Package 'SiPSiC'

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Type Package
Title Calculate Pathway Scores for Each Cell in scRNA-Seq Data
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Description Infer biological pathway activity of cells from single-cell RNA-sequencing data by calcu-
      lating a pathway score for each cell (pathway genes are specified by the user). It is recom-
      mended to have the data in Transcripts-Per-Million (TPM) or Counts-Per-
      Million (CPM) units for best results. Scores may change when adding cells to or remov-
      ing cells off the data. SiPSiC stands for Single Pathway analysis in Single Cells.
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biocViews Software, DifferentialExpression, GeneSetEnrichment,
      BiomedicalInformatics, CellBiology, Transcriptomics, RNASeq,
      SingleCell, Transcription, Sequencing, ImmunoOncology,
      DataImport
Depends Matrix, SingleCellExperiment
Suggests knitr, rmarkdown, BiocStyle
VignetteBuilder knitr
BugReports https://github.com/DanielDavis12/SiPSiC/issues
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2 SiPSiC-package

R topics documented:

Index				:	
SiPSiC-nackage	Calculate Dath	um Cooper for Ea	ch Cell in scRNA-Sea I	Data	

Description

Infer biological pathway activity of cells from single-cell RNA-sequencing data by calculating a pathway score for each cell (pathway genes are specified by the user). It is recommended to have the data in Transcripts-Per-Million (TPM) or Counts-Per-Million (CPM) units for best results. Scores may change when adding cells to or removing cells off the data. SiPSiC stands for Single Pathway analysis in Single Cells.

Details

Use this package to calculate per-cell scores for a biological pathway of your choice, from single-cell RNA-seq data. Transcripts-Per-Million (TPM) or Counts-Per-Million (CPM) units of the data are recommended for best results.

Author(s)

Daniel Davis, Yotam Drier. Maintainer: Daniel Davis

References

https://medicine.ekmd.huji.ac.il/en/research/yotamd/Pages/default.aspx

See Also

getPathwayScores

getPathwayScores 3

getPathwayScores	Calculate Pathway Scores for all Cells in a scRNA-Seq Data

Description

Calculate a pathway score for each cell included in the input scRNA-seq data, using the cell's transcription level of the pathway's genes.

Usage

```
getPathwayScores(dataMatrix, pathwayGenes)
```

Arguments

a list of two items:

dataMatrix a matrix whose rows and columns represent genes and cells, respectively, con-

taining the scRNA-seq data; Counts-Per-Million (CPM) or Transcripts-Per-Million (TPM) units should be used for best results. The matrix should be of type sparse

matrix (dgCMatrix), otherwise errors might be raised.

pathwayGenes a character vector of the gene names of which the relevant biological pathway

consists.

Value

pathwayScores an array (type double) of the pathway score of each cell in the input dataMatrix,

unless none of the pathway genes is found in the data, in which case NA is

returned.

index a numeric array of the row indices in the dataMatrix where genes of the pathway

were found.

Author(s)

Daniel Davis, Yotam Drier

References

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https://medicine.ekmd.huji.ac.il/en/research/yotamd/Pages/default.aspx
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Examples

```
library(SiPSiC)
geneCountsMatrix <- matrix(rpois(16, lambda = 10), ncol = 4, nrow = 4)
geneCountsMatrix <- as(geneCountsMatrix, "dgCMatrix")
rownames(geneCountsMatrix) <- c("Gene1", "Gene2", "Gene3", "Gene4")
colnames(geneCountsMatrix) <- c("Cell1", "Cell2", "Cell3", "Cell4")
assayData <- SingleCellExperiment(assays = list(counts = geneCountsMatrix))
pathwayGenesList <- c("Gene1", "Gene2", "Gene4")</pre>
```

4 normalizeCountsForGene

```
scoresAndIndices <- getPathwayScores(counts(assayData), pathwayGenesList)
pathwayScoresOfCells <- scoresAndIndices$pathwayScores
pathwayGeneIndices <- scoresAndIndices$index</pre>
```

normalizeCountsForGene

Gene counts normalization

Description

Get the counts of a single gene normalized by the median of the top 5 percent cells, unless it's zero; In this case, the counts are all divided by the maximum value across all cells. If all counts are zeros, they are returned untouched.

Usage

normalizeCountsForGene(expressionValues)

Arguments

expressionValues

An array of type double, containing the counts (in any units, e.g. CPM or TPM) of a single gene across different cells.

Format

The format is: num 5

Value

An array (type double) of the normalized input counts.

Author(s)

Daniel Davis, Yotam Drier

Index