowclust\\_2d](#)  
`quantileGate` → [gate\\_quantile](#)  
`cytokine` → [gate\\_tail](#)  
`quadGate.seq` → [gate\\_quad\\_sequential](#)  
`quadGate.tmix` → [gate\\_quad\\_tmix](#)  
`gating` → [gt\\_gating](#)  
`getNode` → [gt\\_get\\_nodes](#)  
`getChildren` → [gt\\_get\\_children](#)  
`getParent` → [gt\\_get\\_parent](#)  
`getGate` → [gt\\_get\\_gate](#)  
`listgtMethods` → [gt\\_list\\_methods](#)  
`registerPlugins` → [register\\_plugins](#)  
`remove_pop` → [gs\\_remove\\_gating\\_method](#)  
`tautString` → [tautstring](#)  
`tautStringGate` → [gate\\_tautstring](#)  
`toggle.helperGates` → [gt\\_toggle\\_helper gates](#)  
`get.helperGates` → [gt\\_get\\_helper gates](#)  
`delete.helperGates` → [gt\\_delete\\_helper gates](#)

---

openCyto.options	<i>Some global options for openCyto See examples for the meaning of these options and how to get/set them.</i>
------------------	--

---

### Description

Get/set some global options for openCyto

### Examples

```
opt <- getOption("openCyto")
#the threshold of minimum cell events required for the gating algorithm to proceed
opt[["gating"]][["minEvents"]]
#to change the threshold
opt[["gating"]][["minEvents"]] <- 100
options(openCyto = opt)

#switch off the validity check flags(Not recommended)
opt[["check.pop"]] <- FALSE
options(openCyto = opt)
```

---

parameters,gtMethod-method	<i>get parameters of the gating method/function</i>
----------------------------	---

---

### Description

get parameters of the gating method/function

### Usage

```
## S4 method for signature 'gtMethod'
parameters(object)
```

### Arguments

object	gtMethod
--------	----------

---

```
plot,fcFilterList,ANY-method
      plot a fcFilterList
```

---

### Description

It is usually called by plot method for fcTree instead of directly by users.

### Usage

```
## S4 method for signature 'fcFilterList,ANY'
plot(
  x,
  y,
  samples = NULL,
  posteriors = FALSE,
  xlim = NULL,
  ylim = NULL,
  node = NULL,
  data = NULL,
  breaks = 20,
  lwd = 1,
  ...
)
```

### Arguments

x	fcFilterList
y	character channel name
samples	character a vector of sample names to be plotted
posteriors	logical indicating whether posteriors should be plotted
xlim, ylim	scale settings for x,y axes
node	character population name associated with the fcFilterList
data	GatingSet object
breaks	passed to <a href="#">hist</a>
lwd	line width
...	other arguments passed to base plot

### Examples

```
## Not run:
env1<-new.env(parent=emptyenv())
#gt is a gatingTemplate, gs is a GatingSet
gt_gating(gt,gs,env1) #the flowClust gating results are stored in env1
plot(env1$fcfct,"nonDebris",post=T) #plot the priors as well as posteriors for the "nonDebris" gate

## End(Not run)
```

---

 plot,fcTree,character-method

*plot the flowClust gating results*


---

### Description

This provides the priors and posteriors as well as the gates for the purpose of debugging flowClust gating algorithm

### Usage

```
## S4 method for signature 'fcTree,character'
plot(x, y, channel = NULL, data = NULL, ...)
```

### Arguments

x	fcTree
y	character node name in the fcTree
channel	character specifying the channel.
data	GatingSet that the fcTree is associated with
...	other arguments

---

 plot,gatingTemplate,missing-method

*plot the gating scheme*


---

### Description

plot the gating scheme using Rgraphviz

### Usage

```
## S4 method for signature 'gatingTemplate,missing'
plot(x, y, ...)
```

### Arguments

x	gatingTemplate object
y	either character specifying the root node which can be used to visualize only the subgraph or missing which display the entire gating scheme
...	other arguments

graphAttr, nodeAttr: graph rendering attributes passed to [renderGraph](#) showRef logical: whether to display the reference gates. Sometime it maybe helpful to hide all those reference gates which are not the cell population of interest and used primarily for generating other population nodes.

**Examples**

```
## Not run:
gt <- gatingTemplate(system.file("extdata/gating_template/tcell.csv", package = "openCyto"))
plot(gt) #plot entire tree
plot(gt, "lymph") #only plot the subtree rooted from "lymph"

## End(Not run)
```

---

polyFunctions-class    *A class to represent a polyFunctions gating method.*

---

**Description**

It extends boolMethod class and will be expanded to multiple boolMethod object.

---

pop\_add.ocRectangleGate  
*bypass the default flowWorkspace:::addGate*

---

**Description**

to support adding gate along with indices without loading flow data and computing  
to support adding rectangleGate yet gating through boolean operations without loading flow data

**Usage**

```
## S3 method for class 'ocRectangleGate'
pop_add(gate, gh, recompute, ...)

## S3 method for class 'ocRectRefGate'
pop_add(gate, gh, recompute, ...)
```

**Arguments**

gate	ocRectangleGate or logicalFilterResult
gh	GatingHierarchy see <a href="#">add</a> in flowWorkspace package
recompute	logical see <a href="#">add</a> in flowWorkspace package
...	see <a href="#">add</a> in flowWorkspace package

**Details**

however it is proven that logical indices are too big to be efficiently passed around

---

posterior,fcFilter,ANY-method  
*get posterior from a fcFilter object*

---

**Description**

get posterior from a fcFilter object

**Usage**

```
## S4 method for signature 'fcFilter,ANY'  
posterior(x, y = "missing")
```

**Arguments**

x	fcFilter
y	character or missing that specify which channel to look for

---

ppMethod,gatingTemplate,character-method  
*get preprocessing method from the node*

---

**Description**

get preprocessing method from the node

**Usage**

```
## S4 method for signature 'gatingTemplate,character'  
ppMethod(obj, y, z)
```

**Arguments**

obj	gatingTemplate
y	character parent node path
z	character child node path

**Examples**

```
## Not run:  
gt <- gatingTemplate(system.file("extdata/gating_template/tcell.csv", package = "openCyto"))  
ppMethod(gt, "/nonDebris/singlets", "/nonDebris/singlets/lymph")  
  
## End(Not run)
```

---

ppMethod-class            *A class to represent a preprocessing method.*

---

### Description

It extends gtMethod class.

### Examples

```
## Not run:
gt <- gatingTemplate(system.file("extdata/gating_template/tcell.csv", package = "openCyto"))
ppMethod(gt, '3', '4')

## End(Not run)
```

---

preprocessing,ppMethod,GatingSet-method  
*apply a [ppMethod](#) to the GatingSet*

---

### Description

apply a [ppMethod](#) to the GatingSet

### Usage

```
## S4 method for signature 'ppMethod,GatingSet'
preprocessing(x, y, ...)
```

### Arguments

x	ppMethod
y	GatingSet or GatingSetList
...	other arguments

---

priors,fcFilter,ANY-method  
*get priors from a fcFilter object*

---

### Description

get priors from a fcFilter object

### Usage

```
## S4 method for signature 'fcFilter,ANY'
priors(x, y = "missing")
```



**Arguments**

x	fcFilter object
y	character specifying channel name. if missing then extract priors for all the channels

---

prior_flowclust	<i>Elicits data-driven priors from a flowSet object for specified channels</i>
-----------------	--

---

**Description**

We elicit data-driven prior parameters from a flowSet object for specified channels. For each sample in the flowSet object, we apply the given prior\_method to elicit the priors parameters.

**Usage**

```
prior_flowclust(
  flow_set,
  channels,
  prior_method = c("kmeans"),
  K = 2,
  nu0 = 4,
  w0 = c(10, 10),
  shrink = 0.000001,
  ...
)
```

**Arguments**

flow_set	a flowSet object
channels	a character vector containing the channels in the flowSet from which we elicit the prior parameters for the Student's t mixture
prior_method	the method to elicit the prior parameters
K	the number of mixture components to identify
nu0	prior degrees of freedom of the Student's t mixture components.
w0	the number of prior pseudocounts of the Student's t mixture components. (only the first element is used and the rest is ignored at the moment)
shrink	the amount of eigenvalue shrinkage to add in the case the prior covariance matrices are singular. See details.
...	Additional arguments passed to the prior elicitation method selected

**Details**

Currently, we have implemented only two methods. In the case that one channel is given, we use the kernel-density estimator (KDE) approach for each sample to obtain K peaks from which we elicit prior parameters. Otherwise, if more than one channel is specified, we apply K-Means to each of the samples in the flowSet and aggregate the clusters to elicit the prior parameters.

In the rare case that a prior covariance matrix is singular, we shrink the eigenvalues of the matrix slightly to ensure that it is positive definite. For instance, if the flow\_set has two samples, this case can occur. The amount of shrinkage is controlled in shrink.

**Value**

list of the necessary prior parameters

---

refGate-class	<i>A class to represent a reference gating method.</i>
---------------	--

---

**Description**

It extends gtMethod class.

**Slots**

**refNodes** character specifying the reference nodes

---

register_plugins	<i>Register a gating or preprocessing function with OpenCyto</i>
------------------	--

---

**Description**

Function registers a new gating or preprocessing method with openCyto so that it may be used in the csv template.

**Usage**

```
register_plugins(fun = NA, methodName, dep = NA, ...)
```

**Arguments**

fun	function to be registered
methodName	character name of the gating or preprocessing method
dep	character name of the library dependency required for the plugin method to work.
...	other arguments type character specifying the type of registering method. Should be either "gating" or "preprocessing".

**Details**

The fun argument should be a wrapper function definition for the gating or preprocessing method. Gating method must have formal arguments:

fr a flowFrame

pp\_res a pre-processing result

xChannel character (optional)

yChannel character (required)

filterId character

... ellipses for the additional parameters.

Preprocessing method must have formal arguments:

fs a flowSet that stores the flow data (could be subgrouped data if groupBy column is defined in the csv template)

gs a GatingSet

gm a gtMethod object that stores the information from gating method

xChannel character (required)

yChannel character (required)

... ellipses for the additional parameters.

The gating function must return a filter (i.e. polygonGate or other instance) from flowCore. The preprocessing can return anything and it will be passed on to the gating function. So it is up to gating function to use and interpret the results of preprocessing. Not all formal parameters need to be used. Additional arguments are passed via the ... and can be processed in the wrapper

### Value

logical TRUE if successful and prints a message. FALSE otherwise.

---

show, boolMethod-method

*show method for boolMethod*

---

### Description

show method for boolMethod

### Usage

```
## S4 method for signature 'boolMethod'
show(object)
```

### Arguments

object            boolMethod

---

show, fcFilter-method    *show method for fcFilter*

---

### Description

show method for fcFilter

### Usage

```
## S4 method for signature 'fcFilter'
show(object)
```

### Arguments

object            fcFilter show method for fcFilter

---

show,gatingTemplate-method  
*show method for gatingTemplate*

---

### Description

show method for gatingTemplate

### Usage

```
## S4 method for signature 'gatingTemplate'
show(object)
```

### Arguments

object           gatingTemplate

---

show,gtMethod-method   *show method for gtMethod*

---

### Description

show method for gtMethod

### Usage

```
## S4 method for signature 'gtMethod'
show(object)
```

### Arguments

object           gtMethod show method for gtMethod

---

tautstring           *Taut String Density Estimator Gating*

---

### Description

The taut string density estimator gating returns 0, 1, or more gates, depending on how many modes it identifies in the data.

**Usage**

```
tautstring(sorted_vector, modeprior = 0)
```

```
gate_tautstring(  
  fr,  
  channel,  
  gate_range = NULL,  
  min = NULL,  
  max = NULL,  
  filterId = ""  
)
```

**Arguments**

sorted_vector	numeric vector of single cell expression from a single cytometric channel.
modeprior	numeric scalar specifying the expected number of modes. Default 0 (autodetect). Rarely should this be set by the user.
fr	a flowFrame object
channel	The channel to gate.
gate_range	The range to look for a gate, no truncation occurs.
min	The min range of the data to truncate the flowFrame
max	The max range of the data to truncate the flowFrame
filterId	The id / name of the gate.

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