

Package ‘altcdfenvs’

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Title alternative CDF environments (aka probeset mappings)

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Description Convenience data structures and functions to handle
cdfenvs

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Collate appendCdfEnvAffy.R buildCdfEnv.matchprobes.R
buildCdfEnv.biostrings.R CdfEnv.R cdfenvs.R copyCdfEnvAffy.R
readFASTA.R removeIndex.R unique.CdfEnvAffy.R

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AffyProbesMatch-class *Class "AffyProbesMatch"*

Description

Store the results of a call to `matchAffyProbes`.

Objects from the Class

Objects can be created by calls of the form `new("AffyProbesMatch", ...)`.

An object will store the result of matching probe sequences against target sequences.

Slots

`pm`: Object of class "list": each element is vector of index values

`mm`: Object of class "list": each element is vector of index values

`labels`: Object of class "character"

`chip_type`: Object of class "character" and of length 1.

`probes`: Object of class "ANY": the probetable object used to perform the matches.

Methods

combine signature(`x = "AffyProbesMatch"`, `y = "AffyProbesMatch"`): combine two instances.

This is can be useful when splitting the list of target sequences to parallelized the job.

show signature(`x = "AffyProbesMatch"`): Show the instance.

toHypergraph signature(`object = "AffyProbesMatch"`): build an [Hypergraph](#) from the matches.

Examples

```
showClass("AffyProbesMatch")
```

| | |
|------------------|--|
| appendCdfEnvAffy | <i>append probe sets to a CdfEnvAffy</i> |
|------------------|--|

Description

append probe sets to a CdfEnvAffy

Usage

```
appendCdfEnvAffy(acdfenv, id, i, nocopy = TRUE)
```

Arguments

| | |
|---------|--|
| acdfenv | instance of class CdfEnvAffy |
| id | identifier for the probe set to add |
| i | a matrix of indexes (see details) |
| nocopy | whether to make a copy of the environment or not (see details) |

Details

The matrix `i` must have one column per probe type. For typical Affymetrix chip types, there are two probe types: "pm" and "mm".

`nocopy` set to TRUE means that the environment is added the probe set 'in-situ' (this can boost execution speed if you add a lot of probe sets).

Value

An CdfEnvAffy is returned

Examples

```
data(cdfenvEx)

## pm and mm probe set
m <- matrix(1:10, ncol = 2)
colnames(m) <- c("pm", "mm")

appendCdfEnvAffy(cdfenvEx, "blabla", m)

indexProbes(cdfenvEx, c("pm", "mm"), "blabla")

## pm only probe set
m <- matrix(6:9, ncol = 1)
colnames(m) <- c("pm")
```

```
appendCdfEnvAffy(cdfenvEx, "blabla2", m)
## note that the unspecified "mm" were set to NA
indexProbes(cdfenvEx, c("pm", "mm"), "blabla2")
```

buildCdfEnv.biostrings

Build CDF environments

Description

Build CDF environment from Biostrings matchPDict results

Usage

```
buildCdfEnv.biostrings(apm, abatch = NULL,
                       nrow.chip = NULL, ncol.chip = NULL,
                       simplify = TRUE,
                       x.colname = "x", y.colname = "y",
                       verbose = FALSE)
```

Arguments

| | |
|-----------|--|
| apm | AffyProbesMatch |
| abatch | AffyBatch |
| nrow.chip | number of rows for the chip type (see details) |
| ncol.chip | number of columns for the chip type (see details) |
| simplify | simplify the environment built (removing target names when there is no matching probe) |
| x.colname | column name |
| y.colname | column name |
| verbose | verbose TRUE/FALSE |

Details

Whenever an abatch is specified, nrow.chip and ncol.chip are not needed. Specifying the an AffyBatch in abatch is the easiest way to specify information about the geometry of a chip type.

Value

An instance of class CdfEnvAffy.

| | |
|------------------|--------------------|
| CdfEnvAffy-class | Class "CdfEnvAffy" |
|------------------|--------------------|

Description

A class to hold the information necessary to handle the grouping of probes in set of probes, and to find XY coordinates of probes on a chip

Objects from the Class

Objects can be created by calls of the form `new("CdfEnvAffy", ...)`. Typically, there is an instance of the class for each type of chip (e.g. Hu6800, HG-U95A, etc...).

Slots

envir: Object of class "environment". It has to be thought of as a hashtable: the keys are probe set identifiers, or gene names, and the values are indexes.

envName: Object of class "character". A name for the environment.

index2xy: Object of class "function". The function used to resolve index into xy coordinates. Unless you are an advanced user, you probably want to ignore this (and rely on the default provided with the package).

xy2index: Object of class "function". The function used to resolve xy coordinates into index. Unless you are an advanced user, you probably want to ignore this (and rely on the default provided with the package).

nrow: Object of class "integer". The number of rows of probes for the chip type.

ncol: Object of class "integer". The number of columns of probes for the chip type.

probeTypes: Object of class "character". The different types of probes stored for each probe set. In the case of Affymetrix chips, the probes are typically perfect match (pm) probes or mismatch probes (mm).

chipType: Object of class "character". The name of the chip type the instance is associated with. This is useful when one starts to create alternative mappings of the probes on a chip (see associated vignette).

Methods

[**signature**(object = "CdfEnvAffy", i = "character", j = "missing", drop = "boolean"): subset a cdf, that is return a new cdf containing only a subset of the probe sets. The subset of probe sets to take is identified as a vector of identifiers (mode "character").

coerce **signature**(object = "CdfEnvAffy", "environment"): coerce an instance of the class to an environment.

coerce **signature**(object = "CdfEnvAffy", "Cdf"): coerce an instance of the class to a Cdf.

geneNames **signature**(object="CdfEnvAffy"): Return the names of the known probe sets (of course, it depends on the associated CDF).

index2xy signature(object = "CdfEnvAffy", i="integer"): convert index values into XY coordinates.

indexProbes signature(object = "CdfEnvAffy", which = "character", probeSetNames = NULL): obtain the indexes for the probes associated with the probe set name probeSetNames. When probeSetNames is set to NULL (default), the indexes are returned for the probe sets defined on the chip. See indexProbes.CdfEnvAffy

plot signature(x = "CdfEnvAffy", y = "missing"): Plot the chip. It mainly sets coordinates for further plotting (see examples). See plot.CdfEnvAffy

show signature(object = "CdfEnvAffy"): Print method.

xy2index signature(object = "CdfEnvAffy", x="integer", y="integer"): convert XY coordinates into index values.

toHypergraph signature(object = "CdfEnvAffy"): convert XY coordinates into index values.

Author(s)

Laurent Gautier

See Also

[indexProbes.CdfEnvAffy](#), [plot.CdfEnvAffy](#)

Examples

```
## build an instance
library(hgu95acdf)
cdfenv.hgu95a <- wrapCdfEnvAffy(hgu95acdf, 640, 640, "HG-U95A")

show(cdfenv.hgu95a)

## find the indexes for a probe set (pm only)
ip <- indexProbes(cdfenv.hgu95a, "pm", "1000_at")[[1]]
## get the XY coordinates for the probe set
xy <- index2xy(cdfenv.hgu95a, ip)

## plot the chip
plot(cdfenv.hgu95a)

## plot the coordinates
plotLocation(xy)

## subset the environment

cdfenv.hgu95a.mini <- cdfenv.hgu95a["1000_at"]
```

`cdfenvEx`*CdfEnvAffy*

Description

An example of CdfEnvAffy

Usage

```
data(cdfenvEx)
```

Format

The format is: Formal class 'CdfEnvAffy' [package "altcdfenvs"] with 8 slots ..@ index2xy :function (object, i) ..@ xy2index :function (object, x, y) ..@ envir :length 2 <environment> ..@ envName : chr "ZG-DU33" ..@ nrow : int 100 ..@ ncol : int 100 ..@ probeTypes: chr [1:2] "pm" "mm" ..@ chipType : chr "ZG-DU33"

Examples

```
data(cdfenvEx)
print(cdfenvEx)
```

`cdfenvs`*functions related to cdfenvs*

Description

A set of functions to handle cdfenvs

Usage

```
wrapCdfEnvAffy(cdfenv, nrow.chip, ncol.chip, chiptype, check = TRUE,
               verbose = FALSE)
getCdfEnvAffy(abatch)
buildCdfEnv.matchprobes(matches, ids, probes.pack, abatch=NULL,
                        nrow.chip=NULL, ncol.chip=NULL, chiptype=NULL,
                        mm=NA, simplify = TRUE,
                        x.colname = "x", y.colname = "y", verbose=FALSE)
```

Arguments

| | |
|----------------------|---|
| abatch | an AffyBatch |
| cdfenv | A cdfenv environment |
| check | perform consistency check or not |
| chiptype | A name for the chip type |
| ids | a vector of probe set identifiers for the matches |
| matches | a list as returned by the function combineAffyBatch |
| mm | The value to store for MMs |
| ncol.chip | The number of columns for the chip type |
| nrow.chip | The number of rows for the chip type |
| probes.pack | The name of the probe package |
| simplify | Simplify the environment created by removing the ids without any matching probe |
| x.colname, y.colname | see the getxy.probeseq |
| verbose | verbosity (TRUE or FALSE) |

Value

An instance of class CdfEnvAffy.

Examples

```
## See the main vignette
```

| | |
|----------------|------------------------------------|
| copyCdfEnvAffy | <i>make a copy of a CdfEnvAffy</i> |
|----------------|------------------------------------|

Description

make a copy of a CdfEnvAffy

Usage

```
copyCdfEnvAffy(acdfenv)
```

Arguments

| | |
|---------|------------------------------|
| acdfenv | instance of class CdfEnvAffy |
|---------|------------------------------|

Details

Make a copy can be needed since a CdfEnvAffy contains an environment

Value

A CdfEnvAffy

See Also

[CdfEnvAffy-class](#), [copyEnv](#)

| | |
|-----------------|--|
| countduplicated | <i>Count the number of times probes are used</i> |
|-----------------|--|

Description

This function counts the number of times the probes in a CdfEnvAffy are found in this object.

Usage

```
countduplicated(x, incomparables = FALSE, verbose = FALSE)
```

Arguments

| | |
|---------------|----------------------------------|
| x | An instance of CdfEnvAffy-class |
| incomparables | (not implemented yet, keep away) |
| verbose | verbose or not |

Value

An environment is returned. Each element in this environment has the same identifier than its corresponding probe set in the CdfEnvAffy-class and contains the number of times a probe is in use in the environment (instead of an index number in the CdfEnvAffy-class).

Author(s)

Laurent

See Also

[CdfEnvAffy-class](#)

geneNames.CdfEnvAffy *get the names of the known probe sets*

Description

get the names of the probe sets known to the CdfEnv

Usage

```
geneNames.CdfEnvAffy(object)
```

Arguments

object CdfEnvAffy-class

Value

a vector of mode character

getxy.probeseq *A function to get the XY coordinates from a probes sequences data frame*

Description

A function to get the XY coordinates from a probes sequences data.frame

Usage

```
getxy.probeseq(ppset.id = NULL, probeseq = NULL, i.row = NULL,
xy.offset = NULL, x.colname = "x", y.colname = "y")
```

Arguments

ppset.id The probe sets of interest (a vector of mode character).

probeseq The probe sequence data.frame (see details).

i.row Row indexes in the data.frame (see details).

xy.offset Offset for the xy coordinates. if NULL, uses the default offset stored as an option for the affy package.

x.colname, y.colname The probe sequence packages have seen the names for the columns in their data.frame. This parameters exists to let us follow these changes.

Details

The `data.frame` passed as argument `probeseq` is expected to have (at least) the following columns: `Probe.X`, `Probe.Y` and `Probe.Set.Name`. When the argument `ppset.id` is not null, the probe sets

Value

A matrix of two columns. The first column contains x coordinates, while the second column contains y coordinates.

Warning

The parameter `xy.offset.one` is here for historical reasons. This should not be touched, the option in the `affy` package should be modified if one wishes to modify this.

This function should not be confused with the methods `index2xy` and similar. Here the the XY coordinate come from a `data.frame` that stores information about an arbitrary number probes on the chip. (See the 'probe sequence' data packages on Bioconductor, and the package `Biostrings`).

The methods `index2xy` are meant to interact with instances of class `AffyBatch`.

Author(s)

Laurent

Examples

```
##---- Should be DIRECTLY executable !! ----
```

| | |
|-----------------------|--|
| <code>index2xy</code> | <i>Functions to shuttle from indexes to XY coordinates</i> |
|-----------------------|--|

Description

Functions to shuttle from indexes to XY coordinates.

Usage

```
index2xy(object, ...)
xy2index(object, ...)
index2xy.CdfEnvAffy(object, i)

xy2index.CdfEnvAffy(object, x, y)
```

Arguments

| | |
|---------------------|--|
| <code>object</code> | An object of class <code>CdfEnvAffy</code> . |
| <code>i</code> | A vector of indexes. |
| <code>x, y</code> | Vectors of X and Y coordinates. |
| <code>...</code> | Optional parameters (not used). |

Value

A vector of integers (for `xy2index` methods), or a matrix of two columns (for `index2xy` methods).

See Also

[CdfEnvAffy-class](#)

Examples

```
## To be done...
```

```
indexProbes.CdfEnvAffy  
                          indexes for probes
```

Description

A function to get the index for probes

Usage

```
indexProbes.CdfEnvAffy(object, which, probeSetNames = NULL)
```

Arguments

| | |
|----------------------------|--|
| <code>object</code> | CdfEnvAffy |
| <code>which</code> | which kind of probe are of interest (see details). |
| <code>probeSetNames</code> | names of the probe sets of interest. If NULL, all the probe sets are considered. |

Details

The parameter `which` let one specify which category of probes are of interest. In the case of Affymetrix chips, probes can be "pm" probes or "mm" probes. If the parameter is set to `c("pm", "mm")`, both are returned. Should other categories be defined, they can be handled as well.

Value

A list of indexes.

See Also

[CdfEnvAffy-class](#), [AffyBatch-class](#)

| | |
|-----------------|--|
| matchAffyProbes | <i>Match the probes on an Affymetrix array</i> |
|-----------------|--|

Description

Match the individual probes on an Affymetrix array to arbitrary targets.

Usage

```
mmProbes(probes)

matchAffyProbes(probes, targets, chip_type,
                matchmm = TRUE,
                selectMatches = function(x) which(elementNROWS(x) > 0),
                ...)
```

Arguments

| | |
|---------------|---|
| probes | a probetable object |
| targets | a vector of references |
| chip_type | a name for the chip type. |
| matchmm | whether to match MM probes or not |
| selectMatches | a function to select matches (see Details). |
| ... | further arguments to be passed to matchPDict. |

Details

The matching is performed by the function [matchPDict](#). The man page for that function will indicate what are the options it accepts.

In the case where a large number targets are given, like when each target represents a possible mRNA, is it expected to have a largely sparse incidence matrix, that is a low number of probes matching every target. For that reason, only the index of matching probes are associated with each given target, with the function `selectMatches` giving the definition of what are matching probes. The default function just count anything matching, but the user can specify a more stringent definition if wanted.

Value

`mmProbes` returns a vector of MM probe sequences.
`matchAffyProbes` returns an instance of `AffyProbesMatch-class`.

Author(s)

Laurent Gautier

See Also

[matchPDict](#) for details on how the matching is performed, [AffyProbesMatch-class](#) and [buildCdfEnv.biostrings](#)

Examples

```
library(hgu133aprobe)

filename <- system.file("exampleData", "sample.fasta",
                        package="altcdfenvs")

fasta.seq <- readDNASTringSet(filename)

targets <- as.character(fasta.seq)
names(targets) <- sub("^>.\|\\|(NM[^ \\|]+|Hs[^ \\|]+)\\| ? .+$$", "",
                    names(targets))

m <- matchAffyProbes(hgu133aprobe,
                    targets,
                    "HG-U133A")
```

plot.CdfEnvAffy *A function to 'plot' a CdfEnvAffy*

Description

A function to set the axis and plot the outline for a CdfEnvAffy

Usage

```
## S3 method for class 'CdfEnvAffy'
plot(x, xlab = "", ylab = "", main = x@chipType, ...)
```

Arguments

| | |
|------|--|
| x | a CdfEnvAffy |
| xlab | label for the rows |
| ylab | label for the columns |
| main | label for the plot. The chip-type by default. |
| ... | optional parameters to be passed to the underlying function plot |

Details

This function does not 'plot' much, but sets the coordinates for further plotting (see the examples).

Author(s)

Laurent

See Also[CdfEnvAffy-class](#)**Examples**

```
## See "CdfEnvAffy-class"
```

read.FASTA.entry *Functions to work with FASTA files / connections*

Description

Set of function to work with biological sequences stored in FASTA format.

Usage

```
countskip.FASTA.entries(con, linebreaks = 3000)
grep.FASTA.entry(pattern, con, ...)
## S3 method for class 'FASTA'
print(x, ...)
read.FASTA.entry(con, linebreaks = 3000)
read.n.FASTA.entries(con, n, linebreaks = 3000)
read.n.FASTA.entries.split(con, n, linebreaks = 3000)
read.n.FASTA.headers(con, n, linebreaks = 3000)
read.n.FASTA.sequences(con, n, linebreaks = 3000)
skip.FASTA.entry(con, skip, linebreaks = 3000)
write.FASTA(x, file="data.fasta", append = FALSE)
```

Arguments

| | |
|------------|--|
| append | append to the file (or not) |
| con | a connection |
| file | a file name |
| linebreaks | (to optimize the parsing, probably safe to leave it as it is) |
| n | number of entries to read |
| pattern | a pattern (to be passed to the function grep) |
| skip | number of entries to skip |
| x | a FASTA sequence object |
| ... | optional arguments to be forwarded to the function print or to the function grep |

Details

countskip.FASTA.entries skips the remaining FASTA entries currently remaining in the connection and return the count. grep.FASTA.entry returns the next FASTA entry in the connection that matches a given regular expression. print.FASTA prints a FASTA object. read.FASTA.entry reads the next FASTA entry in the connection. read.n.FASTA.entries reads the n next FASTA entries and returns a list of FASTA objects. read.n.FASTA.entries.split reads the n next FASTA entries and returns a list of two elements: headers and sequences. read.n.FASTA.headers reads the n next FASTA headers. read.n.FASTA.sequences reads the n next FASTA sequences. skip.FASTA.entry skips a given number of FASTA entries. write.FASTA write a FASTA object into a connection.

Value

The value returned depends on the function. See above.

Author(s)

Laurent Gautier

Examples

```
filename <- system.file("exampleData", "sample.fasta",
                        package="altcdfenvs")
con <- file(filename, open="r")

fasta.seq <- grep.FASTA.entry("NM_001544\\.2", con)
close(con)

print(fasta.seq)
```

removeIndex

A function to remove probes in an environment

Description

A function to remove probes in an environment, given their index.

Usage

```
removeIndex(x, i, simplify = TRUE, verbose = FALSE)
```

Arguments

| | |
|----------|--|
| x | An instance of CdfEnvAffy-class |
| i | A vector of indexes (integers !). |
| simplify | Simply the resulting CdfEnvAffy (see details). |
| verbose | verbose output or not. |

Details

The probes to be removed are set to NA in the CdfEnvAffy. When `simplify` is set to TRUE the probe sets are simplified whenever possible. For example, if both pm and mm for the same probe pair are set to NA, then the probe pair is removed from the probe set.

Value

An instance of CdfEnvAffy-class is returned.

Author(s)

Laurent Gautier

See Also

[CdfEnvAffy-class](#)

Examples

```
## use plasmodiumanopheles chip as an example
if (require(plasmodiumanophelescdf)) {

  ## wrap in a (convenient) CdfEnvAffy object
  planocdf <- wrapCdfEnvAffy(plasmodiumanophelescdf, 712, 712, "plasmodiumanophelescdf")
  print(planocdf)

  ## ask for the probe indexed '10759' to be removed
  ## (note: if one wishes to remove from X/Y coordinates,
  ## the function xy2index can be of help).
  planocdfCustom <- removeIndex(planocdf, as.integer(10759))

  ## let see what happened (we made this example knowing in which
  ## probe set the probe indexed '10759' is found).
  indexProbes(planocdf, "pm", "200000_s_at")
  indexProbes(planocdfCustom, "pm", "200000_s_at")
  ## The 'second' pm probe (indexed '10579') in the probe set is now set
  ## to NA.
}
```

toHypergraph

Transform to an hypergraph

Description

Transform to an hypergraph

Usage

```
toHypergraph(object, ...)
```

Arguments

object Object derived from class AffyProbesMatch.
 ... Unused.

Value

An [Hypergraph-class](#) object.

unique.CdfEnvAffy *Remove duplicated elements from a CdfEnvAffy*

Description

Remove duplicated elements from a CdfEnvAffy

Usage

```
## S3 method for class 'CdfEnvAffy'
unique(x, incomparables = FALSE, simplify = TRUE, verbose = FALSE, ...)
```

Arguments

x An instance of CdfEnvAffy-class
 incomparables (not yet implemented)
 simplify simplify the result
 verbose verbose or not
 ... (here for compatibility with the generic unique)

Details

The parameter `simplify` has the same function as the one with the same name in `countduplicated`.

Value

An instance of CdfEnvAffy-class in which probes used several times are removed.

Warning

The function differs slightly from the generic `unique`. Here the elements found in several place a merely removed.

Author(s)

Laurent

See Also

[countduplicated](#)

Examples

```
##not yet here...
```

| | |
|----------------|--|
| validAffyBatch | <i>Check validity of a CdfEnvAffy.</i> |
|----------------|--|

Description

Tries to see if a CdfEnvAffy, or a pair of AffyBatch / CdfEnvAffy is valid.

Usage

```
validAffyBatch(abatch, cdfenv)  
validCdfEnvAffy(cdfenv, verbose=TRUE)  
printValidCdfEnvAffy(x)
```

Arguments

| | |
|---------|--|
| abatch | instance of AffyBatch-class |
| cdfenv | instance of CdfEnvAffy-class |
| verbose | verbose or not |
| x | object returned by validCdfEnvAffy |

Details

The function validAffyBatch calls in turn validCdfEnvAffy.

See Also

[AffyBatch-class](#), [CdfEnvAffy-class](#)

Examples

```
## To be done...
```

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