

# Package ‘ballgown’

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**License** Artistic-2.0

**Title** Flexible, isoform-level differential expression analysis

**Description** Tools for statistical analysis of assembled transcriptomes, including flexible differential expression analysis, visualization of transcript structures, and matching of assembled transcripts to annotation.

**Depends** R (>= 3.1.1), methods

**Imports** GenomicRanges (>= 1.17.25), IRanges (>= 1.99.22), S4Vectors (>= 0.9.39), RColorBrewer, splines, sva, limma, rtracklayer (>= 1.29.25), Biobase (>= 2.25.0), GenomeInfoDb

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**VignetteBuilder** knitr

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---

ballgown-package

*The ballgown package for analysis of transcript assemblies*


---

## Description

Super awesome transcript-level expression analysis

---

annotate\_assembly      *match assembled transcripts to annotated transcripts*

---

### Description

match assembled transcripts to annotated transcripts

### Usage

```
annotate_assembly(assembled, annotated)
```

### Arguments

assembled      GRangesList object representing assembled transcripts  
 annotated      GRangesList object representing annotated transcripts

### Details

If gown is a ballgown object, assembled can be structure(gown)\$trans (or any subset). You can generate a GRangesList object containing annotated transcripts from a gtf file using the [gffReadGR](#) function and setting splitByTranscripts=TRUE.

### Value

data frame, where each row contains assembledInd and annotatedInd (indexes of overlapping transcripts in assembled and annotated), and the percent overlap between the two transcripts.

### Author(s)

Alyssa Frazee

### Examples

```
data(bg)
gtfPath = system.file('extdata', 'annot.gtf.gz', package='ballgown')
annot = gffReadGR(gtfPath, splitByTranscript=TRUE)
info = annotate_assembly(assembled=structure(bg)$trans, annotated=annot)
```

---

ballgown-class      *Ballgown*

---

### Description

S4 class for storing and manipulating expression data from assembled transcriptomes

**Slots**

**expr** tables containing expression data for genomic features (introns, exons, transcripts)  
**structure** genomic locations of features and their relationships to one another  
**indexes** tables connecting components of the assembly and providing other experimental information (e.g., phenotype data and locations of read alignment files)  
**dirs** directories holding data created by tablemaker  
**mergedDate** date the ballgown object was created  
**meas** which expression measurement(s) the object contains in its data slot. Vector of one or more of "rcount", "ucount", "mrcount", "cov", "cov\_sd", "mcov", "mcov\_sd", or "FPKM", if Tablemaker output is used, or one of "TPM" or "FPKM" if RSEM output is used. Can also be "all" for all measurements. See vignette for details.  
**RSEM** TRUE if object was made from RSEM output, FALSE if object was made from Tablemaker/Cufflinks output.

**Author(s)**

Alyssa Frazee, Leonardo Collado-Torres, Jeff Leek

**Examples**

```

data(bg)
class(bg) #"ballgown"
dim(bg@expr$exon)
bg@structure$exon
head(bg@indexes$t2g)
head(bg@dirs)
bg@mergedDate
bg@meas
bg@RSEM
  
```

---

ballgown-constructor    *constructor function for ballgown objects*

---

**Description**

constructor function for ballgown objects

**Usage**

```

ballgown(
  samples = NULL,
  dataDir = NULL,
  samplePattern = NULL,
  bamfiles = NULL,
  pData = NULL,
  verbose = TRUE,
  meas = "all"
)
  
```

**Arguments**

<code>samples</code>	vector of file paths to folders containing sample-specific ballgown data (generated by <code>tablemaker</code> ). If <code>samples</code> is provided, <code>dataDir</code> and <code>samplePattern</code> are not used.
<code>dataDir</code>	file path to top-level directory containing sample-specific folders with ballgown data in them. Only used if <code>samples</code> is <code>NULL</code> .
<code>samplePattern</code>	regular expression identifying the subdirectories of <code>dataDir</code> containing data to be loaded into the ballgown object (and only those subdirectories). Only used if <code>samples</code> is <code>NULL</code> .
<code>bamfiles</code>	optional vector of file paths to read alignment files for each sample. If provided, make sure to sort properly (e.g., in the same order as <code>samples</code> ). Default <code>NULL</code> .
<code>pData</code>	optional <code>data.frame</code> with rows corresponding to <code>samples</code> and columns corresponding to phenotypic variables.
<code>verbose</code>	if <code>TRUE</code> , print status messages and timing information as the object is constructed.
<code>meas</code>	character vector containing either "all" or one or more of: "rcount", "ucount", "mrcount", "cov", "cov_sd", "mcov", "mcov_sd", or "FPKM". The resulting ballgown object will only contain the specified expression measurements, for the appropriate features. See vignette for which expression measurements are available for which features. "all" creates the full object.

**Details**

Because experimental data is recorded so variably, it is the user's responsibility to format `pData` correctly. In particular, it's really important that the rows of `pData` (corresponding to `samples`) are ordered the same way as `samples` or the `dataDir/samplePattern` combo. You can run `list.files(path = dataDir, pattern = samplePattern)` to see the sample order if `samples` was not used.

If you are creating a ballgown object for a large experiment, this function may run slowly and use a large amount of RAM. We recommend running this constructor as a batch job and saving the resulting ballgown object as an `rda` file. The `rda` file usually has reasonable size on disk, and the object in it shouldn't take up too much RAM when loaded, so the time and memory use in creating the object is a one-time cost.

**Value**

an object of class `ballgown`

**Author(s)**

Leonardo Collado-Torres, Alyssa Frazee

**See Also**

[ballgownrsem](#), for loading RSEM output into a ballgown object

**Examples**

```
bg = ballgown(dataDir=system.file('extdata', package='ballgown'),
             samplePattern='sample')
pData(bg) = data.frame(id=sampleNames(bg), group=rep(c(1,0), each=10))
```

---

ballgownrsem

*load RSEM data into a ballgown object*


---

### Description

Loads results of rsem-calculate-expression into a ballgown object for easy visualization, processing, and statistical testing

### Usage

```
ballgownrsem(
  dir = "",
  samples,
  gtf,
  UCSC = TRUE,
  tfield = "transcript_id",
  attrsep = "; ",
  bamout = "transcript",
  pData = NULL,
  verbose = TRUE,
  meas = "all",
  zipped = FALSE
)
```

### Arguments

dir	output directory containing RSEM output for all samples (i.e. for each run of rsem-calculate-expression)
samples	vector of sample names (i.e., of the sample_name arguments used in each RSEM run)
gtf	path to GTF file of genes/transcripts used in your RSEM reference. (where the reference location was denoted by the reference_name argument used in rsem-calculate-expression). RSEM references can be created with or without a GTF file, but currently the ballgown reader requires the GTF file.
UCSC	set to TRUE if gtf comes from UCSC: quotes will be stripped from transcript identifiers if so.
tfield	What keyword identifies transcripts in the "attributes" field of gtf? Default 'transcript_id'.
attrsep	How are attributes separated in the "attributes" field of gtf? Default ' ; ' (semicolon-space).
bamout	set to 'genome' if --output-genome-bam was used when running rsem-calculate-expression; set to 'none' if --no-bam-output was used when running rsem-calculate-expression; otherwise use the default ('transcript').
pData	data frame of phenotype data, with rows corresponding to samples. The first column of pData must be equal to samples, and rows must be in the same order as samples.
verbose	If TRUE (as by default), status messages are printed during data loading.

meas	character vector containing either "all" or one of "FPKM" or "TPM". The resulting ballgown object will only contain the specified expression measurement for the transcripts. "all" creates the full object.
zipped	set to TRUE if all RSEM results files have been gzipped (end in ".gz").

### Details

Currently exon- and intron-level measurements are not available for RSEM-generated ballgown objects, but development is ongoing.

### Value

a ballgown object with the specified expression measurements and structure specified by GTF.

### See Also

[ballgown](#) for reading Cufflinks/Tablemaker output

### Examples

```
dataDir = system.file('extdata', package='ballgown')
gtf = file.path(dataDir, 'hg19_genes_small.gtf.gz')
rsemobj = ballgownrsem(dir=dataDir, samples=c('tiny', 'tiny2'), gtf=gtf,
  bamout='none', zipped=TRUE)
rsemobj
```

---

bg	<i>Toy ballgown object</i>
----	----------------------------

---

### Description

Small ballgown object created with simulated toy data, for demonstration purposes

### Format

a ballgown object: 100 transcripts, 633 exons, 536 introns

### Author(s)

Alyssa Frazee

### Examples

```
data(bg)
bg
# ballgown instance with 100 transcripts and 20 samples
```

---

checkAssembledTx      *plot annotated and assembled transcripts together*

---

### Description

plot annotated and assembled transcripts together

### Usage

```
checkAssembledTx(  
  assembled,  
  annotated,  
  ind = 1,  
  main = "Assembled and Annotated Transcripts",  
  customCol = NULL  
)
```

### Arguments

assembled	a GRangesList object where the GRanges objects in the list represent sets of exons comprising assembled transcripts
annotated	a GRangesList object where the GRanges objects in the list represent sets of exons comprising annotated transcripts
ind	integer; index of annotated specifying which annotated transcript to plot. All transcripts (assembled and annotated) overlapping annotated[[ind]] will be plotted. Default 1.
main	optional character string giving the title for the resulting plot. Default: "Assembled and Annotated Transcripts"
customCol	optional vector of custom colors for the annotated transcripts. If not the same length as the number of annotated transcripts in the plot, recycling or truncation might occur.

### Value

Plots annotated transcripts on the bottom panel (shaded in gray) and assembled transcripts on the top panel (shaded with diagonal lines).

### Author(s)

Alyssa Frazee

### Examples

```
gtfPath = system.file('extdata', 'annot.gtf.gz', package='ballgown')  
annot = gffReadGR(gtfPath, splitByTranscript=TRUE)  
data(bg)  
checkAssembledTx(annotated=annot, assembled=structure(bg)$trans, ind=4)
```



---

clusterTranscripts     *group a gene's assembled transcripts into clusters*

---

### Description

group a gene's assembled transcripts into clusters

### Usage

```
clusterTranscripts(gene, gown, k = NULL, method = c("hclust", "kmeans"))
```

### Arguments

gene	name of gene whose transcripts will be clustered. When using Cufflinks output, usually of the form "XLOC_#####"
gown	ballgown object containing experimental data
k	number of clusters to use
method	clustering method to use. Must be one of "hclust", for hierarchical clustering, or "kmeans", for k-means clustering.

### Value

list with elements `clusters` and `pctvar`. `clusters` contains columns "cluster" and "t\_id", and denotes which transcripts belong to which clusters. `pctvar` is only non-NULL when using k-means clustering and is the percentage of variation explained by these clusters, defined as the ratio of the between-cluster sum of squares to the total sum of squares.

### Author(s)

Alyssa Frazee

### See Also

[hclust](#), [kmeans](#), [plotLatentTranscripts](#) for visualizing the transcript clusters

### Examples

```
data(bg)
clusterTranscripts('XLOC_000454', bg, k=2, method='kmeans')
# transcripts 1294 and 1301 cluster together, 91% variation explained.
```

---

collapseTranscripts    *cluster a gene's transcripts and calculate cluster-level expression*

---

### Description

cluster a gene's transcripts and calculate cluster-level expression

### Usage

```
collapseTranscripts(
  gene,
  gown,
  meas = "FPKM",
  method = c("hclust", "kmeans"),
  k = NULL
)
```

### Arguments

gene	which gene's transcripts should be clustered
gown	ballgown object
meas	which transcript-level expression measurement to use ('cov', average per-base coverage, or 'FPKM')
method	which clustering method to use: 'hclust' (hierarchical clustering) or 'kmeans' (k-means clustering).
k	how many clusters to use.

### Value

list with two elements:

- `tab`, a cluster-by-sample table of expression measurements (`meas`, either `cov` or `FPKM`), where the expression measurement for each cluster is the mean (for `cov`) or aggregate (for `FPKM`, as in `gexpr`) expression measurement for all the transcripts in that cluster. This table can be used as the `gowntable` argument to `stattest`, if differential expression results for transcript `*clusters*` are desired.
- `c1` output from `clusterTranscripts` that was run to produce `tab`, for reference. Cluster IDs in the `cluster` component correspond to row names of `tab`

### Author(s)

Alyssa Frazee

### See Also

[hclust](#), [kmeans](#), [clusterTranscripts](#), [plotLatentTranscripts](#)

### Examples

```
data(bg)
collapseTranscripts(bg, gene='XLOC_000454', meas='FPKM', method='kmeans')
```

---

contains	<i>determine if one set of GRanges fully contains any of another set of GRanges</i>
----------	---

---

**Description**

determine if one set of GRanges fully contains any of another set of GRanges

**Usage**

```
contains(transcripts, cds)
```

**Arguments**

transcripts	GRangesList object (assume for now that it represents transcripts)
cds	GRangesList object (assume for now that it represents sets of coding sequences)

**Details**

If gown is a ballgown object, transcripts can be `structure(gown)$trans` (or any subset).

**Value**

vector with length equal to `length(transcripts)`, where each entry is TRUE if the corresponding transcript contains a coding sequence (i.e., is a superset of at least one entry of `cds`).

**Author(s)**

Alyssa Frazee

**Examples**

```
## pretend this annotation is coding sequence:
gtfPath = system.file('extdata', 'annot.gtf.gz', package='ballgown')
annot = gffReadGR(gtfPath, splitByTranscript=TRUE)
data(bg)
results = contains(structure(bg)$trans, annot)
# results is a boolean vector
sum(results) #61
```

---

dirs	<i>extract paths to tablemaker output</i>
------	---

---

**Description**

extract paths to tablemaker output

**Usage**

```

dirs(x)

## S4 method for signature 'ballgown'
dirs(x)

```

**Arguments**

x                    a ballgown object

**Examples**

```

data(bg)
dirs(bg)

```

---

eexpr

*extract exon-level expression measurements from ballgown objects*


---

**Description**

extract exon-level expression measurements from ballgown objects

**Usage**

```

eexpr(x, meas = "rcount")

## S4 method for signature 'ballgown'
eexpr(x, meas = "rcount")

```

**Arguments**

x                    a ballgown object

meas                type of measurement to extract. Can be "rcount", "ucount", "mrcount", "cov", "mcov", or "all". Default "rcount".

**Value**

exon-by-sample matrix containing exon-level expression values (measured by meas). If meas is "all", or x@RSEM is TRUE, a data frame is returned, containing all measurements and location information.

**Examples**

```

data(bg)
exon_rcount_matrix = eexpr(bg)
exon_ucount_matrix = eexpr(bg, 'ucount')
exon_data_frame = eexpr(bg, 'all')

```

---

expr	<i>extract expression components from ballgown objects</i>
------	--

---

**Description**

extract expression components from ballgown objects

**Usage**

```
expr(x)
```

```
## S4 method for signature 'ballgown'  
expr(x)
```

**Arguments**

x                    a ballgown object

**Value**

list containing elements intron, exon, and trans, which are feature-by-sample data frames of expression data.

**See Also**

[texpr](#), [gexpr](#), [eexpr](#), [iexpr](#)

**Examples**

```
data(bg)  
names(expr(bg))  
class(expr(bg))  
dim(expr(bg)$exon)
```

---

expr<-	<i>Replacement method for expr slot in ballgown objects</i>
--------	---

---

**Description**

Replacement method for expr slot in ballgown objects

**Usage**

```
expr(x) <- value
```

```
## S4 replacement method for signature 'ballgown'  
expr(x) <- value
```

**Arguments**

x	a ballgown object
value	the updated value for expr(x) or a subcomponent

**Examples**

```
data(bg)
n = ncol(bg@expr$trans)
#multiply all transcript expression measurements by 10:
bg@expr$trans[,11:n] = 10*bg@expr$trans[11:n]
```

---

exprfilter	<i>subset ballgown objects using an expression filter</i>
------------	---

---

**Description**

Create a new ballgown object containing only transcripts passing a mean expression filter

**Usage**

```
exprfilter(gown, cutoff, meas = "FPKM")
```

**Arguments**

gown	a ballgown object
cutoff	transcripts must have mean expression across samples above this value to be included in the return
meas	how should transcript expression be measured? Default FPKM, but can also be 'cov'.

**Value**

A new ballgown object derived from gown, but only containing transcripts (and associated exons/introns) with mean meas greater than cutoff across all samples.

**See Also**

[subset](#)

**Examples**

```
data(bg)
# make a ballgown object containing only transcripts with mean FPKM > 100:
over100 = exprfilter(bg, cutoff=100)
```

---

geneIDs	<i>get gene IDs from a ballgown object</i>
---------	--

---

**Description**

get gene IDs from a ballgown object

**Usage**

```
geneIDs(x)  
  
## S4 method for signature 'ballgown'  
geneIDs(x)
```

**Arguments**

x                    a ballgown object

**Details**

This vector differs from that produced by `geneNames` in that `geneIDs` produces names of loci created during the assembly process, not necessarily annotated genes.

**Value**

named vector of gene IDs included in the ballgown object. If object was created using `Tablemaker`, these gene IDs will be of the form "XLOC\_\*". Vector is named and ordered by corresponding numeric transcript ID.

**See Also**

[geneNames](#)

**Examples**

```
data(bg)  
geneIDs(bg)
```

---

geneNames	<i>get gene names from a ballgown object</i>
-----------	--

---

**Description**

get gene names from a ballgown object

**Usage**

```
geneNames(x)  
  
## S4 method for signature 'ballgown'  
geneNames(x)
```

**Arguments**

x                    a ballgown object

**Details**

This vector differs from that produced by `geneIDs` in that `geneNames` produces *\*annotated\** gene names that correspond to assembled transcripts. The return will be empty/blank/NA if the transcriptome assembly is de novo (i.e., was not compared to an annotation before the ballgown object was created). See [getGenes](#) for matching transcripts to gene names. Some entries of this vector will be empty/blank/NA if the corresponding transcript did not overlap any annotated genes.

**Value**

named vector of gene names included in the ballgown object, named and ordered by corresponding numeric transcript ID.

**See Also**

[geneIDs](#)

**Examples**

```
data(bg)
# this is a de novo assembly, so it does not contain gene info as it stands
# but we can add it:
annot = system.file('extdata', 'annot.gtf.gz', package='ballgown')
gnames = getGenes(annot, structure(bg)$trans, UCSC=FALSE)
gnames_first = lapply(gnames, function(x) x[1]) #just take 1 overlapping gene
expr(bg)$trans$gene_name = gnames_first

# now we can extract these gene names:
geneNames(bg)
```

---

<code>getAttributeField</code>	<i>extract a specific field of the "attributes" column of a data frame created from a GTF/GFF file</i>
--------------------------------	--

---

**Description**

extract a specific field of the "attributes" column of a data frame created from a GTF/GFF file

**Usage**

```
getAttributeField(x, field, attrsep = "; ")
```

**Arguments**

x                    vector representing the "attributes" column of GTF/GFF file  
field                name of the field you want to extract from the "attributes" column  
attrsep             separator for the fields in the attributes column. Defaults to '; ', the separator for GTF files outputted by Cufflinks.



**Value**

vector of nucleotide positions included in the transcript

**Author(s)**

Wolfgang Huber, in the davidTiling R package (LGPL license)

**See Also**

[gffRead](#) for creating a data frame from a GTF/GFF file, and <http://useast.ensembl.org/info/website/upload/gff.html> for specifics of the GFF/GTF file format.

**Examples**

```
gtfPath = system.file('extdata', 'annot.gtf.gz', package='ballgown')
gffdata = gffRead(gtfPath)
gffdata$transcriptID = getAttributeField(gffdata$attributes,
  field = "transcript_id")
```

---

getGenes

*label assembled transcripts with gene names*


---

**Description**

label assembled transcripts with gene names

**Usage**

```
getGenes(gtf, assembled, UCSC = TRUE, attribute = "gene_id")
```

**Arguments**

gtf	path to a GTF file containing locations of annotated transcripts
assembled	GRangesList object, with each set of ranges representing exons of an assembled transcript.
UCSC	set to TRUE if you're using a UCSC gtf file. (Requires some extra text processing).
attribute	set to attribute name in gtf that gives desired gene identifiers. Default "gene_id"; another common one is "gene_name" (for the gene symbol).

**Details**

chromosome labels in gtf and assembled should match. (i.e., you should provide the path to a gtf corresponding to the same annotation you used when constructing assembled)

**Value**

an IRanges CharacterList of the same length as assembled, providing the name(s) of the gene(s) that overlaps each transcript in assembled.

**Author(s)**

Alyssa Frazee, Andrew Jaffe

**Examples**

```
data(bg)
gtfPath = system.file('extdata', 'annot.gtf.gz', package='ballgown')
geneoverlaps = getGenes(gtfPath, structure(bg)$trans, UCSC=FALSE)
```

---

gexpr	<i>extract gene-level expression measurements from ballgown objects</i>
-------	---

---

**Description**

For objects created with Cufflinks/Tablemaker, gene-level measurements are calculated by appropriately combining FPKMs from the transcripts comprising the gene. For objects created with RSEM, gene-level measurements are extracted directly from the RSEM output.

**Usage**

```
gexpr(x)

## S4 method for signature 'ballgown'
gexpr(x)
```

**Arguments**

x                    a ballgown object

**Value**

gene-by-sample matrix containing per-sample gene measurements.

**Examples**

```
data(bg)
gene_matrix = gexpr(bg)
```

---

gffRead	<i>read in GTF/GFF file as a data frame</i>
---------	---

---

**Description**

read in GTF/GFF file as a data frame

**Usage**

```
gffRead(gffFile, nrows = -1, verbose = FALSE)
```

**Arguments**

gffFile            name of GTF/GFF on disk  
 nrows             number of rows to read in (default -1, which means read all rows)  
 verbose            if TRUE, print status info at beginning and end of file read. Default FALSE.

**Value**

data frame representing the GTF/GFF file

**Author(s)**

Kasper Hansen

**See Also**

[getAttributeField](http://useast.ensembl.org/info/website/upload/gff.html) to extract data from "attributes" column; <http://useast.ensembl.org/info/website/upload/gff.html> for more information on the GTF/GFF file format.

**Examples**

```
gtfPath = system.file('extdata', 'annot.gtf.gz', package='ballgown')
annot = gffRead(gtfPath)
```

---

gffReadGR	<i>read in gtf file as GRanges object</i>
-----------	---

---

**Description**

(very) light wrapper for `rtracklayer::import`

**Usage**

```
gffReadGR(
  gtf,
  splitByTranscript = FALSE,
  identifier = "transcript_id",
  sep = "; "
)
```

**Arguments**

gtf                name of GTF/GFF file on disk  
 splitByTranscript    if TRUE, return a `GRangesList` of transcripts; otherwise return a `GRanges` object containing all genomic features in `gtf`. Default FALSE.  
 identifier        name of transcript identifier column of `attributes` field in `gtf`. Default "transcript\_id". Only used if `splitByTranscript` is TRUE.  
 sep                field separator in the `attributes` field of `gtf`. Default "; " (semicolon + space). Only used if `splitByTranscript` is TRUE.

**Value**

if `splitByTranscript` is `FALSE`, an object of class `GRanges` representing the genomic features in `gtf`. If `splitByTranscript` is `TRUE`, an object of class `GRangesList`, where each element is a `GRanges` object corresponding to an annotated transcript (designated in names).

**Author(s)**

Alyssa Frazee

**See Also**

[gffRead](#) for reading in a GTF file as a data frame rather than a `GRanges/GRangesList` object.

**Examples**

```
gtfPath = system.file('extdata', 'annot.gtf.gz', package='ballgown')

# read in exons as GRanges:
annotgr = gffReadGR(gtfPath)

# read in groups of exons as transcripts, in GRangesList:
transcripts_grl = gffReadGR(gtfPath, splitByTranscript=TRUE)
```

---

iexpr	<i>extract transcript-level expression measurements from ballgown objects</i>
-------	---

---

**Description**

extract transcript-level expression measurements from ballgown objects

**Usage**

```
iexpr(x, meas = "rcount")

## S4 method for signature 'ballgown'
iexpr(x, meas = "rcount")
```

**Arguments**

x	a ballgown object
meas	type of measurement to extract. Can be "rcount", "ucount", "mrcount", or "all". Default "rcount".

**Value**

intron-by-sample matrix containing the number of reads (measured as specified by `meas`) supporting each intron, in each sample. If `meas` is "all", a data frame is returned, containing all measurements and location information.

**Examples**

```
data(bg)
intron_rcount_matrix = iexpr(bg)
intron_data_frame = iexpr(bg, 'all')
```

---

indexes	<i>extract the indexes from ballgown objects</i>
---------	--

---

**Description**

extract the indexes from ballgown objects

**Usage**

```
indexes(x)

## S4 method for signature 'ballgown'
indexes(x)
```

**Arguments**

x                    a ballgown object

**Value**

list containing elements e2t, i2t, t2g, bamfiles, and pData, where e2t and i2t are data frames linking exons and introns (respectively) to transcripts, t2g is a data frame linking transcripts to genes, and bamfiles and pData are described in ?ballgown.

**Examples**

```
data(bg)
names(indexes(bg))
class(indexes(bg))
head(indexes(bg)$t2g)
```

---

indexes<-	<i>Replace method for indexes slot in ballgown objects</i>
-----------	--

---

**Description**

Replace method for indexes slot in ballgown objects

**Usage**

```
indexes(x) <- value

## S4 replacement method for signature 'ballgown'
indexes(x) <- value
```

**Arguments**

x                    a ballgown object  
value                the updated value for `indexes(x)` or a subcomponent

**Examples**

```
data(bg)  
indexes(bg)$bamfiles = paste0('/path/to/bamfolder/',  
  sampleNames(bg), '_accepted_hits.bam')
```

---

last	<i>get the last element</i>
------	-----------------------------

---

**Description**

get the last element

**Usage**

```
last(x)
```

**Arguments**

x                    anything you can call `tail` on (vector, data frame, etc.)

**Details**

this function is made of several thousand lines of complex code, so be sure to read it carefully.

**Value**

the last element of x

**Author(s)**

Alyssa Frazee

**Examples**

```
last(c('h', 'e', 'l', 'l', 'o'))
```

---

mergedDate	<i>extract package version &amp; creation date from ballgown object</i>
------------	---

---

**Description**

extract package version & creation date from ballgown object

**Usage**

```
mergedDate(x)
```

```
## S4 method for signature 'ballgown'  
mergedDate(x)
```

**Arguments**

x	a ballgown object
---	-------------------

**Examples**

```
data(bg)  
mergedDate(bg)
```

---

pctOverlap	<i>calculate percent overlap between two GRanges objects</i>
------------	--

---

**Description**

calculate percent overlap between two GRanges objects

**Usage**

```
pctOverlap(tx1, tx2)
```

**Arguments**

tx1	GRanges object
tx2	GRanges object

**Details**

In the ballgown context, tx1 and tx2 are two transcripts, each represented by GRanges objects whose ranges represent the exons comprising the transcripts. The percent overlap is the number of nucleotides falling within both transcripts divided by the number of nucleotides falling within either transcript. Useful as a measure of transcript closeness (as it is essentially Jaccard distance).

**Value**

percent overlap between tx1 and tx2, as defined by the ratio of the intersection of tx1 and tx2 to the union of tx1 and tx2.

**Author(s)**

Alyssa Frazee

**Examples**

```
data(bg)
gtfPath = system.file('extdata', 'annot.gtf.gz', package='ballgown')
annot_gr1 = gffReadGR(gtfPath, splitByTranscript=TRUE)
pctOverlap(structure(bg)$trans[[2]], annot_gr1[[369]]) #79.9%
```

---

pData	<i>extract phenotype data from a ballgown object</i>
-------	--

---

**Description**

extract phenotype data from a ballgown object

**Usage**

```
pData(object)

## S4 method for signature 'ballgown'
pData(object)
```

**Arguments**

object            a ballgown object

**Value**

sample-by-phenotype data frame

**Examples**

```
data(bg)
pData(bg)
```

---

pData<-	<i>Replacement method for pData slot in ballgown objects</i>
---------	--

---

**Description**

Replacement method for pData slot in ballgown objects

**Usage**

```
pData(object) <- value

## S4 replacement method for signature 'ballgown,ANY'
pData(object) <- value
```



**Arguments**

object	a ballgown object
value	the updated value for pData(x).

**Examples**

```
# add "timepoint" covariate to ballgown object:
data(bg) # already contains pData
pData(bg) = data.frame(pData(bg), timepoint=rep(1:10, 2))
head(pData(bg))
```

---

plotLatentTranscripts *cluster assembled transcripts and plot the results*

---

**Description**

This is an experimental, first-pass function that clusters assembled transcripts based on their overlap percentage, then plots and colors the transcript clusters.

**Usage**

```
plotLatentTranscripts(
  gene,
  gown,
  method = c("hclust", "kmeans"),
  k = NULL,
  choosek = c("var90", "thumb"),
  returncluster = TRUE,
  labelTranscripts = TRUE,
  ...
)
```

**Arguments**

gene	string, name of gene whose transcripts should be clustered (e.g., "XLOC_000001")
gown	object of class ballgown being used for analysis
method	clustering method to use. Currently can choose from hierarchical clustering (hclust) or K-means (kmeans). More methods are in development.
k	number of transcripts clusters to use. By default, k is NULL and thus is chosen using a rule of thumb, but providing k overrides those rules of thumb.
choosek	if k is not provided, how should the number of clusters be chosen? Must be one of "var90" (choose a k that explains 90 percent of the observed variation) or "thumb" (k is set to be approximately $\sqrt{n}$ , where n is the total number of transcripts for gene)
returncluster	if TRUE (as it is by default), return the results of the call to clusterTranscripts so the data is available for later use. Nothing is returned if FALSE.
labelTranscripts	if TRUE (as it is by default), print transcript IDs on the y-axis
...	other arguments to pass to plotTranscripts

**Value**

if `returncluster` is `TRUE`, the transcript clusters are returned as described in [clusterTranscripts](#). A plot of the transcript clusters is also produced, in the style of [plotTranscripts](#).

**Author(s)**

Alyssa Frazee

**See Also**

[clusterTranscripts](#), [plotTranscripts](#)

**Examples**

```
data(bg)
plotLatentTranscripts('XLOC_000454', bg, method='kmeans', k=2)
```

---

plotMeans

*visualize transcript abundance by group*

---

**Description**

visualize transcript abundance by group

**Usage**

```
plotMeans(
  gene,
  gown,
  overall = FALSE,
  groupvar,
  groupname = "all",
  meas = c("cov", "FPKM", "rcount", "ucount", "mrcount", "mcov"),
  colorby = c("transcript", "exon"),
  legend = TRUE,
  labelTranscripts = FALSE
)
```

**Arguments**

gene	name of gene whose transcripts will be plotted. When using Cufflinks/Tablemaker output, usually of the form "XLOC_#####"
gown	ballgown object containing experimental and phenotype data
overall	if <code>TRUE</code> , color features by the overall (experiment-wide) mean rather than a group-specific mean
groupvar	string representing the name of the variable denoting which sample belongs to which group. Can be "none" (if you want the study-wide mean), or must correspond to the name of a column of <code>pData(gown)</code> . Usually a categorical variable.

groupname	string representing which group's expression means you want to plot. Can be "none" (if you want the study-wide mean), "all" (if you want a multipanel plot of each group's mean expression), or any of the levels of groupvar.
meas	type of expression measurement to plot. One of "cov", "FPKM", "rcount", "ucount", "mrcount", or "mcov". Not all types are valid for all features. (See description of tablemaker output for more information).
colorby	one of "transcript" or "exon", indicating which feature's abundances should dictate plot coloring.
legend	if TRUE (as it is by default), a color legend is drawn on top of the plot indicating the scale for feature abundances.
labelTranscripts	if TRUE, transcript ids are labeled on the left side of the plot. Default FALSE.

**Value**

produces a plot of the transcript structure for the specified gene in the current graphics device, colored by study-wide or group-specific mean expression level.

**Author(s)**

Alyssa Frazee

**See Also**

[plotTranscripts](#)

**Examples**

```
data(bg)
plotMeans('XLOC_000454', bg, groupvar='group', meas='FPKM',
          colorby='transcript')
```

---

plotTranscripts	<i>visualize structure of assembled transcripts</i>
-----------------	---

---

**Description**

visualize structure of assembled transcripts

**Usage**

```
plotTranscripts(
  gene,
  gown,
  samples = NULL,
  colorby = "transcript",
  meas = "FPKM",
  legend = TRUE,
  labelTranscripts = FALSE,
  main = NULL,
```

```

blackBorders = TRUE,
log = FALSE,
logbase = 2,
customCol = NULL,
customOrder = NULL
)

```

### Arguments

gene	name of gene whose transcripts will be plotted. When using Cufflinks output, usually of the form "XLOC_#####"
gown	ballgown object containing experimental and phenotype data
samples	vector of sample(s) to plot. Can be 'none' if only one plot (showing transcript structure in gray) is desired. Use sampleNames(gown) to see sample names for gown. Defaults to sampleNames(gown)[1].
colorby	one of "transcript", "exon", or "none", indicating which feature's abundances should dictate plot coloring. If "none", all transcripts are drawn in gray.
meas	which expression measurement to color features by, if any. Must match an available measurement for whatever feature you're plotting.
legend	if TRUE (as it is by default), a color legend is drawn on top of the plot indicating scales for feature abundances.
labelTranscripts	if TRUE, transcript ids are labeled on the left side of the plot. Default FALSE.
main	optional string giving the desired plot title.
blackBorders	if TRUE, exon borders are drawn in black. Otherwise, they are drawn in the same color as their transcript or exon. Switching blackBorders to FALSE can be useful for visualizing abundances for skinny exons and/or smaller plots, which can be the case when length(samples) is large.
log	if TRUE, color transcripts on the log scale. Default FALSE. To account for expression values of 0, we add 1 to all expression values before taking the log.
logbase	log base to use if log = TRUE. Default 2.
customCol	an optional vector of custom colors to color transcripts by. There must be the same number of colors as transcripts in the gene being plotted.
customOrder	an optional vector of transcript ids (matching ids in texpr(gown, 'all')\$t_id), indicating which order transcripts will appear in the plot. All transcripts in gene must appear in the vector exactly once.

### Value

produces a plot of the transcript structure for the specified gene in the current graphics device.

### Author(s)

Alyssa Frazee

### See Also

[plotMeans](#), [plotLatentTranscripts](#)

**Examples**

```
data(bg)

# plot one gene for one sample:
plotTranscripts(gene='XLOC_000454', gown=bg, samples='sample12', meas='FPKM',
  colorby='transcript',
  main='transcripts from gene XLOC_000454: sample 12, FPKM')

# plot one gene for many samples:
plotTranscripts('XLOC_000454', bg,
  samples=c('sample01', 'sample06', 'sample12', 'sample19'),
  meas='FPKM', colorby='transcript')
```

---

sampleNames	<i>get names of samples in a ballgown objects</i>
-------------	---

---

**Description**

get names of samples in a ballgown objects

**Usage**

```
sampleNames(object)

## S4 method for signature 'ballgown'
sampleNames(object)
```

**Arguments**

object            a ballgown object

**Value**

vector of sample IDs for x. If pData exists, samples in its rows correspond to samples in sampleNames(x) (in order).

**Examples**

```
data(bg)
sampleNames(bg)
```

---

seqnames	<i>get sequence (chromosome) names from ballgown object</i>
----------	---

---

### Description

get sequence (chromosome) names from ballgown object

### Usage

```
seqnames(x)
```

```
## S4 method for signature 'ballgown'  
seqnames(x)
```

### Arguments

x                    a ballgown object

### Value

vector of sequence (i.e., chromosome) names included in the ballgown object

### Examples

```
data(bg)  
seqnames(bg)
```

---

stattest	<i>statistical tests for differential expression in ballgown</i>
----------	--

---

### Description

Test each transcript, gene, exon, or intron in a ballgown object for differential expression, using comparisons of linear models.

### Usage

```
stattest(  
  gown = NULL,  
  gowntable = NULL,  
  pData = NULL,  
  mod = NULL,  
  mod0 = NULL,  
  feature = c("gene", "exon", "intron", "transcript"),  
  meas = c("cov", "FPKM", "rcount", "ucount", "mrcount", "mcov"),  
  timecourse = FALSE,  
  covariate = NULL,  
  adjustvars = NULL,  
  gexpr = NULL,  
  df = 4,  
)
```

```

    getFC = FALSE,
    libadjust = NULL,
    log = TRUE
  )

```

### Arguments

<code>gown</code>	name of an object of class <code>ballgown</code>
<code>gowntable</code>	matrix or matrix-like object with rownames representing feature IDs and columns representing samples, with expression estimates in the cells. Provide the feature name with <code>feature</code> . You must provide exactly one of <code>gown</code> or <code>gowntable</code> . NB: <code>gowntable</code> is log-transformed within <code>stattest</code> if <code>log</code> is <code>TRUE</code> , so provide unlogged expression values in <code>gowntable</code> .
<code>pData</code>	Required if <code>gowntable</code> is provided: data frame giving phenotype data for the samples in the columns of <code>gowntable</code> . (Rows of <code>pData</code> correspond to columns of <code>gowntable</code> ). If <code>gown</code> is used instead, it must have a non-null, valid <code>pData</code> slot (and the <code>pData</code> argument to <code>stattest</code> should be left <code>NULL</code> ).
<code>mod</code>	object of class <code>model.matrix</code> representing the design matrix for the linear regression model including covariates of interest
<code>mod0</code>	object of class <code>model.matrix</code> representing the design matrix for the linear regression model without the covariates of interest.
<code>feature</code>	the type of genomic feature to be tested for differential expression. If <code>gown</code> is used, must be one of "gene", "transcript", "exon", or "intron". If <code>gowntable</code> is used, this is just used for labeling and can be whatever the rows of <code>gowntable</code> represent.
<code>meas</code>	the expression measurement to use for statistical tests. Must be one of "cov", "FPKM", "rcount", "ucount", "mrcount", or "mcov". Not all expression measurements are available for all features. Leave as default if <code>gowntable</code> is provided.
<code>timecourse</code>	if <code>TRUE</code> , tests whether or not the expression profiles of genomic features vary over time (or another continuous covariate) in the study. Default <code>FALSE</code> . Natural splines are used to fit time profiles, so you must have more timepoints than degrees of freedom used to fit the splines. The default <code>df</code> is 4.
<code>covariate</code>	string representing the name of the covariate of interest for the differential expression tests. Must correspond to the name of a column of <code>pData(gown)</code> . If <code>timecourse=TRUE</code> , this should be the study's time variable.
<code>adjustvars</code>	optional vector of strings representing the names of potential confounders. Must correspond to names of columns of <code>pData(gown)</code> .
<code>gexpr</code>	optional data frame that is the result of calling <code>gexpr(gown)</code> . (You can speed this function up by pre-creating <code>gexpr(gown)</code> .)
<code>df</code>	degrees of freedom used for modeling expression over time with natural cubic splines. Default 4. Only used if <code>timecourse=TRUE</code> .
<code>getFC</code>	if <code>TRUE</code> , also return estimated fold changes (adjusted for library size and confounders) between populations. Only available for 2-group comparisons at the moment. Default <code>FALSE</code> .
<code>libadjust</code>	library-size adjustment to use in linear models. By default, the adjustment is defined as the sum of the sample's log expression measurements below the 75th percentile of those measurements. To use a different library-size adjustment, provide a numeric vector of each sample's adjustment value. Entries of this

vector correspond to samples in in rows of pData. If no library size adjustment is desired, set to FALSE.

log if TRUE, outcome variable in linear models is  $\log(\text{expression}+1)$ , otherwise it's expression. Default TRUE.

## Details

At minimum, you need to provide a ballgown object or count table, the type of feature you want to test (gene, transcript, exon, or intron), the expression measurement you want to use (FPKM, cov, rcount, etc.), and the covariate of interest, which must be the name of one of the columns of the 'pData' component of your ballgown object (or provided pData). This covariate is automatically converted to a factor during model fitting in non-timecourse experiments.

By default, models are fit using  $\log_2(\text{meas} + 1)$  as the outcome for each feature. To disable the log transformation, provide 'log = FALSE' as an argument to 'stattest'. You can use the `gowntable` option if you'd like to use a different transformation.

Library size adjustment is performed by default by using the sum of the log nonzero expression measurements for each sample, up to the 75th percentile of those measurements. This adjustment can be disabled by setting `libadjust=FALSE`. You can use `mod` and `mod0` to specify alternative library size adjustments.

`mod` and `mod0` are optional arguments. If `mod` is specified, you must also specify `mod0`. If neither is specified, `mod0` defaults to the design matrix for a model including only a library-size adjustment, and `mod` defaults to the design matrix for a model including a library-size adjustment and covariate. Note that if you supply `mod` and `mod0`, `covariate`, `timecourse`, `adjustvars`, and `df` are ignored, so make sure your covariate of interest and all appropriate confounder adjustments, including library size, are specified in `mod` and `mod0`. By default, the library-size adjustment is the sum of all counts below the 75th percentile of nonzero counts, on the log scale ( $\log_2 + 1$ ).

Full model details are described in the supplement of <http://biorxiv.org/content/early/2014/03/30/003665>.

## Value

data frame containing the columns `feature`, `id` representing feature id, `pval` representing the p-value for testing whether this feature was differentially expressed according to `covariate`, and `qval`, the estimated false discovery rate using this feature's signal strength as a significance cutoff. An additional column, `fc`, is included if `getFC` is TRUE.

## Author(s)

Jeff Leek, Alyssa Frazee

## References

<http://biorxiv.org/content/early/2014/03/30/003665>

## Examples

```
data(bg)

# two-group comparison:
stat_results = stattest(bg, feature='transcript', meas='FPKM',
  covariate='group')

# timecourse test:
```



```

pData(bg) = data.frame(pData(bg), time=rep(1:10, 2)) #dummy time covariate
timecourse_results = stattest(bg, feature='transcript', meas='FPKM',
  covariate='time', timecourse=TRUE)

# timecourse test, adjusting for group:
group_adj_timecourse_results = stattest(bg, feature='transcript',
  meas='FPKM', covariate='time', timecourse=TRUE, adjustvars='group')

# custom model matrices:
### create example data:
set.seed(43)
sex = sample(c('M', 'F'), size=nrow(pData(bg)), replace=TRUE)
age = sample(21:52, size=nrow(pData(bg)), replace=TRUE)

### create design matrices:
mod = model.matrix(~ sex + age + pData(bg)$group + pData(bg)$time)
mod0 = model.matrix(~ pData(bg)$group + pData(bg)$time)

### build model:
adjusted_results = stattest(bg, feature='transcript', meas='FPKM',
  mod0=mod0, mod=mod)

```

---

structure

*extract structure components from ballgown objects*


---

## Description

extract structure components from ballgown objects

## Usage

```

structure(x)

## S4 method for signature 'ballgown'
structure(x)

```

## Arguments

x                    a ballgown object

## Value

list containing elements intron, exon, and trans. exon and intron are GRanges objects, where each range is an exon or intron, and trans is a GRangesList object, where each GRanges element is a set of exons representing a transcript.

## Examples

```

data(bg)
names(structure(bg))
class(structure(bg))
structure(bg)$exon

```

---

subset                      *subset ballgown objects to specific samples or genomic locations*

---

### Description

subset ballgown objects to specific samples or genomic locations

### Usage

```
subset(x, ...)

## S4 method for signature 'ballgown'
subset(x, cond, genomesubset = TRUE)
```

### Arguments

x	a ballgown object
...	further arguments to generic subset
cond	Condition on which to subset. See details.
genomesubset	if TRUE, subset x to a specific part of the genome. Otherwise, subset x to only include specific samples. TRUE by default.

### Details

To use subset, you must provide the cond argument as a string representing a logical expression specifying your desired subset. The subset expression can either involve column names of `texpr(x, "all")` (if `genomesubset` is TRUE) or of `pData(x)` (if `genomesubset` is FALSE). For example, if you wanted a ballgown object for only chromosome 22, you might call `subset(x, "chr == 'chr22'")`. (Be sure to handle quotes within character strings appropriately).

### Value

a subsetted ballgown object, containing only the regions or samples satisfying cond.

### Author(s)

Alyssa Frazee

### Examples

```
data(bg)
bg_twogenes = subset(bg, "gene_id=='XLOC_000454' | gene_id=='XLOC_000024'")
bg_twogenes
# ballgown instance with 4 assembled transcripts and 20 samples

bg_group0 = subset(bg, "group == 0", genomesubset=FALSE)
bg_group0
# ballgown instance with 100 assembled transcripts and 10 samples
```

---

texpr	<i>extract transcript-level expression measurements from ballgown objects</i>
-------	---

---

### Description

extract transcript-level expression measurements from ballgown objects

### Usage

```
texpr(x, meas = "FPKM")

## S4 method for signature 'ballgown'
texpr(x, meas = "FPKM")
```

### Arguments

x	a ballgown object
meas	type of measurement to extract. Can be "cov", "FPKM", or "all". Default "FPKM".

### Value

transcript-by-sample matrix containing expression values (measured by meas). If meas is "all", a data frame is returned, containing all measurements and location information.

### Examples

```
data(bg)
transcript_fpkm_matrix = texpr(bg)
transcript_data_frame = texpr(bg, 'all')
```

---

tGene	<i>Connect a transcript to its gene</i>
-------	---

---

### Description

find the gene to which a transcript belongs

### Usage

```
tGene(bg, transcript, tid = TRUE, gid = TRUE, warnme = TRUE)
```

**Arguments**

bg	ballgown object
transcript	transcript identifier
tid	set to TRUE if transcript is a numeric transcript identifier (i.e., t_id in expression tables), or FALSE if transcript is a named identifier (e.g., TCONS_000001 or similar).
gid	if FALSE, return the gene *name* associated with transcript in bg instead of the gene *id*, which is returned by default. Take care to remember that not all ballgown objects include gene *name* information. (They do all include gene IDs).
warnme	if TRUE, and if gid is FALSE, print a warning if no gene name is available for the transcript. This could either mean the transcript didn't overlap an annotated gene, or that no gene names were included when bg was created.

**Examples**

```
data(bg)
tGene(bg, 10)
tGene(bg, 'TCONS_00000010', tid=FALSE)
tGene(bg, 10, gid=FALSE) #empty: no gene names included in bg.
```

---

transcriptIDs	<i>get numeric transcript IDs from a ballgown object</i>
---------------	--

---

**Description**

get numeric transcript IDs from a ballgown object

**Usage**

```
transcriptIDs(x)

## S4 method for signature 'ballgown'
transcriptIDs(x)
```

**Arguments**

x	a ballgown object
---	-------------------

**Value**

vector of numeric transcript IDs included in the ballgown object

**Examples**

```
data(bg)
transcriptIDs(bg)
```

---

transcriptNames	<i>get transcript names from a ballgown object</i>
-----------------	--

---

**Description**

get transcript names from a ballgown object

**Usage**

```
transcriptNames(x)
```

```
## S4 method for signature 'ballgown'  
transcriptNames(x)
```

**Arguments**

x                    a ballgown object

**Value**

vector of transcript names included in the ballgown object. If object was created using Cufflinks/Tablemaker, these transcript names will be of the form "TCONS\_\*". Return vector is named and ordered by corresponding numeric transcript ID.

**Examples**

```
data(bg)  
transcriptNames(bg)
```

---

writeFiles	<i>write files to disk from ballgown object</i>
------------	---

---

**Description**

create tablemaker-like files on disk from a ballgown object

**Usage**

```
writeFiles(gown, dataDir)
```

**Arguments**

gown                ballgown object  
dataDir            top-level directory for sample-specific folders

**Examples**

```
data(bg)  
writeFiles(bg, dataDir=getwd())
```

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