# Package 'msdata'

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Title Various Mass Spectrometry raw data example files

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Suggests xcms, mzR

ZipData no

Description Ion Trap positive ionization mode data in mzData file format. Subset from 500-850 m/z and 1190-1310 seconds, incl. MS2 and MS3, intensity threshold 100.000. Extracts from FTICR Apex III, m/z 400-450. Subset of UPLC - Bruker micrOTOFq data, both mzData, mzML and mz5. LC-MSMS and MRM files from proteomics experiments. PSI mzIdentML example files for various search engines.

biocViews ExperimentData, MassSpectrometryData

**License** GPL ( $\geq 2$ )

NeedsCompilation no

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msdata

Sample FTICR, LC/MS and MS\$^n\$ data

#### Description

x object containing a subset of LC/MS raw data from a Thermo Finnigan LCQ Deca XP The data is a subset from 500-850 m/z and 1190-1310 seconds, incl. MS2 and MS3, intensity threshold 100.000. It was collected in positive ionization mode.

xs object containing a subset of FTICR data from a Bruker APex III FTICR. The data is a subset from 400-450 m/z, collected in positive ionization mode.

#### Usage

data(xs)

## Format

The format is:

xs

# Details

The corresponding raw mzdata files are located in the fticr and iontrap subdirectory of this package.

#### See Also

xcmsSet, xcmsRaw

## Examples

```
## The directory with the mzData LC/MS files
data(xs)
mzdatapath <- file.path(find.package("msdata"), "iontrap")
mzdatapath
files <- list.files(mzdatapath, recursive = TRUE, full.names = TRUE)
files
if (require(xcms)) {
## xcmsSet Summary
show(xs)
## Access raw data file
x <- xcmsRaw(files[1])
x
}
```

proteomics

# Proteomics data in msdata

#### Description

This function returns proteomics mass spectrometry files. These files are all stored in the proteomics directory in the msdata package. Each file/data is described in more details below.

#### Usage

proteomics(...)

#### Arguments

. . .

Additional arguments passed to list.files.

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#### proteomics

#### Details

• TMT\_Erwinia\_1uLSike\_Top10HCD\_isol2\_45stepped\_60min\_01.mzML.gz: A LC-MSMS data file containing iTRAQ 4-plex data. The data is described in more details in Gatto L. and Christoforou A. *Using R and Bioconductor for proteomics data analysis* (PMID 23692960). This file only contains a subset of the fill data (spectra 1002 to 1510) and was generated from the full data using msconvert (ProteoWizard release: 3.0.9283 (2016-1-11)) using following command

msconvert TMT\_Erwinia\_1uLSike\_Top10HCD\_isol2\_45stepped\_60min\_01-20141210.mzML
--filter "index [1002,1510]" -o subset

The original file is available in the ProteomeXchange PXD000001 project.

An MS2 identification file, ident/TMT\_Erwinia\_1uLSike\_Top10HCD\_isol2\_45stepped\_60min\_01-20141210.m generated searching the raw data against the *Erwinia carotovora* database (see reference above) is also available through the ident function.

- MS3TMT10\_01022016\_32917-33481.mzML.gz:A subset of 565 spectra from a currenly unpublished TMT 10-plex experiment run on an Thermo Orbitrap Lumos with synchronous precursor selection (SPS) MS3. Only the MS2 spectra were centroided during convertion using msconvert (ProteoWizard release: 3.0.9283 (2016-1-11)) using vendor libraries.
- MRM-standmix-5.mzML.gz:Sample from mouse brain acquired by HILIC ESI-QqQ/MS in Dynamic multiple reaction monitoring mode (MRM). HPLC system was a 1290 Infinity (Agilent Technologies) coupled to ion-Funnel Triple quadrupole 6490 mass spectrometer (Agilent Technologies). This file was contributed by Xavi Domingo-Almenara from theThe Scripps Research Institute, San Diego, CA.

#### Value

A character with file names.

#### Author(s)

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#### See Also

For more access to mass spectrometry-based proteomics data, see the rpx and ProteomicsAnnotationHubData packages.

#### Examples

```
(f <- proteomics(full.names = TRUE))</pre>
```

library(mzR)
openMSfile(f[2])

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