RNA-seq Data Employed in The Sequencing Quality Control (SEQC) Project

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1 Introduction

This vignette briefly describes the content in the seqc package.

The seqc package provides a series of data frames derived from the SEQC project between 2011 to 2014. Three types of data frames are included in this package: RNA-seq read count tables, junction tables and an additional gene intensity table generated by using the TaqMan RT-PCR technology. All the data frames are ready to use in the R environment once the library is loaded.

```
> library(seqc)
> options(width=110, digits=2)
> 1s(2)
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 [5] "ILM_aceview_gene_MAY" "ILM_aceview_gene_NVS" "ILM_junction_AGR_A"
                                                                           "ILM_junction_AGR_B"
                            "ILM_junction_AGR_D"
                                                    "ILM_junction_BGI_A"
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[13] "ILM_junction_BGI_C"
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                                                                           "ILM_junction_CNL_B"
                            "ILM_junction_CNL_D"
[17] "ILM_junction_CNL_C"
                                                    "ILM_junction_COH_A"
                                                                           "ILM_junction_COH_B"
                            "ILM_junction_COH_D"
                                                    "ILM_junction_MAY_A"
[21] "ILM_junction_COH_C"
                                                                           "ILM_junction_MAY_B"
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                            "ILM_junction_MAY_D"
                                                    "ILM_junction_NVS_A"
                                                                           "ILM_junction_NVS_B"
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                            "ILM_junction_NVS_D"
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                                                   "ILM_refseq_gene_MAY"
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                            "LIF_junction_LIV_B"
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                                                                           "LIF_junction_LIV_D"
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                            "ROC_junction_NYU_A"
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[73] "seqc.eSet"
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                                                    "taqman"
```

Six (6) samples were employed in the SEQC project [1]. Sample A and B were obtained from two human RNA reference libraries, while a small amount of Ambion ERCC RNA Spike-in Mix was added into both samples. Sample A and B were then mixed up to different ratios, creating Sample C and D. The pure ERCC Spike-in Mix 1 and 2 were also used as sample E and F, ending up with totally six samples being studied in this

project. The details about the samples are further described in the next section. Huge amounts of RNA-seq data (short reads) were yielded from the six samples at twelve (12) sequencing site by using three (3) different platforms (sequencers).

Genes that have different expression levels among samples can be detected from the RNA-seq data. The read count tables in this package are useful for the purpose of Differently Expressed (DE) gene analysis. For generating the read count tables, the RNA-seq data was mapped to the human genome by using the Subread aligner [2], and the mapping result was assigned to the annotated genes by using the featureCounts program [3] for estimating their expression levels. Two sets of gene annotations were referred when the mapping result was assigned to the genomes: the AceView annotations and the RefSeq annotations, both from the National Center for Biotechnology Information (NCBI) in the United States.

The RNA-seq technologies give an opportunity to precisely call the exon-exon junctions that are coded in the RNA transcriptome. We used the Subjunc program [2] to process the mapping result from the Subread aligner, therefore detecting the exon-exon junctions by using the seed-and-vote strategy. The junction tables from Subjunc were summarized on the sample level and are included in this package.

In addition to the analyses on the RNA-seq data, samples A, B, C and D were also analyzed by using the TaqMan RT-PCR technology. The intensity values of the selected genes are included in this package as a reference.

2 Samples and Sequencing

The SEQC consortium prepared six RNA samples and distributed the samples to twelve sequencing sites. The first two samples, A and B, were derived from Agilent's Universal Human Reference RNA (UHRR) and Life Technologies' Human Brain Reference RNA (HBRR) cell lines respectively. Sample A and B were then mixed with Ambion ERCC RNA Spike-In Mix 1 and 2 accordingly, which are two mixtures of RNA molecules from 92 contigs, but each contig is known to have different density levels in the two mixtures. Sample C and D were then created by mixing sample A and B to different ratios. In sample C, there is 75% of the volume from sample A and 25% of the volume from sample B; the ratio between sample A and B is 25% and 75% in sample D. As we mentioned above, sample E and F are the pure ERCC RNA Spike-In Mix 1 and 2. All the sequencing sites sequenced Sample A and B, but some sites did not sequence sample C, D, E and/or F.

Each of the samples has a number of replicates. Samples A, B, C and D each has five (5) replicates, and Sample E and F each has two (2) replicates. Some sequencing sites sequenced all of these replicates, but the other sites only sequenced a part of them. The set of replicates sequenced at each site can be found from the column names of the data frames in this package. The naming scheme is described in the next chapter.

Three RNA-seq platforms were examined in the SEQC project: the Illumina HiSeq 2000 devices (ILM), the Roche 454 GS FLX platform (ROC) and Life Technologies'

SOLiD 5500 instruments (LIF). Each sequencing site uses a number of lanes and flowcells to sequence each replicate, but the numbers of lanes and flowcells used in the SEQC project differ between the platforms and event between the sequencing sites using the same platform. This project generated 2758 libraries in total: 1832 of them from the Illumnia HiSeq platform, 12 from the 454 Life Science platform and 914 from the SOLiD systems. Table 1 lists the sequencing sites, the platforms employed, the number of samples/replicates that were sequenced at each site, and the numbers of short read libraries yielded.

The 2758 libraries have different read formats. All the libraries from Illumnia HiSeq are paired-end, and the length of every single read is around 100bp. The 454 Life Science libraries, on the other hand, are all single-end, and the read length is variable from tens of bases to above one thousand bases. Different to the libraries from Illumnia HiSeq and 454 Life Science, which contain only base-space reads, all the SOLiD libraries contain only color-space reads. There are 50 single-end libraries and 864 paired-end libraries among all the 914 SOLiD libraries. The read lengths in the SOLiD library vary from 36 to 76 colors long.

Site	Platform	$\mathbf{Samples}$	Replicates	Libraries	PE/SE
AGR	Illumina (ILM)	A, B, C, D	4 for each sample	256	PE
BGI	Illumina (ILM)	A, B, C, D, E, F	5 for A, B, C & D; 2 for E & F	384	$_{ m PE}$
CNL	Illumina (ILM)	A, B, C, D, E, F	5 for A, B, C & D; 2 for E & F	384	${ m PE}$
COH	Illumina (ILM)	A, B, C, D	4 for each sample	128	${ m PE}$
MAY	Illumina (ILM)	A, B, C, D, E, F	5 for A, B, C & D; 2 for E & F	384	${ m PE}$
NVS	Illumina (ILM)	A, B, C, D, E, F	4 for A, B, C & D; 2 for E & F	320	${ m PE}$
LIV	SOLiD (LIF)	A, B, C, D	2 for each sample	50	SE
NWU	SOLiD (LIF)	A, B, C, D, E, F	5 for A, B, C & D; 2 for E & F	288	${ m PE}$
PSU	SOLiD (LIF)	A, B, C, D, E, F	5 for A, B, C & D; 2 for E & F	288	${ m PE}$
SQW	SOLiD (LIF)	A, B, C, D, E, F	5 for A, B, C & D; 2 for E & F	288	${ m PE}$
$\overline{\text{SQW}}$	454 (ROC)	A, B	1 for each sample	4	SE
MGP	454 (ROC)	A, B	1 for each sample	4	${ m SE}$
NYU	454 (ROC)	A, B	1 for each sample	4	SE

Table 1: List of the sequencing sites and processed samples/replicates. SQW used two platforms: SOLiD and 454. The last column indicates if the libraries are paired-end (PE) or single-end (SE)

3 Read Mapping and Read Assignment

All the libraries were mapped to the human genome by using the Subread package.

The reference genome index was first built from the GRCg37/hg19 human genome, plus the 92 ERCC Spike-In contigs¹, the Subread aligner then mapped the RNA-seq libraries to the index using the default settings. The mapping results were then assigned

 $^{^1}$ http://tools.lifetechnologies.com/downloads/ERCC_Controls_Annotation.txt

to the annotated genes by featureCounts, running on the single-end mode or the paired-end mode according to the format of the library. Two sets of annotations were used in the study: the AceView annotations and the RefSeq annotations. There are 1,383,372 exons (from 55,950 genes) in the AceView annotations, and there are 225,074 exons (from 25,072 genes) in the RefSeq annotations; only the exon regions of the genes were extracted from the annotations for read assignment. It is noticed that, although there are much more genes in the AceView annotations, the genes in the RefSeq annotations are not a subset of genes in the AceView annotations. Both sets of annotations have some unique genes not in the other set.

The gene-level read assignment results are provided in this package as 22 data frames, which contain the numbers of single-end reads or paired-end fragments overlapping with exons of each annotated gene. The names of the data frames are synthesized following this scheme: (PLATFORM)_(ANNOTATION)_gene_(SITE), for example, data frame LIF_refseq_gene_NWU includes the read assignment results on the libraries sequenced at NMU by using the SOLiD platform, and the reads were assigned to the RefSeq annotations. The column names in the data frames describe the sample names, the replicate numbers, the lane numbers and the flowcell numbers (if applicable) associated with the libraries. For example, the read counts in LIF_refseq_gene_NWU\$B_4_L02_FlowCell2 are the fourth replicate of Sample B, sequenced by using the second flowcell in lane 2. Another example, ROC_aceview_gene_MGP\$A_1_R02 include the RNA-seq library generated from the first replicate of Sample A at MGP by using the second region of the 454 platform.

> ROC_aceview_gene_MGP[1:15,]

	EntrezID	Symbol	GeneLength	IsERCC	A_1_R01	A_1_R02	B_1_R01	B_1_R02
1	<na></na>	2-OXOACID_DH	1624	FALSE	0	0	0	0
2	<na></na>	A1BGAS	5364	FALSE	1	3	0	2
3	29974	A1CF	2511	FALSE	1	3	0	0
4	< N A >	A2BP1	9426	FALSE	0	0	68	78
5	<na></na>	A2LD1	3848	FALSE	5	2	2	0
6	2	A2M	8153	FALSE	213	169	46	61
7	144568	A2ML1	7492	FALSE	0	1	3	0
8	3	A2MP1	1741	FALSE	0	0	0	0
9	53947	A4GALT	3184	FALSE	5	2	2	1
10	51146	A4GNT	1771	FALSE	0	0	0	1
11	100329167	AAA1	7197	FALSE	0	3	0	0
12	8086	AAAS	4614	FALSE	19	14	7	4
13	65985	AACS	13098	FALSE	14	11	27	20
14	< N A >	AACSL	3594	FALSE	2	0	0	0
15	13	AADAC	3106	FALSE	0	0	0	0

> colnames(ILM_aceview_gene_BGI)

```
[1] "EntrezID" "Symbol" "GeneLength" "ISERCC" "A_1_L01_FlowCellA" [6] "A_1_L01_FlowCellB" "A_1_L02_FlowCellA" "A_1_L02_FlowCellB" "A_1_L03_FlowCellA" "A_1_L03_FlowCellB" [1] "A_1_L04_FlowCellA" "A_1_L04_FlowCellB" "A_1_L05_FlowCellA" "A_1_L05_FlowCellB" "A_1_L06_FlowCellA" [6] "A_1_L06_FlowCellB" "A_1_L06_FlowCellA" "A_1_L06_FlowCellA" "A_1_L06_FlowCellB" "A_1_L06_FlowCellB" "A_1_L06_FlowCellB" "A_1_L06_FlowCellB" "A_1_L06_FlowCellB" "A_1_L05_FlowCellA" "A_1_L08_FlowCellB" "A_1_L08_FlowCellB" [2] "A_2_L01_FlowCellA" "A_2_L01_FlowCellB" "A_2_L02_FlowCellA" "A_2_L02_FlowCellB" "A_2_L03_FlowCellB" "A_2_L03_FlowCellB" "A_2_L03_FlowCellB" "A_2_L05_FlowCellB" "A_2_L05_FlowCellB" "A_2_L05_FlowCellB" "A_2_L05_FlowCellB" "A_2_L05_FlowCellB" "A_2_L05_FlowCellB" "A_2_L05_FlowCellB" "A_3_L02_FlowCellB" "A_3
```

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[321] "D_5_L07_FlowCellA" "D_5_L07_FlowCellB" "D_5_L08_FlowCellA" "D_5_L08_FlowCellB" "E_1_L01_FlowCellA"
[326] "E_1_L01_FlowCellB" "E_1_L02_FlowCellA" "E_1_L02_FlowCellB" "E_1_L03_FlowCellA" "E_1_L03_FlowCellB"
[331] "E_1_L04_FlowCellA" "E_1_L04_FlowCellB" "E_1_L05_FlowCellA" "E_1_L05_FlowCellB" "E_1_L06_FlowCellA"
[336] "E_1_L06_FlowCellB" "E_1_L07_FlowCellA" "E_1_L07_FlowCellB" "E_1_L08_FlowCellA" "E_1_L08_FlowCellB"
[341] "E_2_L01_FlowCellA" "E_2_L01_FlowCellB" "E_2_L02_FlowCellA" "E_2_L02_FlowCellB" "E_2_L03_FlowCellA"
[346] "E_2_L03_FlowCellB" "E_2_L04_FlowCellA" "E_2_L04_FlowCellB" "E_2_L05_FlowCellA" "E_2_L05_FlowCellB"
[351] "E_2_L06_FlowCellA" "E_2_L06_FlowCellB" "E_2_L07_FlowCellA" "E_2_L07_FlowCellB" "E_2_L08_FlowCellA"
[356] "E_2_L08_FlowCellB" "F_1_L01_FlowCellA" "F_1_L01_FlowCellB" "F_1_L02_FlowCellA" "F_1_L02_FlowCellB"
```

```
[361] "F_1_L03_FlowCellA" "F_1_L03_FlowCellB" "F_1_L04_FlowCellA" "F_1_L04_FlowCellB" "F_1_L05_FlowCellA" [366] "F_1_L05_FlowCellB" "F_1_L06_FlowCellA" "F_1_L06_FlowCellB" "F_1_L07_FlowCellB" "F_1_L07_FlowCellB" "F_1_L07_FlowCellB" "F_1_L07_FlowCellB" "F_1_L07_FlowCellB" "F_1_L07_FlowCellB" "F_1_L07_FlowCellB" "F_1_L07_FlowCellA" "F_1_1_L07_FlowCellA" "F_1
```

> ILM_aceview_gene_BGI[1:15,1:7]

	EntrezID	Symbol	GeneLength	IsERCC	A_1_L01_FlowCellA	A_1_L01_FlowCellB	A_1_L02_FlowCellA
1	< N A >	2-0X0ACID_DH	1624	FALSE	0	0	0
2	< N A >	A1BGAS	5364	FALSE	17	15	18
3	29974	A1CF	2511	FALSE	21	14	21
4	< N A >	A2BP1	9426	FALSE	7	5	1
5	< N A >	A2LD1	3848	FALSE	9	15	17
6	2	A2M	8153	FALSE	2683	2777	2660
7	144568	A2ML1	7492	FALSE	4	0	6
8	3	A2MP1	1741	FALSE	1	3	0
9	53947	A4GALT	3184	FALSE	44	58	56
10	51146	A4GNT	1771	FALSE	1	0	0
11	100329167	AAA1	7197	FALSE	5	10	8
12	8086	AAAS	4614	FALSE	369	358	327
13	65985	AACS	13098	FALSE	202	207	184
14	< N A >	AACSL	3594	FALSE	15	12	10
15	13	AADAC	3106	FALSE	0	0	0

4 Junction Detection

The subread aligner can very efficiently map reads to the reference genome. From the mapping results, subjunc furthers detected exon-exon junctions that were coded in the RNA-seq reads², and generated a junction table for each library. We merged the junction tables from the libraries on the platform-site-sample level, then provide the resulted data frames in this package.

The data frames are named following this scheme: (PLATFORM)_junction_(SITE)_SAMPLE. For example, data frame ILM_junction_AGR_B includes the junctions that were detected from all the libraries of Sample B sequenced at AGR by using the Illumina HiSeq platform. Each data frame contains four columns: the chromosome name, the first junction side, the second junction side and the number of supporting reads (i.e., the reads that spanning the junction location and include the two exons surrounding the junction). The two junction sides are defined as the chromosomal position (one starting) of the last base in the first exon, and the chromosomal position of the first base in the second exon both surrounding this junction point.

> ILM_junction_AGR_B[1:15,]

	Chromosome	Location1	Location2	nSupportingReads
1	ERCC-00002	195	228	2
2	ERCC-00002	407	430	2

²The libraries were processed by using an old version of the subread package. The subjunc in that package only takes SAM files from the subread aligner as input. The subjunc program in the newer package (after version 1.4.0) is able to directly take FASTQ and FASTA files as input.

```
ERCC-00002
                      75
                                228
   ERCC-00096
                     527
                                563
   ERCC - 00113
                     130
                                146
                                                   2
5
6
    ERCC-00130
                     757
                                850
                                                   6
    GL000210.1
                    5430
                               5545
                                                   2
8 NT_113878.1
                   29833
                              30763
                                                  12
9 NT_113878.1
                   30969
                              31271
                                                  19
10 NT_113878.1
                   37549
                              41405
                                                  11
11 NT_113878.1
                   37782
                              41405
                                                   1
12 NT 113878.1
                   41272
                              41405
                                                   .3
13 NT_113878.1
                   41456
                              41682
14 NT_113878.1
                    41459
                              41682
                                                   20
15 NT 113885.1
                              31602
                   27618
```

5 TaqMan RT-PCR

The gene expression levels in Samples A, B, C and D were further measured by using the TaqMan RT-PCR technologies. The expression levels of the 1044 selected genes in replicates 1, 2, 3 and 4 of the four samples are stored in data frame taqman. The first column in this data frame are the entrez ids and symbols of the 1044 genes, followed by the gene intensity values in 16 columns, which are named as (SAMPLE).(SAMPLE)(REPLICATE)_values, for example, taqman\$C.C3_values are the gene intensity values in the third replicate of sample C.

```
> colnames(taqman)
                                              "A1_detection" "A2_value"
                                                                          "A2_detection" "A3_value"
 [1] "EntrezID"
                  "Svmbol"
                                "A1_value"
 [8] "A3_detection" "A4_value"
                                "A4_detection" "B1_value" "B1_detection" "B2_value" "B2_detection"
                  "B3_detection" "B4_value"
                                              "B4_detection" "C1_value"
                                                                      "C1_detection" "C2_value"
[15] "B3_value"
[22] "C2_detection" "C3_value"
                                "C3_detection" "C4_value"
                                                           "C4_detection" "D1_value"
                                                                                        "D1_detection"
[29] "D2_value"
                                              "D3_detection" "D4_value"
                  "D2_detection" "D3_value"
                                                                          "D4_detection"
> tagman[1:15, 1:9]
  EntrezID Symbol A1_value A1_detection A2_value A2_detection A3_value A3_detection A4_value
      1543 CYP1A1 0.00861
                           P 0.00875
                                                 P 0.00827
                                                                            P 0.00819
                                   P 0.03002
                                                        P 0.02657
                                                                             P 0.03169
2
      1950
              EGF 0.02697
      1153
            CIRBP
                 2.11583
                                   P 2.11533
                                                        P 2.15163
                                                                            P 2.08393
3
                                   P 0.20006
                                                        P 0.16115
                                                                            P 0.19256
      1613 DAPK3 0.19717
4
      1665 DHX15 0.11647
                                  P 0.11893
                                                        P 0.10562
                                                                            P 0.11335
      1786 DNMT1 0.77968
                                  P 0.81197
                                                        P 0.84169
                                                                            P 0.76226
7
      1982 EIF4G2 7.14649
                                   P 6.76277
                                                        P 7.65979
                                                                             Р
                                                                               7.17128
8
      2069
            EREG
                  0.03583
                                   P 0.03379
                                                        P 0.03350
                                                                            P 0.03665
                                   P 0.00432
                                                        P 0.00412
                                                                            P 0.00488
9
      2196
            FAT2 0.00447
10
      3491 CYR61 0.65991
                                  P 0.66509
                                                       P 0.68998
                                                                            P 0.65407
                                   A 0.00043
11
      1527 CXorf2 0.00043
                                                        A 0.00043
                                                                             A 0.00043
12
      1346 COX7A1
                  0.00291
                                   P 0.00231
                                                        P 0.00222
                                                                             Р
                                                                               0.00177
13
      1357
            CPA1 0.00043
                                   A 0.00043
                                                        A 0.00043
                                                                             A 0.00043
14
      1360
            CPB1 0.00043
                                   A 0.00043
                                                        A 0.00043
                                                                             A 0.00043
      1503
            CTPS 1.10256
                                   P 1.06406
                                                        P 1.07379
                                                                             P 1.12382
```

6 Citation

[1] Su Z, Labaj PP, Li S, Thierry-Mieg J, Thierry-Mieg D, Shi W, Wang C, Schroth GP, Setterquist RA, Thompson JF, et al. (SEQC/MAQC-III Consortium). A comprehensive assessment of RNA-seq accuracy, reproducibility and information content by the

Sequencing Quality Control Consortium. Nature Biotechnology, 32(9), 903-914

- [2] Yang Liao, Gordon K Smyth and Wei Shi (2013). The Subread aligner: fast, accurate and scalable read mapping by seed-and-vote. Nucleic Acids Research, 41(10):e108.
- [3] Yang Liao, Gordon K Smyth and Wei Shi (2014). featureCounts: an efficient general purpose program for assigning sequence reads to genomic features. Bioinformatics, 30(7):923-30

7 Authors

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8 Contact

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