

In-silico cleavage of polypeptides using the **cleaver** package

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1 Introduction

Most proteomics experiments need protein (peptide) separation and cleavage procedures before these molecules could be analyzed or identified by mass spectrometry or other analytical tools.

cleaver allows in-silico cleavage of polypeptide sequences to e.g. create theoretical mass spectrometry data.

The cleavage rules are taken from the [ExPASy PeptideCutter tool](#)[?].

2 Simple Usage

Loading the **cleaver** package:

```
> library("cleaver")
```

Getting help and list all available cleavage rules:

```
> help("cleave")
```

Cleaving of *Gastric juice peptide 1 (P01358)* using *Trypsin*:

```
> ## cleave it
> cleave("LAAGKVEDSD", enzym="trypsin")

$LAAGKVEDSD
[1] "LAAGK" "VEDSD"

> ## get the cleavage ranges
> cleavageRanges("LAAGKVEDSD", enzym="trypsin")
```

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```
$LAAGKVEDSD
      start end
[1,]      1   5
[2,]      6  10

> ## get only cleavage sites
> cleavageSites("LAAGKVEDSD", enzym="trypsin")

$LAAGKVEDSD
[1] 5
```

Sometimes cleavage is not perfect and the enzyme miss some cleavage positions:

```
> ## miss one cleavage position
> cleave("LAAGKVEDSD", enzym="trypsin", missedCleavages=1)

$LAAGKVEDSD
[1] "LAAGKVEDSD"

> cleavageRanges("LAAGKVEDSD", enzym="trypsin", missedCleavages=1)

$LAAGKVEDSD
      start end
[1,]      1  10

> ## miss zero or one cleavage positions
> cleave("LAAGKVEDSD", enzym="trypsin", missedCleavages=0:1)

$LAAGKVEDSD
[1] "LAAGK"      "VEDSD"      "LAAGKVEDSD"

> cleavageRanges("LAAGKVEDSD", enzym="trypsin", missedCleavages=0:1)

$LAAGKVEDSD
      start end
[1,]      1   5
[2,]      6  10
[3,]      1  10
```

Combine *cleaver* and the *Biostrings* R package⁷ :

```
> ## create AAStringSet object
> p <- AAStringSet(c(gaju="LAAGKVEDSD", pnm="AGEPKLDAGV"))
>
> ## cleave it
> cleave(p, enzym="trypsin")

AAStringSetList of length 2
[["gaju"]] LAAGK VEDSD
[["pnm"]] AGEPK LDAGV

> cleavageRanges(p, enzym="trypsin")

IRangesList of length 2
$gaju
```

```
IRanges object with 2 ranges and 0 metadata columns:
```

	start	end	width
	<integer>	<integer>	<integer>
[1]	1	5	5
[2]	6	10	5

```
$pnm
```

```
IRanges object with 2 ranges and 0 metadata columns:
```

	start	end	width
	<integer>	<integer>	<integer>
[1]	1	5	5
[2]	6	10	5

```
> cleavageSites(p, enzym="trypsin")
```

```
$gaju
```

```
[1] 5
```

```
$pnm
```

```
[1] 5
```

3 Insulin & Somatostatin Example

Downloading *Insulin* (P01308) and *Somatostatin* (P61278) sequences from the [UniProt](#)[?] database using the *UniProt.ws* R package[?].

```
> ## load UniProt.ws library
> library("UniProt.ws")
>
> ## select species Homo sapiens
> UniProt.ws <- UniProt.ws(taxId=9606)
>
> ## download sequences of Insulin/Somatostatin
> s <- select(UniProt.ws, keys=c("P01308", "P61278"), columns=c("SEQUENCE"))
```

Getting extra data for P01308,P61278

'select()' returned 1:1 mapping between keys and columns

```
> ## fetch only sequences
> sequences <- setNames(s$SEQUENCE, s$UNIPROTKB)
>
> ## remove whitespaces
> sequences <- gsub(pattern="[:space:]", replacement="", x=sequences)
```

Cleaving using *Pepsin*:

```
> cleave(sequences, enzym="pepsin")
```

```
$P01308
```

```

[1] "MA"      "L"      "WMRLLP"  "LL"
[5] "A"       "WGPDPAAA" "F"       "VNQH"
[9] "CGSH"    "VEA"     "Y"       "VCGERG"
[13] "FF"      "YTPKTRREAED" "QVGQVE"  "GGGPGAGS"
[17] "LQP"     "LA"      "EGS"     "QKRGIVEQCCTSICS"
[21] "YQ"      "ENYCN"

$P61278
[1] "ML"      "SCRL"    "QCA"
[4] "L"       "AA"      "SIV"
[7] "A"       "GCVTGAPSDPRL" "RQ"
[10] "FL"      "QKS"     "LAAAAGKQEL"
[13] "AKY"     "AE"      "SEPNQTENDA"
[16] "LEPED"   "SQAAEQDEMRL" "EL"
[19] "QRSANSNPAMAPRERKAGCKN" "FF"      "WKT"
[22] "FTSC"

```

4 Isotopic Distribution Of Tryptic Digested Insulin

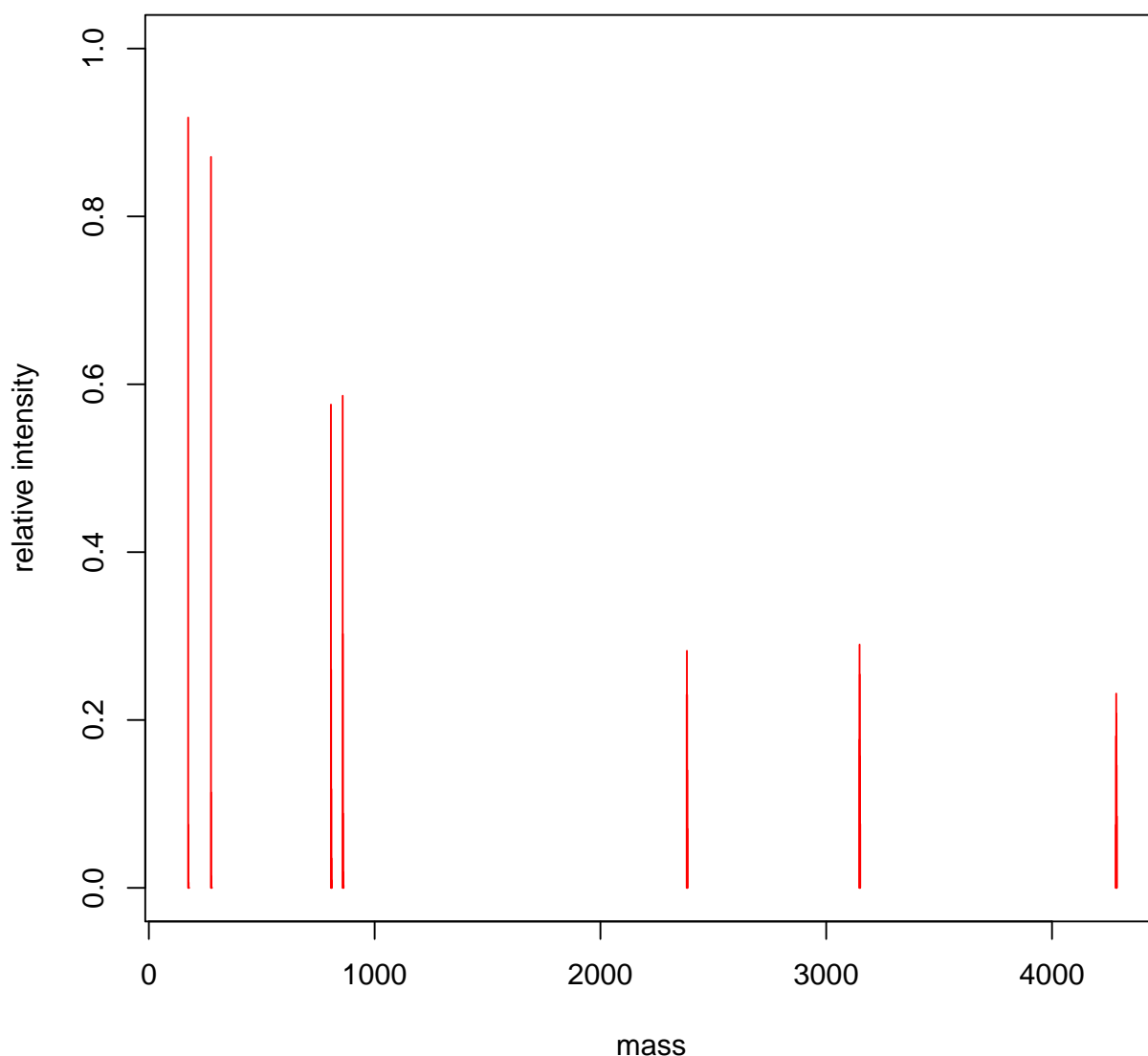
A common use case of in-silico cleavage is the calculation of the isotopic distribution of peptides (which were enzymatic digested in the in-vitro experimental workflow). Here the *BRAIN* R package^{??} is used to calculate the isotopic distribution of *cleaver*'s output. (please note: it is only a toy example, e.g. the relation of intensity values between peptides isn't correct).

```

> ## load BRAIN library
> library("BRAIN")
>
> ## cleave insulin
> cleavedInsulin <- cleave(sequences[1], enzym="trypsin")[[1]]
>
> ## create empty plot area
> plot(NA, xlim=c(150, 4300), ylim=c(0, 1),
+      xlab="mass", ylab="relative intensity",
+      main="tryptic digested insulin - isotopic distribution")
>
> ## loop through peptides
> for (i in seq(along=cleavedInsulin)) {
+   ## count C, H, N, O, S atoms in current peptide
+   atoms <- BRAIN::getAtomsFromSeq(cleavedInsulin[[i]])
+   ## calculate isotopic distribution
+   d <- useBRAIN(atoms)
+   ## draw peaks
+   lines(d$masses, d$isoDistr, type="h", col=2)
+ }

```

tryptic digested insulin – isotopic distribution



5 Session Information

- R version 3.3.1 (2016-06-21), x86_64-apple-darwin13.4.0
- Locale: C/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
- Base packages: base, datasets, grDevices, graphics, methods, parallel, stats, stats4, utils
- Other packages: BRAIN 1.20.0, BiocGenerics 0.20.0, Biostrings 2.42.0, DBI 0.5-1, IRanges 2.8.0, PolynomF 0.94, RCurl 1.95-4.8, RSQLite 1.0.0, S4Vectors 0.12.0, UniProt.ws 2.14.0, XVector 0.14.0, bitops 1.0-6, cleaver 1.12.0, knitr 1.14, lattice 0.20-34
- Loaded via a namespace (and not attached): AnnotationDbi 1.36.0, Biobase 2.34.0, BiocStyle 2.2.0, evaluate 0.10, formatR 1.4, grid 3.3.1, highr 0.6, magrittr 1.5, stringi 1.1.2, stringr 1.1.0, tools 3.3.1, zlibbioc 1.20.0