

# Introduction to RBM package

Dongmei Li

November 9, 2022

Clinical and Translational Science Institute, University of Rochester School of Medicine and Dentistry, Rochester, NY 14642-0708

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## 1 Overview

This document provides an introduction to the `RBM` package. The `RBM` package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the `RBM` package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

## 2 Getting started

The RBM package can be installed and loaded through the following R code.  
Install the RBM package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

## 3 RBM\_T and RBM\_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The  $p$ -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(myresult$permutation_p<=0.05)
```

```

[1] 26

> which(myresult$permutation_p<=0.05)

[1] 49 58 105 107 129 171 275 278 311 422 472 476 569 611 694 806 810 813 815
[20] 823 834 899 951 980 984 991

> sum(myresult$bootstrap_p<=0.05)

[1] 0

> which(myresult$bootstrap_p<=0.05)

integer(0)

> permutation_adj_p <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adj_p<=0.05)

[1] 6

> bootstrap_adj_p <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adj_p<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 38

> which(myresult2$bootstrap_p<=0.05)

[1] 20 33 37 57 70 83 143 152 162 256 279 308 335 461 468 493 495 538 575
[20] 597 617 641 679 698 699 737 747 764 774 789 797 808 837 873 885 895 971 977

> bootstrap2_adj_p <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adj_p<=0.05)

[1] 0

```

- Examples using the RBM\_F function: normdata\_F simulates a standardized gene expression data and unifdata\_F simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```

> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)

              Length Class  Mode
ordfit_t      3000   -none-  numeric
ordfit_pvalue 3000   -none-  numeric
ordfit_beta1  3000   -none-  numeric
permutation_p 3000   -none-  numeric
bootstrap_p   3000   -none-  numeric

> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 43

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 52

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 42

> which(myresult_F$permutation_p[, 1]<=0.05)

[1] 17 20 36 44 48 116 122 240 254 340 349 360 369 381 398 411 415 423 425
[20] 444 458 468 530 541 556 565 577 600 607 612 618 670 732 746 747 772 775 845
[39] 892 909 959 985 988

> which(myresult_F$permutation_p[, 2]<=0.05)

[1] 9 17 36 44 48 55 103 116 133 204 254 281 340 349 360 381 391 398 411
[20] 415 423 425 431 444 458 468 503 530 538 541 556 565 577 600 607 612 618 644
[39] 670 727 732 746 772 783 845 892 909 959 980 985 988 995

> which(myresult_F$permutation_p[, 3]<=0.05)

[1] 17 20 36 44 108 122 204 254 340 349 360 369 375 376 381 398 415 423 425
[20] 444 468 530 541 556 565 577 580 600 612 618 670 727 732 746 772 845 892 909
[39] 933 959 985 988

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

[1] 2

```

```

> con2_adjp <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adjp<=0.05/3)

[1] 4

> con3_adjp <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adjp<=0.05/3)

[1] 3

> which(con2_adjp<=0.05/3)

[1] 360 444 468 577

> which(con3_adjp<=0.05/3)

[1] 36 444 746

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

              Length Class  Mode
ordfit_t      3000   -none-  numeric
ordfit_pvalue 3000   -none-  numeric
ordfit_beta1  3000   -none-  numeric
permutation_p 3000   -none-  numeric
bootstrap_p    3000   -none-  numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 37

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 47

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 44

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 8 34 38 108 123 163 210 248 259 272 278 279 302 315 320 351 423 427 450
[20] 487 502 505 628 651 658 706 770 778 779 826 868 877 903 933 967 986 993

```

```

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 8 34 38 54 108 157 189 196 248 259 266 272 278 279 302 315 320 325 351
[20] 383 399 428 450 460 487 502 515 539 567 628 651 657 658 663 706 770 772 778
[39] 779 826 868 877 889 933 978 986 993

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 8 34 38 157 163 171 192 196 210 241 259 266 278 279 302 315 320 325 351
[20] 383 450 487 488 502 539 628 631 638 651 658 706 751 768 772 778 779 816 833
[39] 868 877 933 944 986 993

> con21_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adj_p<=0.05/3)

[1] 6

> con22_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adj_p<=0.05/3)

[1] 5

> con23_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adj_p<=0.05/3)

[1] 5

```

## 4 Ovarian cancer methylation example using the RBM\_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM\_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the RBM\_T function and presenting the results for further validation and investigations.

```

> system.file("data", package = "RBM")

[1] "/private/tmp/RtmpwN2u0N/Rinst114fb4b364a37/RBM/data"

> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)

```

IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]
cg00000292: 1	Min. :0.01058	Min. :0.01187	Min. :0.009103
cg00002426: 1	1st Qu.:0.04111	1st Qu.:0.04407	1st Qu.:0.041543
cg00003994: 1	Median :0.08284	Median :0.09531	Median :0.087042
cg00005847: 1	Mean :0.27397	Mean :0.28872	Mean :0.283729
cg00006414: 1	3rd Qu.:0.52135	3rd Qu.:0.59032	3rd Qu.:0.558575
cg00007981: 1	Max. :0.97069	Max. :0.96937	Max. :0.970155
(Other) :994		NA's :4	

  

exmdata4[, 2]	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]
Min. :0.01019	Min. :0.01108	Min. :0.01937	Min. :0.01278
1st Qu.:0.04092	1st Qu.:0.04059	1st Qu.:0.05060	1st Qu.:0.04260
Median :0.09042	Median :0.08527	Median :0.09502	Median :0.09362
Mean :0.28508	Mean :0.28482	Mean :0.27348	Mean :0.27563
3rd Qu.:0.57502	3rd Qu.:0.57300	3rd Qu.:0.52099	3rd Qu.:0.52240
Max. :0.96658	Max. :0.97516	Max. :0.96681	Max. :0.95974
	NA's :1		

  

exmdata8[, 2]
Min. :0.01357
1st Qu.:0.04387
Median :0.09282
Mean :0.28679
3rd Qu.:0.57217
Max. :0.96268

```

> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)

```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```

> sum(diff_results$ordfit_pvalue<=0.05)

```

```

[1] 45

```

```

> sum(diff_results$permutation_p<=0.05)

```

```

[1] 72

```

```

> sum(diff_results$bootstrap_p<=0.05)

```

```
[1] 56
```

```
> ordfit_adj_p <- p.adjust(diff_results$ordfit_pvalue, "BH")
> sum(ordfit_adj_p<=0.05)
```

```
[1] 0
```

```
> perm_adj_p <- p.adjust(diff_results$permutation_p, "BH")
> sum(perm_adj_p<=0.05)
```

```
[1] 30
```

```
> boot_adj_p <- p.adjust(diff_results$bootstrap_p, "BH")
> sum(boot_adj_p<=0.05)
```

```
[1] 3
```

```
> diff_list_perm <- which(perm_adj_p<=0.05)
> diff_list_boot <- which(boot_adj_p<=0.05)
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t[diff_list_perm, ])
> print(sig_results_perm)
```

	IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]	exmdata4[, 2]
16	cg00014085	0.05906804	0.04518973	0.04211710	0.03665208
19	cg00016968	0.80628480	NA	0.81440820	0.83623180
83	cg00072216	0.04505377	0.04598964	0.04000674	0.03231534
95	cg00081975	0.03633894	0.04975194	0.06024723	0.05598723
103	cg00094319	0.73784280	0.73532960	0.75574900	0.73830220
106	cg00095674	0.07076291	0.05045181	0.03861991	0.03337576
131	cg00121904	0.15449580	0.17949750	0.23608110	0.24354150
146	cg00134539	0.61101320	0.53321780	0.45999340	0.46787420
237	cg00215066	0.94926640	0.95311870	0.94634910	0.94561120
245	cg00224508	0.04479948	0.04972043	0.04152814	0.04189373
259	cg00234961	0.04192170	0.04321576	0.05707140	0.05327565
280	cg00260778	0.64319890	0.60488960	0.56735060	0.53150910
285	cg00263760	0.09050395	0.10197760	0.14801710	0.12242400
349	cg00332745	0.04703361	0.04634372	0.03676908	0.04518837
437	cg00424946	0.04122172	0.04325330	0.03339863	0.02876798
520	cg00502442	0.03163993	0.03581662	0.02785063	0.02549502
627	cg00612467	0.04777553	0.03783457	0.05380982	0.05582291
632	cg00615377	0.11265030	0.16140570	0.19404450	0.17468600
743	cg00717862	0.07999436	0.07873347	0.06089359	0.06171374
764	cg00730260	0.90471270	0.90542290	0.91002680	0.91258610
772	cg00743372	0.03922780	0.02919634	0.02187972	0.02568053
804	cg00777121	0.04540701	0.05430304	0.04154242	0.04221162
848	cg00826384	0.05721674	0.05612171	0.06644259	0.06358381
851	cg00830029	0.58362500	0.59397870	0.64739610	0.67269640



887	cg00862290	0.43640520	0.54047160	0.60786800	0.56325950
911	cg00888479	0.07388961	0.07361080	0.10149800	0.09985076
928	cg00901493	0.03737166	0.03903724	0.04684618	0.04981432
931	cg00901704	0.05734342	0.04812868	0.04478214	0.03878488
939	cg00906183	0.03949030	0.04365079	0.03720015	0.03575748
979	cg00945507	0.13432250	0.23854600	0.34749760	0.28903340
	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]	exmdata8[, 2]	
16	0.04222944	0.05324246	0.03728026	0.04062589	
19	0.80831380	0.73306440	0.82968340	0.84917800	
83	0.04965089	0.04833366	0.03466159	0.04390894	
95	0.04561792	0.05115624	0.06068253	0.06168212	
103	0.67349260	0.73510200	0.75715920	0.78981220	
106	0.04693030	0.06837343	0.04534005	0.03709488	
131	0.17352980	0.12564280	0.18193170	0.20847670	
146	0.67191510	0.63137380	0.47929610	0.45428300	
237	0.94837410	0.94665570	0.94089070	0.94600090	
245	0.04208405	0.05284988	0.03775905	0.03955271	
259	0.04030003	0.03996053	0.05086962	0.05445672	
280	0.61920530	0.61925200	0.46753250	0.55632410	
285	0.11693600	0.10650430	0.12281160	0.12310430	
349	0.04975075	0.05253778	0.04444665	0.03717721	
437	0.03353116	0.03719167	0.03096761	0.03234779	
520	0.03111720	0.03189393	0.02415307	0.02941176	
627	0.04740551	0.05332965	0.05775211	0.05579710	
632	0.12573100	0.14483660	0.16338240	0.20130510	
743	0.07594936	0.09062161	0.06475791	0.07271878	
764	0.90575890	0.88760470	0.90756300	0.90946790	
772	0.02796053	0.03512214	0.02575992	0.02093909	
804	0.04911277	0.04872797	0.04261405	0.04474881	
848	0.05230160	0.06119713	0.06542751	0.06240686	
851	0.50820240	0.34657470	0.66276570	0.64634510	
887	0.50259740	0.40111730	0.56646700	0.54552980	
911	0.08633986	0.06765189	0.09070268	0.12417730	
928	0.04490690	0.04204062	0.05050039	0.05268215	
931	0.04497277	0.05751033	0.03089829	0.04423603	
939	0.03856975	0.06024309	0.03594439	0.03502819	
979	0.11848510	0.16653850	0.30718420	0.26624740	
	diff_results\$ordfit_t[diff_list_perm]				
16		2.325659			
19		-2.446404			
83		2.514109			
95		-3.252063			
103		-2.268711			
106		3.100324			
131		-3.451679			

146	5.394750
237	1.419654
245	1.962457
259	-4.052697
280	4.170347
285	-3.093997
349	2.165826
437	2.102892
520	1.873471
627	-2.239498
632	-3.661161
743	3.444684
764	-1.808081
772	2.416991
804	1.995220
848	-2.314412
851	-2.841244
887	-3.217939
911	-3.621731
928	-2.716443
931	2.464709
939	1.762879
979	-4.750997
diff_results\$permutation_p[diff_list_perm]	
16	0
19	0
83	0
95	0
103	0
106	0
131	0
146	0
237	0
245	0
259	0
280	0
285	0
349	0
437	0
520	0
627	0
632	0
743	0
764	0
772	0

```

804
848
851
887
911
928
931
939
979

```

```

> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t[
> print(sig_results_boot)

```

```

      IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
131 cg00121904 0.1544958      0.1794975      0.2360811      0.2435415
146 cg00134539 0.6110132      0.5332178      0.4599934      0.4678742
632 cg00615377 0.1126503      0.1614057      0.1940445      0.1746860
      exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
131      0.1735298      0.1256428      0.1819317      0.2084767
146      0.6719151      0.6313738      0.4792961      0.4542830
632      0.1257310      0.1448366      0.1633824      0.2013051
diff_results$ordfit_t[diff_list_boot]
131      -3.451679
146      5.394750
632     -3.661161
diff_results$bootstrap_p[diff_list_boot]
131      0
146      0
632      0

```