# Package 'ACME'

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|---|
| Title Algorithms for Calculating Microarray Enrichment (ACME)   |
| Author Sean Davis <sdavis2@mail.nih.gov></sdavis2@mail.nih.gov>   |
| Maintainer Sean Davis <sdavis2@mail.nih.gov></sdavis2@mail.nih.gov>   |
| <b>Depends</b> R (>= 2.10), Biobase (>= 2.5.5), methods, BiocGenerics   |
| Imports graphics, stats   |
| Description ACME (Algorithms for Calculating Microarray Enrichment) is a set of tools for analysing tiling array ChIP/chip, DNAse hypersensitivity, or other experiments that result in regions of the genome showing ``enrichment". It does not rely on a specific array technology (although the array should be a ``tiling" array), is very general (can be applied in experiments resulting in regions of enrichment), and is very insensitive to array noise or normalization methods. It is also very fast and can be applied on whole-genome tiling array experiments quite easily with enough memory. |
| License GPL (>= 2)  |
| <pre>URL http://watson.nci.nih.gov/~sdavis</pre>  |
| biocViews Technology, Microarray, Normalization   |
| NeedsCompilation yes  |
| R topics documented:  |
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#### **Description**

A subclass of ACMESet that can also store the parameters and results of an ACME calculation

## **Objects from the Class**

Objects can be created by calls of the form new("ACMECalcSet", assayData, phenoData, featureData, experimentData and addition to the constraints defined by ACMESet, this class can also hold the results (in the assay-DataElement vals) and the threshold and cutpoints from an ACME do.aGFF.calc run

#### **Slots**

```
cutpoints: Object of class "numeric" The values of the cutpoints used in an analysis by do.aGFF.calc,
    one per sample.
threshold: Object of class "numeric" The threshold used in an analysis.
assayData: Object of class "AssayData". See ExpressionSet for details.
phenoData: Object of class "AnnotatedDataFrame" See ExpressionSet for details.
featureData: Object of class "AnnotatedDataFrame" See ExpressionSet for details.
experimentData: Object of class "MIAME" See ExpressionSet for details.
annotation: Object of class "character" See ExpressionSet for details.
.__classVersion__: Object of class "Versions" See ExpressionSet for details.
```

### **Extends**

```
Class "ACMESet", directly. Class "ExpressionSet", by class "ACMESet", distance 2. Class "eSet", by class "ACMESet", distance 3. Class "VersionedBiobase", by class "ACMESet", distance 4. Class "Versioned", by class "ACMESet", distance 5.
```

#### Methods

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#### Author(s)

Sean Davis <sdavis2@mail.nih.gov>

#### See Also

**ACMESet** 

# **Examples**

```
showClass("ACMECalcSet")
data(example.agff)
b <- do.aGFF.calc(example.agff,thresh=0.95,window=1000)
b
head(vals(b))
threshold(b)
cutpoints(b)</pre>
```

ACMESet-class

Class "ACMESet"

## **Description**

An extension of ExpressionSet to deal with ACME data including chromosome locations

#### **Objects from the Class**

Objects can be created by calls of the form new("ACMESet", assayData, phenoData, featureData, experimentData, The exprs assayDataElement stores the data. The featureData slot stores the chromosome location.

In practice, the data.frame underlying the featureData MUST contain three columns named chromosome, start, and end; this is enforced by the class validity method.

## Slots

```
assayData: Object of class "AssayData". See ExpressionSet for details.

phenoData: Object of class "AnnotatedDataFrame" See ExpressionSet for details.

featureData: Object of class "AnnotatedDataFrame" See ExpressionSet for details.

experimentData: Object of class "MIAME" See ExpressionSet for details.

annotation: Object of class "character" See ExpressionSet for details.

.__classVersion__: Object of class "Versions" See ExpressionSet for details.
```

#### **Extends**

```
Class "ExpressionSet", directly. Class "eSet", by class "ExpressionSet", distance 2. Class "VersionedBiobase", by class "ExpressionSet", distance 3. Class "Versioned", by class "ExpressionSet", distance 4.
```

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#### Methods

**chromosome** signature(object = "ACMESet"): Accessor for the chromosome. Returns a vector of chromosomes.

end signature(x = "ACMESet"): Accessor for the end location for a probe. If that is not known, this could be set to the same value as the start location.

**start** signature(x = "ACMESet"): Accessor for the start location for a probe.

#### Author(s)

Sean Davis <sdavis2@mail.nih.gov>

#### See Also

ExpressionSet, ACMECalcSet

## **Examples**

```
showClass("ACMESet")
data(example.agff)
example.agff
head(chromosome(example.agff))
head(start(example.agff))
head(end(example.agff))
```

aGFF-class

Class for storing GFF-like data

## **Description**

The GFF format is quite versatile while remaining simple. This class simply stores the annotation associated with a set of GFF files from the same regions of the genome along with some information about the samples from which the data came and the data (from the "score" column of the GFF file) themselves.

## **Objects from the Class**

Objects can be created by calls of the form new("aGFF", ...). Also, the read.resultsGFF() function returns aGFF objects.

#### **Slots**

annotation: Object of class "data.frame" with two columns absolutely necessary, "Chromosome" and "Location". Other columns can be included.

data: Object of class "matrix" of the same number of rows as the annotation slot and the same number of columns as the number of rows in the samples slot, containing data for later analysis samples: Object of class "data.frame" for describing the samples, one row per sample

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#### Methods

```
plot signature(x = "aGFF"): to plot a region along the genome.
print signature(x = "aGFF"): simple method to display summary of aGFF object
show signature(object = "aGFF"): simple method to display summary of aGFF object
```

#### Author(s)

Sean Davis

#### See Also

```
read.resultsGFF andaGFFCalc-class
```

#### **Examples**

```
# Load an example
data(example.agff)
example.agff
```

aGFFCalc-class

Class "aGFFCalc"

#### **Description**

Store results of ACME calculations

# **Objects from the Class**

Objects can be created by calls of the form new("aGFFCalc", ...).

#### **Slots**

```
call: Object of class "call", contains the exact call to do.aGFF.calc, for historical purposes threshold: Object of class "numeric", the threshold used in the calculation cutpoints: Object of class "numeric", the data value above which probes were considered positive
vals: Object of class "matrix", equivalent in size to the original data matrix, containing the calculated p-values from the ACME algorithm
annotation: Object of class "data.frame", currently a copy of the original annotation, possibly reordered in chromosome order
data: Object of class "matrix", the original data, possibly reordered
samples: Object of class "data.frame", sample metadata
```

#### **Extends**

```
Class "aGFF", directly.
```

6 do.aGFF.calc

#### Methods

```
plot signature(x = "aGFFCalc", ask=FALSE): plot the results of an ACME calculation
print signature(x = "aGFFCalc"): brief overview of the object
show signature(object = "aGFFCalc"): brief overview of the object
```

# Author(s)

Sean Davis <sdavis2@mail.nih.gov>

#### See Also

```
do.aGFF.calc, aGFF-class
```

## **Examples**

```
data(example.agff)
example.agffcalc <- do.aGFF.calc(example.agff,window=1000,thresh=0.9)
example.agffcalc</pre>
```

do.aGFF.calc

Perform ACME calculation

#### **Description**

This function performs the moving window chi-square calculation. It is written in C, so is quite fast.

#### Usage

```
do.aGFF.calc(x, window, thresh)
```

# **Arguments**

x An aGFF class object

window An integer value, representing the number of basepairs to include in the win-

dowed chi-square calculation

thresh The quantile of the data distribution for each sample that will be used to classify

a probe as positive

## **Details**

A window size on the order of 2-3 times the average size of fragments from sonication, digestion, etc. and containing at least 8-10 probes is the recommended size. Larger size windows are probably more sensitive, but obviously reduce the accuracy with which boundaries of signal can be called.

A threshold of between 0.9 and 0.99 seems empirically to be adequate. If one plots the histogram of data values and there is an obvious better choice (such as a bimodal distribution, with one peak representing enrichment), a more data-driven approach may yield better results.

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# Value

An object of class aGFFCalc

# Author(s)

Sean Davis <sdavis2@mail.nih.gov>

# **Examples**

```
data(example.agff)
example.agffcalc <- do.aGFF.calc(example.agff,window=1000,thresh=0.9)
example.agffcalc</pre>
```

example.agff

An example ACME data structure of class ACMESet

# Description

An ACMESet data structure from two Nimblegen arrays, custom tiled to include multiple HOX genes.

# Usage

```
data(example.agff)
```

#### **Format**

The format is: chr "example.agff"

#### **Source**

From Scacheri et al., Plot Genet, 2006. Pubmed ID 16604156

```
data(example.agff)
example.agff
```

8 findClosestGene

| findClosestGene | Find closest refseq gene |  |
|-----------------|--------------------------|--|
|-----------------|--------------------------|--|

# **Description**

This function is used to find the nearest refseq transcript(s) to a point in the genome specified. Note that it is limited to the refseq transcripts listed at genome.ucsc.edu, where this function goes for information.

# Usage

```
findClosestGene(chrom, pos, genome = "hg17", position = "txStart")
```

## Arguments

chrom Usually specified like 'chr1', 'chr2', etc.

pos A position in base pairs in the genome
genome Something like 'hg16', 'hg17', 'mm6', etc.

position The location to measure distance from: one of 'txStart', 'txEnd', 'cdsStart',

'cdsEnd'

#### **Details**

The first time the function is run, it checks to see if the refflat table for the given genome is present in the package environment. If not, it downloads it to the /tmp directory and gunzips it (using getRefflat. It is then stored so that in future calls, there is no re-download required.

# Value

A data frame with the gene name, refseq id(s), txStart, txEnd, cdsStart, cdsEnd, exon count, and distance. Note that distance is measured as pos-position, so negative values mean that the point in the gene is to the left of the point specified in the function call (with the p-tel on the left).

# Note

The function may return more than one transcript, as several transcripts may have the same start site

#### Author(s)

Sean Davis <sdavis2@mail.nih.gov>

```
findClosestGene('chr1',100000000,'hg17')
```

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| findRegions | Find all regions in data above p-value threshold |  |
|-------------|--|--|
|             |  |  |

#### **Description**

After the ACME calculation, each probe is associated with a p-value of enrichment. However, one often wants the contiguous regions associated with runs of p-values above a given p-value threshold.

# Usage

```
findRegions(x, thresh = 1e-04)
```

## Arguments

x An ACMESetCalc object thresh The p-value threshold

#### **Details**

Runs of p-values above the p-value threshold will be reported as one "region". These can be used for downstream analyses, export to browsers, submitted for transcription factor binding enrichment analyses, etc.

#### Value

A data frame with these columns:

Length The length of the region in probes

TF Either TRUE or FALSE; TRUE regions represent regions of enrichment while

FALSE regions are the regions between the TRUE regions

StartInd The starting Index of the region EndInd The ending Index of the region Sample The sample containing the region The Chromosome of the region Chromosome The starting basepairof the region Start The ending basepair of the region End Median The median p-value in the region Mean The mean p-value in the region

#### Author(s)

Sean Davis <sdavis2@mail.nih.gov>

#### See Also

```
do.aGFF.calc, findClosestGene
```

10 generics

## **Examples**

```
data(example.agff)
example.agffcalc <- do.aGFF.calc(example.agff,window=1000,thresh=0.9)
foundregions <- findRegions(example.agffcalc,thresh=0.001)
foundregions[1:6,]</pre>
```

generics

Generics defined within ACME

## **Description**

See methods descriptions for details.

# Usage

```
vals(x, ...)
chromosome(object, ...)
end(x, ...)
start(x, ...)
plot(x, y, ...)
cutpoints(x, ...)
threshold(x, ...)
```

# **Arguments**

x An ACMESet or ACMECalcSet object (for cutpoints and threshold)
 object An ACMESet or ACMECalcSet object (for cutpoints and threshold)
 y Treated as missing for plotting these types of objects
 ... Passed into method

#### **Details**

These are all getters for ACMESet and ACMECalcSet objects.

# Value

See methods descriptions for details

## Author(s)

Sean Davis <sdavis2@mail.nih.gov>

#### See Also

ACMESet, ACMECalcSet

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# **Examples**

```
data(example.agff)
head(chromosome(example.agff))
head(end(example.agff))
head(start(example.agff))
```

getRefflat

Get the refflat table from ucsc for the given genome

# Description

Fetches the refflat table from ucsc, stores in temp dir and then gunzips it and reads it in.

# Usage

```
getRefflat(genome = "hg17")
```

### **Arguments**

genome

The genome code from ucsc, like 'hg16', 'mm6', etc.

# Value

A data frame mirroring the UCSC table structure.

# Author(s)

Sean Davis <sdavis2@mail.nih.gov>

# References

http://genome.ucsc.edu

#### See Also

findClosestGene

```
rf <- getRefflat('hg17')</pre>
```

12 read.resultsGFF

|  | read.resultsGFF | Read Nimblegen GFF files |  |
|--|-----------------|--------------------------|--|
|--|-----------------|--------------------------|--|

# Description

A GFF format file is a quite flexible format for storing genomic data. Nimblegen uses these format files as one format for making chip-chip data available. This function reads these files, one per experiment and creates a resulting aGFF-class object.

# Usage

```
read.resultsGFF(fnames, path = ".", samples = NULL, notes = NULL, skip = 0, sep = "\t^{"}", quote = "\t^{"}", .
```

# **Arguments**

| fnames  | A vector of filenames  |
|---------|--|
| path    | The path to the filenames  |
| samples | A data frame containing sample information, one row per sample, in the same order as the files in fnames |
| notes   | A character vector for notes-not currently stored  |
| skip    | Number of lines to skip if the file contains a header  |
| sep     | The field separator–should be a tab character for gff files, but can be set if necessary.                |
| quote   | The text quote character-again not used for gff file, typically  |
|         |  |

# Details

The output is an ACMESet object.

#### Value

A single ACMESet object.

# Author(s)

Sean Davis <sdavis2@mail.nih.gov>

# References

http://www.sanger.ac.uk/Software/formats/GFF/

# See Also

ACMESet

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#### **Examples**

```
datdir <- system.file('extdata',package='ACME')
fnames <- dir(datdir)
example.agff <- read.resultsGFF(fnames,path=datdir)</pre>
```

write.bedGraph

Write bedGraph format tracks for UCSC genome browser

# Description

Generate bedGraph format files for the UCSC genome browser. This function will write the bed-Graph files associated with a aGFFcalc object. There will be either one or two files (default two) representing the raw data and the calculated data (which is output as -log10(val) for visualization purposes for EACH sample).

#### Usage

```
write.bedGraph(x, raw = TRUE, vals = TRUE, directory = ".")
```

### **Arguments**

x An ACMESet or ACMECalcSet object

raw Boolean. Create a file for the raw data?

vals Boolean. Create a file for the calculated p-values?

directory Give a directory for storing the files

# Author(s)

Sean Davis

```
data(example.agff)
write.bedGraph(example.agff)
```

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write.sgr

Write Affy IGB .sgr format files

# **Description**

The affy Integrated Genome Browser (IGB) is a powerful, fast browser for genomic data. The file format is simple (three columns: chromosome, location, and score) to generate. This function will write the sgr files associated with a aGFFcalc object. There will be either one or two files (default two) representing the raw data and the calculated data (which is output as -log10(val) for visualization purposes).

# Usage

```
write.sgr(x, raw = TRUE, vals = TRUE, directory = ".")
```

# **Arguments**

x An ACMESet or ACMECalcSet objectraw Boolean. Create a file for the raw data?

vals Boolean. Create a file for the calculated p-values?

directory Give a directory for storing the files

#### Author(s)

Sean Davis

```
data(example.agff)
write.sgr(example.agff)
```

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