

# Package ‘PathoStat’

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**Type** Package

**Title** PathoStat Statistical Microbiome Analysis Package

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**Description** The purpose of this package is to perform Statistical Microbiome Analysis on metagenomics results from sequencing data samples. In particular, it supports analyses on the PathoScope generated report files. PathoStat provides various functionalities including Relative Abundance charts, Diversity estimates and plots, tests of Differential Abundance, Time Series visualization, and Core OTU analysis.

**URL** <https://github.com/mani2012/PathoStat>

**BugReports** <https://github.com/mani2012/PathoStat/issues>

**License** GPL (>= 2)

**Depends** R (>= 3.5)

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---

Bootstrap\_LOOCV\_LR\_AUC  
*Do bootstrap and LOOCV*

---

**Description**

Do bootstrap and LOOCV

**Usage**

Bootstrap\_LOOCV\_LR\_AUC(df, targetVec, nboot = 50)

**Arguments**

df	Row is sample, column is feature. Required
targetVec	y vector. Required
nboot	number of BOOTSTRAP

**Value**

bootstrap loocv result dataframe

**Examples**

```
data('iris')
Bootstrap_LOOCV_LR_AUC(iris[,1:4],
c(rep(1,100), rep(0,50)), nboot = 3)
```

---

Chisq_Test_Pam	<i>Given PAM and disease/control annotation, do Chi-square test for each row of PAM</i>
----------------	---

---

**Description**

Given PAM and disease/control annotation, do Chi-square test for each row of PAM

**Usage**

```
Chisq_Test_Pam(pam, label.vec.num, pvalue.cutoff = 0.05)
```

**Arguments**

pam	Input data object that contains the data to be tested. Required
label.vec.num	The target binary condition. Required
pvalue.cutoff	choose p-value cut-off

**Value**

df.output object

**Examples**

```
tmp <- matrix(rbinom(12,1,0.5), nrow = 3)
rownames(tmp) <- c("a", "b", "c")
Chisq_Test_Pam(tmp, c(1,1,0,0))
```

---

findRAfromCount	<i>Return the Relative Abundance (RA) data for the given count OTU table</i>
-----------------	--

---

**Description**

Return the Relative Abundance (RA) data for the given count OTU table

**Usage**

```
findRAfromCount(count_otu)
```

**Arguments**

count_otu	Count OTU table
-----------	-----------------

**Value**

ra\_otu Relative Abundance (RA) OTU table

**Examples**

```
data_dir <- system.file("data", package = "PathoStat")
infileName <- "pstat_data.rda"
pstat_test <- loadPstat(data_dir, infileName)
ra_otu <- findRAfromCount(phyloseq::otu_table(pstat_test))
```

---

findTaxonMat	<i>Find the Taxonomy Information Matrix</i>
--------------	---

---

**Description**

Find the Taxonomy Information Matrix

**Usage**

```
findTaxonMat(names, taxonLevels)
```

**Arguments**

names	Row names of the taxonomy matrix
taxonLevels	Taxon Levels of all tids

**Value**

taxmat Taxonomy Information Matrix

**Examples**

```
example_data_dir <- system.file("example/data", package = "PathoStat")
pathoreport_file_suffix <- "-sam-report.tsv"
datlist <- readPathoscopeData(example_data_dir, pathoreport_file_suffix,
input.files.name.vec = as.character(1:6))
dat <- datlist$data
ids <- rownames(dat)
tids <- unlist(lapply(ids, FUN = grepTid))
# taxonLevels <- findTaxonomy(tids[1:5])
# taxmat <- findTaxonMat(ids[1:5], taxonLevels)
```

findTaxonomy

*Find the taxonomy for unlimited tids*

---

**Description**

Find the taxonomy for unlimited tids

**Usage**

```
findTaxonomy(tids)
```

**Arguments**

tids                    Given taxonomy ids

**Value**

taxondata Data with the taxonomy information

**Examples**

```
example_data_dir <- system.file("example/data", package = "PathoStat")
pathoreport_file_suffix <- "-sam-report.tsv"
datlist <- readPathoscopeData(example_data_dir, pathoreport_file_suffix,
input.files.name.vec = as.character(1:6))
dat <- datlist$data
ids <- rownames(dat)
tids <- unlist(lapply(ids, FUN = grepTid))
# taxonLevels <- findTaxonomy(tids[1:5])
```

---

findTaxonomy300*Find the taxonomy for maximum 300 tids*

---

**Description**

Find the taxonomy for maximum 300 tids

**Usage**

```
findTaxonomy300(tids)
```

**Arguments**

tids                    Given taxonomy ids

**Value**

taxondata Data with the taxonomy information

**Examples**

```
example_data_dir <- system.file("example/data", package = "PathoStat")
pathoreport_file_suffix <- "-sam-report.tsv"
datlist <- readPathoscopeData(example_data_dir,
  pathoreport_file_suffix, input.files.name.vec = as.character(1:6))
dat <- datlist$data
ids <- rownames(dat)
tids <- unlist(lapply(ids, FUN = grepTid))
# taxonLevels <- findTaxonomy300(tids[1:5])
```

---

Fisher_Test_Pam	<i>Given PAM and disease/control annotation, do Chi-square test for each row of PAM</i>
-----------------	---

---

**Description**

Given PAM and disease/control annotation, do Chi-square test for each row of PAM

**Usage**

```
Fisher_Test_Pam(pam, label.vec.num, pvalue.cutoff = 0.05)
```

**Arguments**

pam	Input data object that contains the data to be tested. Required
label.vec.num	The target binary condition. Required
pvalue.cutoff	choose p-value cut-off

**Value**

df.output object

**Examples**

```
tmp <- matrix(rbinom(12,1,0.5), nrow = 3)
rownames(tmp) <- c("a", "b", "c")
Fisher_Test_Pam(tmp, c(1,1,0,0))
```

---

formatTaxTable	<i>Format taxonomy table for rendering</i>
----------------	--

---

**Description**

Format taxonomy table for rendering

**Usage**

```
formatTaxTable(ttable)
```

**Arguments**

ttable	Taxonomy table
--------	----------------

**Value**

Formatted table suitable for rendering with. DT::renderDataTable

---

getShinyInput	<i>Getter function to get the shinyInput option</i>
---------------	---

---

**Description**

Getter function to get the shinyInput option

**Usage**

```
getShinyInput()
```

**Value**

shinyInput option

**Examples**

```
getShinyInput()
```



---

`getShinyInputCombat`     *Getter function to get the shinyInputCombat option*

---

**Description**

Getter function to get the shinyInputCombat option

**Usage**

```
getShinyInputCombat()
```

**Value**

shinyInputCombat option

**Examples**

```
getShinyInputCombat()
```

---

`getShinyInputOrig`     *Getter function to get the shinyInputOrig option*

---

**Description**

Getter function to get the shinyInputOrig option

**Usage**

```
getShinyInputOrig()
```

**Value**

shinyInputOrig option

**Examples**

```
getShinyInputOrig()
```

getSignatureFromMultipleGlmnet

*Use Lasso to do feature selection*

---

### Description

Use Lasso to do feature selection

### Usage

```
getSignatureFromMultipleGlmnet(df.input, target.vec, nfolds = 10,  
  logisticRegression = TRUE, nRun = 100, alpha = 1)
```

### Arguments

df.input	Row is sample, column is feature. Required
target.vec	y vector. Required
nfolds	glmnet CV nfolds
logisticRegression	doing logistic regression or linear regression.
nRun	number of glmnet runs
alpha	same as in glmnet

### Value

signature

### Examples

```
data('iris')  
getSignatureFromMultipleGlmnet(iris[,1:4],  
  c(rep(1,100), rep(0,50)), nfolds = 3, nRun = 10)
```

---

GET\_PAM

*transform cpm counts to presence-absence matrix*

---

### Description

transform cpm counts to presence-absence matrix

### Usage

```
GET_PAM(df)
```

**Arguments**

df                    Input data object that contains the data to be tested. Required

**Value**

df.output object

**Examples**

```
GET_PAM(data.frame(a = c(1,3,0), b = c(0,0.1,2)))
```

---

grepTid                    *Greps the tid from the given identifier string*

---

**Description**

Greps the tid from the given identifier string

**Usage**

```
grepTid(id)
```

**Arguments**

id                    Given identifier string

**Value**

tid string

**Examples**

```
grepTid("ti|700015|org|Coriobacterium_glomerans_PW2")
```

---

`loadPathoscopeReports` *Loads all data from a set of PathoID reports. For each column in the PathoID report, construct a matrix where the rows are genomes and the columns are samples. Returns a list where each element is named according to the PathoID column. For example, `ret[["Final.Best.Hit.Read.Numbers"]]` on the result of this function will get you the final count matrix. Also includes elements "total\_reads" and "total\_genomes" from the first line of the PathoID report.*

---

### Description

Loads all data from a set of PathoID reports. For each column in the PathoID report, construct a matrix where the rows are genomes and the columns are samples. Returns a list where each element is named according to the PathoID column. For example, `ret[["Final.Best.Hit.Read.Numbers"]]` on the result of this function will get you the final count matrix. Also includes elements "total\_reads" and "total\_genomes" from the first line of the PathoID report.

### Usage

```
loadPathoscopeReports(reportfiles, nrows = NULL)
```

### Arguments

<code>reportfiles</code>	Paths to report files
<code>nrows</code>	Option to read first N rows of PathoScope reports

### Value

Returns a list where each element is named according to the PathoID column. For example, `ret[["Final.Best.Hit.Read.Numbers"]]` on the result of this function will get you the final count matrix. Also includes elements "total\_reads" and "total\_genomes" from the first line of the PathoID report.

### Examples

```
input_dir <- system.file("example/data", package = "PathoStat")
reportfiles <- list.files(input_dir, pattern = "*-sam-report.tsv",
full.names = TRUE)
```

---

loadPstat	<i>Load the R data(.rda) file with pathostat object</i>
-----------	---

---

**Description**

Load the R data(.rda) file with pathostat object

**Usage**

```
loadPstat(indir = ".", infileName = "pstat_data.rda")
```

**Arguments**

indir	Input Directory of the .rda file
infileName	File name of the .rda file

**Value**

pstat pathostat object (NULL if it does not exist)

**Examples**

```
data_dir <- system.file("data", package = "PathoStat")
infileName <- "pstat_data.rda"
pstat <- loadPstat(data_dir, infileName)
```

---

log2CPM	<i>Compute log2(counts per mil reads) and library size for each sample</i>
---------	--

---

**Description**

Compute log2(counts per mil reads) and library size for each sample

**Usage**

```
log2CPM(qcounts, lib.size = NULL)
```

**Arguments**

qcounts	quantile normalized counts
lib.size	default is colsums(qcounts)

**Value**

list containing log2(quantile counts per mil reads) and library sizes

**Examples**

```
log2CPM(matrix(1:12, nrow = 3))
```

---

```
LOOAUC_simple_multiple_noplot_one_df
      LOOCV
```

---

**Description**

LOOCV

**Usage**

```
LOOAUC_simple_multiple_noplot_one_df(df, targetVec)
```

**Arguments**

df                    Row is sample, column is feature. Required  
targetVec            y vector. Required

**Value**

mean auc

**Examples**

```
data('iris')
LOOAUC_simple_multiple_noplot_one_df(iris[,1:4],
c(rep(1,100), rep(0,50)))
```

---

```
LOOAUC_simple_multiple_one_df
      LOOCV with ROC curve
```

---

**Description**

LOOCV with ROC curve

**Usage**

```
LOOAUC_simple_multiple_one_df(df, targetVec)
```

**Arguments**

df                    Row is sample, column is feature. Required  
targetVec            y vector. Required

**Value**

the ROC

**Examples**

```
data('iris')
LOOAUC_simple_multiple_one_df(iris[,1:4],
c(rep(1,100), rep(0,50)))
```

---

PathoStat-class	<i>PathoStat class to store PathoStat input data including phyloseq object</i>
-----------------	--

---

**Description**

Contains all currently-supported BatchQC output data classes:

**Details**

slots:

**average\_count** a single object of class `otu_tableOrNULL`

**besthit\_count** a single object of class `otu_tableOrNULL`

**highconf\_count** a single object of class `otu_tableOrNULL`

**lowconf\_count** a single object of class `otu_tableOrNULL`

**Examples**

```
otumat = matrix(sample(1:100, 100, replace = TRUE), nrow = 10, ncol = 10)
rownames(otumat) <- paste0("OTU", 1:nrow(otumat))
colnames(otumat) <- paste0("Sample", 1:ncol(otumat))
taxmat = matrix(sample(letters, 70, replace = TRUE),
nrow = nrow(otumat), ncol = 7)
rownames(taxmat) <- rownames(otumat)
colnames(taxmat) <- c("Domain", "Phylum", "Class",
"Order", "Family", "Genus", "Species")
OTU = phyloseq::otu_table(otumat, taxa_are_rows = TRUE)
TAX = phyloseq::tax_table(taxmat)
physeq = phyloseq::phyloseq(OTU, TAX)
pathostat1(physeq)
```

---

percent	<i>Compute percentage</i>
---------	---------------------------

---

**Description**

Compute percentage

**Usage**

```
percent(x, digits = 2, format = "f")
```

**Arguments**

x	a number or a vector
digits	how many digit of percentage
format	numeric format, "f" for float

**Value**

the percentage

**Examples**

```
percent.vec <- percent(c(0.9, 0.98))
```

---

phyloseq_to_edgeR	<i>Convert phyloseq OTU count data into DGEList for edgeR package</i>
-------------------	---

---

**Description**

Further details.

**Usage**

```
phyloseq_to_edgeR(physeq, group, method = "RLE", ...)
```

**Arguments**

physeq	(Required).
group	(Required). A character vector or factor giving the experimental group/condition for each sample/library.
method	(Optional).
...	Additional arguments passed on to



**Value**

dispersion

**Examples**

```
data_dir_test <- system.file("data", package = "PathoStat")
pstat_test <- loadPstat(indir=data_dir_test,
  infileName="pstat_data_2_L1.rda")
phyloseq_to_edgeR(pstat_test, group="Sex")
```

---

plotPCAPlotly

*Plot PCA*

---

**Description**

Plot PCA

**Usage**

```
plotPCAPlotly(df.input, condition.color.vec,
  condition.color.name = "condition", condition.shape.vec = NULL,
  condition.shape.name = "condition", columnTitle = "Title",
  pc.a = "PC1", pc.b = "PC2")
```

**Arguments**

df.input	Input data object that contains the data to be plotted. Required
condition.color.vec	color vector. Required
condition.color.name	color variable name. Required
condition.shape.vec	shape vector. Required
condition.shape.name	shape variable name. Required
columnTitle	Title to be displayed at top of heatmap.
pc.a	pc.1
pc.b	pc.2

**Value**

the plot

**Examples**

```
data('iris')
plotPCAPlotly(t(iris[,1:4]),
condition.color.vec = c(rep(1,100), rep(0,50)),
condition.shape.vec = c(rep(0,100), rep(1,50)))
```

---

plotPCoAPlotly

*Plot PCoA*


---

**Description**

Plot PCoA

**Usage**

```
plotPCoAPlotly(physeq.input, condition.color.vec,
condition.color.name = "condition", condition.shape.vec = NULL,
condition.shape.name = "condition", method = "bray",
columnTitle = "Title", pc.a = "Axis.1", pc.b = "Axis.2")
```

**Arguments**

physeq.input	Input data object that contains the data to be plotted. Required
condition.color.vec	color vector. Required
condition.color.name	color variable name. Required
condition.shape.vec	shape vector. Required
condition.shape.name	shape variable name. Required
method	which distance metric
columnTitle	Title to be displayed at top of heatmap.
pc.a	pc.1
pc.b	pc.2

**Value**

the plot

**Examples**

```
data_dir_test <- system.file("data", package = "PathoStat")
pstat_test <- loadPstat(indir=data_dir_test,
infileName="pstat_data_2_L1.rda")
plotPCoAPlotly(pstat_test, condition.color.vec = rbinom(33,1,0.5),
condition.shape.vec = rbinom(33,1,0.5))
```

---

pstat_data	<i>pathostat object generated from example pathoscope report files</i>
------------	--

---

**Description**

This example data consists of 33 samples from a diet study with 11 subjects taking 3 different diets in random order

**Usage**

```
pstat
```

**Format**

pathostat object extension of phyloseq-class experiment-level object:

**otu\_table** OTU table with 41 taxa and 33 samples

**sample\_data** Sample Data with 33 samples by 18 sample variables

**tax\_table** Taxonomy Table with 41 taxa by 9 taxonomic ranks

**sample\_data** Phylogenetic Tree with 41 tips and 40 internal nodes

**Value**

pathostat object

---

readPathoscopeData	<i>Reads the data from PathoScope reports and returns a list of final guess relative abundance and count data</i>
--------------------	---

---

**Description**

Reads the data from PathoScope reports and returns a list of final guess relative abundance and count data

**Usage**

```
readPathoscopeData(input_dir = ".",
  pathoreport_file_suffix = "-sam-report.tsv", use.input.files = FALSE,
  input.files.path.vec = NULL, input.files.name.vec = NULL)
```

**Arguments**

Directory where the tsv files from PathoScope are located  
 pathoreport\_file\_suffix PathoScope report files suffix  
 use.input.files whether input dir to pathoscope files or directly pathoscope files  
 input.files.path.vec vector of pathoscope file paths  
 input.files.name.vec vector of pathoscope file names

**Value**

List of final guess relative abundance and count data

**Examples**

```

example_data_dir <- system.file("example/data", package = "PathoStat")
pathoreport_file_suffix <- "-sam-report.tsv"
datlist <- readPathoscopeData(example_data_dir, pathoreport_file_suffix,
input.files.name.vec = as.character(1:6))
  
```

---

runPathoStat	<i>Statistical Microbiome Analysis on the pathostat input and generates a html report and produces interactive shiny app plots</i>
--------------	--

---

**Description**

Statistical Microbiome Analysis on the pathostat input and generates a html report and produces interactive shiny app plots

**Usage**

```

runPathoStat(pstat = NULL, report_dir = ".",
report_option_binary = "111111111", interactive = TRUE)
  
```

**Arguments**

pstat phyloseq extension pathostat object  
 report\_dir Output report directory path  
 report\_option\_binary 9 bits Binary String representing the plots to display and hide in the report  
 interactive when TRUE, opens the interactive shinyApp

**Value**

outputfile The output file with all the statistical plots

**Examples**

```
runPathoStat(interactive = FALSE)
```

---

savePstat	<i>Save the pathostat object to R data(.rda) file</i>
-----------	---

---

**Description**

Save the pathostat object to R data(.rda) file

**Usage**

```
savePstat(pstat, outdir = ".", outfileName = "pstat_data.rda")
```

**Arguments**

pstat	pathostat object
outdir	Output Directory of the .rda file
outfileName	File name of the .rda file

**Value**

outfile .rda file

**Examples**

```
data_dir_test <- system.file("data", package = "PathoStat")
pstat_test <- loadPstat(indir=data_dir_test,
  infileName="pstat_data_2_L1.rda")
outfile <- savePstat(pstat_test)
```

---

setShinyInput	<i>Setter function to set the shinyInput option</i>
---------------	---

---

**Description**

Setter function to set the shinyInput option

**Usage**

```
setShinyInput(x)
```

**Arguments**

x	shinyInput option
---	-------------------

**Value**

shinyInput option

**Examples**

```
setShinyInput(NULL)
```

---

setShinyInputCombat     *Setter function to set the shinyInputCombat option*

---

**Description**

Setter function to set the shinyInputCombat option

**Usage**

```
setShinyInputCombat(x)
```

**Arguments**

x                    shinyInputCombat option

**Value**

shinyInputCombat option

**Examples**

```
setShinyInputCombat(NULL)
```

---

setShinyInputOrig     *Setter function to set the shinyInputOrig option*

---

**Description**

Setter function to set the shinyInputOrig option

**Usage**

```
setShinyInputOrig(x)
```

**Arguments**

x                    shinyInputOrig option

**Value**

shinyInputOrig option

**Examples**

```
setShinyInputOrig(NULL)
```

---

summarizeTable	<i>Summarize sample</i>
----------------	-------------------------

---

**Description**

Creates a table of summary metrics

**Usage**

```
summarizeTable(pstat)
```

**Arguments**

pstat            Input pstat

**Value**

A data.frame object of summary metrics.

**Examples**

```
data_dir_test <- system.file("data", package = "PathoStat")
pstat_test <- loadPstat(indir=data_dir_test,
  infileName="pstat_data_2_L1.rda")
st.tmp <- summarizeTable(pstat_test)
```

---

TranslateIdToTaxLevel	<i>Find the taxonomy for the given taxon id name</i>
-----------------------	--

---

**Description**

Find the taxonomy for the given taxon id name

**Usage**

```
TranslateIdToTaxLevel(pstat, input.id.vec, tax.level)
```

**Arguments**

pstat	pathostat object
input.id.vec	names containing id
tax.level	target taxon level

**Value**

target taxon level names

**Examples**

```
data_dir_test <- system.file("data", package = "PathoStat")
pstat_test <- loadPstat(indir=data_dir_test,
  infileName="pstat_data_2_L1.rda")
names.new <- TranslateIdToTaxLevel(pstat_test,
  c("ti|862962|org|Bacteroides_fragilis_638R",
    "ti|697329|org|Ruminococcus_albus_7" ),
  "genus")
```

---

Wilcox\_Test\_df

*Mann-whitney test for a dataframe*

---

**Description**

Mann-whitney test for a dataframe

**Usage**

```
Wilcox_Test_df(df, label.vec.num, pvalue.cutoff = 0.05)
```

**Arguments**

df	Input data object that contains the data to be tested. Required
label.vec.num	The target binary condition. Required
pvalue.cutoff	choose p-value cut-off

**Value**

df.output object

**Examples**

```
data('iris')
Wilcox_Test_df(t(iris[,1:4]),
  c(rep(1,100), rep(0,50)))
```



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