

# Introduction to RBM package

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## 1 Overview

This document provides an introduction to the `RBM` package. The `RBM` package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the `RBM` package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

## 2 Getting started

The RBM package can be installed and loaded through the following R code.  
Install the RBM package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

## 3 RBM\_T and RBM\_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The  $p$ -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(myresult$permutation_p<=0.05)
```

```

[1] 87

> which(myresult$permutation_p<=0.05)

[1] 2 11 12 29 32 36 44 52 53 67 71 90 104 120 121 125 129 133 145
[20] 151 181 189 193 205 206 207 214 222 224 226 246 253 256 284 314 358 375 382
[39] 398 418 432 437 444 448 465 499 503 533 539 542 551 558 563 610 646 652 655
[58] 673 685 695 727 731 736 743 753 764 779 800 802 815 821 824 835 872 880 891
[77] 906 916 920 930 938 948 952 956 966 976 981

> sum(myresult$bootstrap_p<=0.05)

[1] 10

> which(myresult$bootstrap_p<=0.05)

[1] 36 121 129 189 237 246 253 452 916 948

> permutation_adj_p <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adj_p<=0.05)

[1] 7

> bootstrap_adj_p <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adj_p<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 15

> which(myresult2$bootstrap_p<=0.05)

[1] 12 19 63 88 234 340 427 594 636 657 660 673 768 805 880

> bootstrap2_adj_p <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adj_p<=0.05)

[1] 0

```

- Examples using the `RBM_F` function: `normdata_F` simulates a standardized gene expression data and `unifdata_F` simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```
> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)
```

	Length	Class	Mode
ordfit_t	3000	-none-	numeric
ordfit_pvalue	3000	-none-	numeric
ordfit_beta1	3000	-none-	numeric
permutation_p	3000	-none-	numeric
bootstrap_p	3000	-none-	numeric

```
> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 80

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 81

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 72

> which(myresult_F$permutation_p[, 1]<=0.05)

[1] 26 35 46 48 85 98 107 130 162 213 217 219 229 272 338 359 360 366 370
[20] 391 393 399 405 411 423 439 446 447 455 458 477 479 481 489 490 515 530 536
[39] 539 565 566 569 584 593 599 614 615 627 634 654 672 689 701 702 724 729 746
[58] 750 768 779 798 801 807 836 841 843 849 850 853 854 859 863 869 887 889 944
[77] 951 980 991 996

> which(myresult_F$permutation_p[, 2]<=0.05)

[1] 26 46 48 85 98 102 104 130 162 213 217 219 224 229 251 270 280 359 360
[20] 366 370 391 393 405 423 439 446 447 455 458 477 479 481 490 536 538 539 565
[39] 566 569 584 587 593 599 614 615 627 634 654 672 684 689 701 702 729 746 750
[58] 768 779 798 801 807 836 841 843 849 850 853 854 859 863 869 876 887 889 944
[77] 951 980 989 991 996

> which(myresult_F$permutation_p[, 3]<=0.05)
```

```

[1] 26 35 46 48 85 93 98 130 162 198 213 217 219 229 249 359 360 366 391
[20] 393 399 405 411 423 446 455 458 477 479 481 490 536 539 565 569 593 599 614
[39] 615 627 634 654 672 689 700 701 702 707 729 746 768 779 798 801 807 836 841
[58] 843 850 853 854 859 863 869 876 887 889 951 974 980 991 996

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

[1] 18

> con2_adjp <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adjp<=0.05/3)

[1] 20

> con3_adjp <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adjp<=0.05/3)

[1] 10

> which(con2_adjp<=0.05/3)

[1] 130 359 360 393 458 479 490 539 615 634 701 768 779 798 801 807 863 951 980
[20] 991

> which(con3_adjp<=0.05/3)

[1] 130 359 360 615 634 702 768 798 801 980

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

      Length Class  Mode
ordfit_t      3000  -none- numeric
ordfit_pvalue 3000  -none- numeric
ordfit_beta1  3000  -none- numeric
permutation_p 3000  -none- numeric
bootstrap_p   3000  -none- numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 65

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

```

```

[1] 63

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 69

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 38 108 117 124 125 127 146 150 190 207 223 233 253 257 261 282 288 311 321
[20] 323 332 336 344 364 377 423 427 431 433 439 450 457 500 527 562 583 600 617
[39] 627 636 638 655 683 688 691 709 730 753 791 818 820 824 840 855 858 883 885
[58] 891 939 944 952 967 974 982 999

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 57 65 108 117 125 127 128 133 146 150 190 204 207 233 253 257 261 282 287
[20] 288 321 323 329 332 336 344 364 377 423 427 431 439 450 457 507 517 527 550
[39] 583 600 617 627 638 655 683 688 709 753 791 811 818 820 835 840 855 870 883
[58] 885 891 967 974 982 988

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 57 65 108 111 117 125 127 146 150 207 223 233 253 257 261 282 321 323 327
[20] 329 332 336 344 364 377 423 427 431 439 450 457 527 557 580 583 589 617 624
[39] 627 638 654 681 683 688 692 707 709 720 730 753 755 773 791 811 818 820 824
[58] 840 855 870 883 891 939 952 955 967 974 982 988

> con21_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adj_p<=0.05/3)

[1] 11

> con22_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adj_p<=0.05/3)

[1] 8

> con23_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adj_p<=0.05/3)

[1] 3

```

## 4 Ovarian cancer methylation example using the RBM\_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM\_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the RBM\_T function and presenting the results for further validation and investigations.

```
> system.file("data", package = "RBM")

[1] "/private/var/folders/v1/y6dg5h4n163dzmrfl6t_r5480000gp/T/RtmpOTZlW5/Rinst655f5bd2af49/RBM/d

> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)
```

IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]
cg00000292: 1	Min. :0.01058	Min. :0.01187	Min. :0.009103
cg00002426: 1	1st Qu.:0.04111	1st Qu.:0.04407	1st Qu.:0.041543
cg00003994: 1	Median :0.08284	Median :0.09531	Median :0.087042
cg00005847: 1	Mean :0.27397	Mean :0.28872	Mean :0.283729
cg00006414: 1	3rd Qu.:0.52135	3rd Qu.:0.59032	3rd Qu.:0.558575
cg00007981: 1	Max. :0.97069	Max. :0.96937	Max. :0.970155
(Other) :994		NA's :4	

exmdata4[, 2]	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]
Min. :0.01019	Min. :0.01108	Min. :0.01937	Min. :0.01278
1st Qu.:0.04092	1st Qu.:0.04059	1st Qu.:0.05060	1st Qu.:0.04260
Median :0.09042	Median :0.08527	Median :0.09502	Median :0.09362
Mean :0.28508	Mean :0.28482	Mean :0.27348	Mean :0.27563
3rd Qu.:0.57502	3rd Qu.:0.57300	3rd Qu.:0.52099	3rd Qu.:0.52240
Max. :0.96658	Max. :0.97516	Max. :0.96681	Max. :0.95974
	NA's :1		

```
exmdata8[, 2]
Min. :0.01357
1st Qu.:0.04387
Median :0.09282
Mean :0.28679
3rd Qu.:0.57217
Max. :0.96268

> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
```

```

> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)

              Length Class  Mode
ordfit_t      1000  -none- numeric
ordfit_pvalue 1000  -none- numeric
ordfit_beta0   1000  -none- numeric
ordfit_beta1   1000  -none- numeric
permutation_p 1000  -none- numeric
bootstrap_p    1000  -none- numeric

> sum(diff_results$ordfit_pvalue<=0.05)

[1] 45

> sum(diff_results$permutation_p<=0.05)

[1] 78

> sum(diff_results$bootstrap_p<=0.05)

[1] 63

> ordfit_adjp <- p.adjust(diff_results$ordfit_pvalue, "BH")
> sum(ordfit_adjp<=0.05)

[1] 0

> perm_adjp <- p.adjust(diff_results$permutation_p, "BH")
> sum(perm_adjp<=0.05)

[1] 7

> boot_adjp <- p.adjust(diff_results$bootstrap_p, "BH")
> sum(boot_adjp<=0.05)

[1] 2

> diff_list_perm <- which(perm_adjp<=0.05)
> diff_list_boot <- which(boot_adjp<=0.05)
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t[
> print(sig_results_perm)

      IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
19  cg00016968 0.80628480          NA    0.81440820    0.83623180
83  cg00072216 0.04505377    0.04598964    0.04000674    0.03231534
237 cg00215066 0.94926640    0.95311870    0.94634910    0.94561120
245 cg00224508 0.04479948    0.04972043    0.04152814    0.04189373

```



```

280 cg00260778 0.64319890    0.60488960    0.56735060    0.53150910
764 cg00730260 0.90471270    0.90542290    0.91002680    0.91258610
911 cg00888479 0.07388961    0.07361080    0.10149800    0.09985076
      exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
19      0.80831380    0.73306440    0.82968340    0.84917800
83      0.04965089    0.04833366    0.03466159    0.04390894
237     0.94837410    0.94665570    0.94089070    0.94600090
245     0.04208405    0.05284988    0.03775905    0.03955271
280     0.61920530    0.61925200    0.46753250    0.55632410
764     0.90575890    0.88760470    0.90756300    0.90946790
911     0.08633986    0.06765189    0.09070268    0.12417730
      diff_results$ordfit_t[diff_list_perm]
19      -2.446404
83      2.514109
237     1.419654
245     1.962457
280     4.170347
764    -1.808081
911    -3.621731
      diff_results$permutation_p[diff_list_perm]
19      0
83      0
237     0
245     0
280     0
764     0
911     0

```

```

> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t[diff_list_boot, ])
> print(sig_results_boot)

```

```

      IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
146 cg00134539 0.61101320    0.53321780    0.45999340    0.46787420
349 cg00332745 0.04703361    0.04634372    0.03676908    0.04518837
      exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
146     0.67191510    0.63137380    0.47929610    0.45428300
349     0.04975075    0.05253778    0.04444665    0.03717721
      diff_results$ordfit_t[diff_list_boot]
146      5.394750
349      2.165826
      diff_results$bootstrap_p[diff_list_boot]
146      0
349      0

```