

Package ‘epistack’

January 13, 2022

Title Heatmaps of Stack Profiles from Epigenetic Signals

Version 1.0.0

Description The epistack package main objective is the visualizations of stacks of genomic tracks (such as, but not restricted to, ChIP-seq, ATAC-seq, DNA methylation or genomic conservation data) centered at genomic regions of interest.

License MIT + file LICENSE

Encoding UTF-8

LazyData false

Imports GenomicRanges, BiocGenerics, S4Vectors, IRanges, viridisLite, graphics, plotrix, grDevices, stats

Roxygen list(markdown = TRUE)

RoxygenNote 7.1.2

Depends R (>= 4.1)

Suggests testthat (>= 3.0.0), BiocStyle, knitr, rmarkdown, EnrichedHeatmap, biomaRt, rtracklayer, covr, vdiff, magick

Config/testthat/edition 3

VignetteBuilder knitr

biocViews RNASeq, Preprocessing, ChIPSeq, GeneExpression

git_url <https://git.bioconductor.org/packages/epistack>

git_branch RELEASE_3_14

git_last_commit fe0a920

git_last_commit_date 2021-10-26

Date/Publication 2022-01-13

Author SACI Safia [aut],
DEVAILLY Guillaume [cre]

Maintainer DEVAILLY Guillaume <gdevailly@hotmail.com>

R topics documented:

addBins	2
addMetricAndArrangeGRanges	3
meanColor	4
plotAverageProfile	5
plotBinning	6
plotBoxMetric	7
plotEpistack	8
plotMetric	10
plotStackProfile	11
plotStackProfileLegend	13
redimMatrix	13
stackepi	15

Index	16
--------------	-----------

addBins	<i>addBins()</i>
---------	------------------

Description

Add an optional bin metadata column to `gr`, to serve as annotations for the epistack plots.

Usage

```
addBins(gr, nbins = 5L, bin = NULL)
```

Arguments

<code>gr</code>	a GRanges object.
<code>nbins</code>	an integer number, the number of bins.
<code>bin</code>	a vector containing pre-determined bins, in the same order as <code>gr</code> .

Details

`nbins` is taken into account only if `bin` is `NULL`. `gr` should be sorted first, usually with the `addMetricAndArrangeGRanges()` function. `addBin(gr, bin = vec)` is equivalent to `gr$bin <-vec`, while `addBin(gr, nbins = 5)` will create 5 bins of equal size based on `gr` order.

Value

the `gr` GRanges object with a new bin metadata column

See Also

[addMetricAndArrangeGRanges](#)

Examples

```
data("stackepi")
addBins(stackepi)

# 3 bins instead of 5
addBins(stackepi, nbins = 3)

# assign bins using a vector
addBins(stackepi, bin = rep(c("a", "b", "c"),
length.out = length(stackepi)))
```

```
addMetricAndArrangeGRanges
  addMetricAndArrangeGRanges()
```

Description

Perform an inner join between a GRanges object and a data.frame. Sort the resulting GRanges based on a metric column.

Usage

```
addMetricAndArrangeGRanges(
  gr,
  order,
  gr_key = "name",
  order_key = "name",
  order_value = "exp",
  shuffle_tie = TRUE
)
```

Arguments

gr	a GRanges object.
order	a data.frame with at least two columns: keys and values.
gr_key	name of the gr metadata column containing unique names for each genomic region in gr. Usually gene names/id or peak id.
order_key	name of the order column that will be used as key for the inner join.
order_value	name of the order column that contain value used for sorting.
shuffle_tie	a boolean Value (TRUE / FALSE). When TRUE, shuffle the GRanges before sorting, mixing the ties.

Details

This utility function allow the addition of a metric column to genomic regions of interest. One of its common use case is to add gene expression values on a set of transcription start sites. The resulting GRanges object will only contain regions presents in both gr and order.

Value

a GRanges sorted in descending order.

Examples

```
data("stackepi")
randomOrder <- data.frame(gene_id = stackepi$gene_id,
  value = rnorm(length(stackepi)))
addMetricAndArrangeGRanges(stackepi,
  randomOrder, gr_key = "gene_id",
  order_key = "gene_id", order_value = "value" )
```

meanColor

meanColor

Description

Return the average color of a vector of colors, computed in the RGB space.

Usage

```
meanColor(colors)
```

Arguments

colors a vector of colors

Details

Input colors can be either in html or color name formats. The alpha channel is supported but optional.

Value

a single color value

See Also

[redimMatrix](#)

Examples

```

meanColor(c("#000000FF", "#FFFFFF00", "#FFFF00FF", "#FF0000FF"))

# works with color names
meanColor(c("blue", "red"))

# Mix color names and HTML codes
meanColor(c("blue", "red", "#FFFF00FF"))

# works without alpha channel in inputs (but outputs an alpha channel):
meanColor(c("#000000", "#FFFFFF", "#FFFF00", "#FF0000"))

```

```
plotAverageProfile    plotAverageProfile()
```

Description

Plot the average stack profiles +/- error (sd or sem). If a bin column is present in gr, one average profile is drawn for each bin.

Usage

```

plotAverageProfile(
  gr,
  pattern = "^window_",
  x_labels = c("Before", "Anchor", "After"),
  palette = colorRampPalette(c("magenta", "black", "green")),
  alpha_for_se = 0.25,
  error_type = c("sd", "sem"),
  reversed_z_order = FALSE,
  ylim = NULL
)

```

Arguments

gr	a GRanges input
pattern	a single character that should match metadata of gr (can be a regular expression).
x_labels	x-axis labels.
palette	a color palette function, by default: colorRampPalette(c("magenta", "black", "green"))
alpha_for_se	the transparency (alpha) value for the error band.
error_type,	can be either sd (standard deviation) or sem (standard error of the mean). Default: sem.
reversed_z_order	should the z-order of the curves be reversed (i.e. first or last bin on top?)
ylim	a vector of two numbers corresponding to the y-limits of the plot

Value

Display a plot.

Examples

```
data("stackepi")
plotAverageProfile(stackepi)
```

plotBinning	<i>plotBinning()</i>
-------------	----------------------

Description

Plot a vertical color bar of the bin column.

Usage

```
plotBinning(
  gr,
  target_height = 650,
  palette = colorRampPalette(c("magenta", "black", "green"))
)
```

Arguments

<code>gr</code>	a GRanges object containing a bin metadata column
<code>target_height</code>	an integer, the approximate height (in pixels) of the final plot. Used to avoid overplotting artefacts.
<code>palette</code>	a function taking a number as a first argument, and returning a vector of colors.

Value

Display a plot.

Examples

```
data("stackepi")
gr <- stackepi
gr <- addBins(gr, nbins = 3)
plot_bin <- plotBinning(gr)

gr2 <- data.frame(bin = rep(c(1,2,3,4), each = 5))
plotBinning(gr2, palette = colorRampPalette(c("blue4", "forestgreen", "coral3", "goldenrod")))
```

plotBoxMetric	<i>plotBoxMetric()</i>
---------------	------------------------

Description

Plot distribution of a metric values as boxplots depending of bins. If the bin is absent from gr, a single boxplot is drawn.

Usage

```
plotBoxMetric(  
  gr,  
  metric = "expr",  
  title = "Metric",  
  trans_func = function(x) x,  
  ylim = NULL,  
  ylab = "metric",  
  palette = colorRampPalette(c("magenta", "black", "green"))  
)
```

Arguments

gr	a GRanges input
metric	name of the column in gr metadata containing scores.
title	title of the plot.
trans_func	A function to transform value of x before plotting. Useful to apply log10 transformation (i.e. with <code>trans_func = function(x) log10(x+1)</code>).
ylim	limit of the y axis; format: <code>ylim = c(min,max)</code>
ylab	y-axis title
palette	a function that returns a palette of n colors.

Value

Display a plot.

Examples

```
data("stackepi")  
plotBoxMetric(  
  stackepi,  
  trans_func = function(x) x,  
  metric = "exp",  
  title = "Metric"  
)
```

plotEpistack	<i>plotEpistack()</i>
--------------	-----------------------

Description

Given a list of genomic regions, epigenetic signals surrounding these regions, and a score for each regions, plot epigenetic stacks depending on the score. An optional bin column allow the grouping of several genomic regions to produce average profiles per bins.

Usage

```
plotEpistack(
  gr,
  patterns = "^window_",
  tints = "gray",
  titles = "",
  legends = "",
  x_labels = c("Before", "Anchor", "After"),
  zlim = c(0, 1),
  ylim = NULL,
  metric_col = "expr",
  metric_title = "Metric",
  metric_label = "metric",
  metric_transfunc = function(x) x,
  bin_palette = colorRampPalette(c("magenta", "black", "green")),
  npix_height = 650,
  n_core = 1,
  high_mar = c(2.5, 0.6, 4, 0.6),
  low_mar = c(2.5, 0.6, 0.3, 0.6),
  error_type = c("sd", "sem"),
  ...
)
```

Arguments

<code>gr</code>	a GRanges input.
<code>patterns</code>	a character vector of column prefixes (can be regular expressions) that should match columns of <code>gr</code> .
<code>tints</code>	a vector of colors to tint the heatmaps.
<code>titles</code>	titles of each heatmap.
<code>legends</code>	legend names for the epistacks.
<code>x_labels</code>	a character vector of length 3 used as x-axis labels.
<code>zlim</code>	the minimum and maximum z values the heatmap. Format: <code>zlim = c (min,max)</code>
<code>ylim</code>	limits of the y axis for bottom plots. Format: <code>ylim = c (min,max)</code>

<code>metric_col</code>	a character, name of a column in <code>gr</code> such as expression value, peak height, pvalue, fold change, etc.
<code>metric_title</code>	title to be display on the leftmost plots.
<code>metric_label</code>	label of the leftmost plots.
<code>metric_transfunc</code>	a function to transform value of <code>metric_col</code> before plotting. Useful to apply <code>log10</code> transformation (i.e. with <code>trans_func = function(x) log10(x+1)</code>).
<code>bin_palette</code>	a palette of color, (i.e. a function of parameter <code>n</code> that should retrun <code>n</code> colors), used to color average profiles per bin in the bottom plots.
<code>npix_height</code>	The matrix height is reduced to this number of rows before plotting. Useful to limit overplotting artefacts. It should roughlyly be set to the pixel height in the final heatmaps
<code>n_core</code>	number of core used to speedup the martrix resizing.
<code>high_mar</code>	a vector of numerical values corresponding to the margins of the top figures. <code>c(bottom, left, top, right)</code>
<code>low_mar</code>	a vector of numerical values corresponding to the margins of the bottom figures. <code>c(bottom, left, top, right)</code>
<code>error_type</code> ,	can be either <code>sd</code> (standard deviation) or <code>sem</code> (standard error of the mean). Default: <code>sem</code> .
<code>...</code>	Arguments to be passed to <code>par</code> such as <code>cex</code>

Details

This function produce a comprehensive figure including epigenetic heatmaps and average epigenetic profiles from a well formatted `GRanges` object with expected metadata columns. It scales resonably well up to hundreds of thousands of genomic regions.

The visualisation is centered on an anchor, a set of genomic coordinated that can be transcription start sites or peak center for example. Anchor coordinates are those of the `GRanges` used as an input (hereafter `gr`).

Anchors are plotted from top to bottom in the same order as in `gr`. One should sort `gr` before plotting if needed.

`gr` should have a metric column that is used in the leftmost plots. The name of the metric column must be specified to `metric_col`. The metric can be transformed before plotting if needed using the `metric_transfunc` parameter.

The matrix or matrices used to display the heatmap(s) should be passed as additional metadata columns of `gr`. Such matrix can be obtained using `EnrichedHeatmap::normalizeToMatrix()` for example. The matrix columns names are then specified through patterns using prefixes, suffixes or regular expressions.

If an optionnal bin column is present in `gr`, it will be used to group genomic regions to performed average profile per bins in the bottom plots.

Epistack are multipanel plots build using `layout()`. Margins for the panels can be specified using `high_mar` and `low_mar` parameters if needed, especially to avoid text overlaps. The default value should be appropriate in most situations. Individual component can be plotted using severa epistack functions such has `plotStackProfile()` or `plotAverageProfile()`.

Plotting more than > 1000 regions can lead to overplotting issued as well as some plotting artefacts (such as horizontal white strips). Both issues can be resolved with fidling with the npix_height parameter. npix_height should be smaller than the number of regions, and in the same order of magnitude of the final heatmap height in pixels. Last minutes call to the redimMatrix() function will hapen before plotting using npix_height as target height. Parameter n_core is passed to redimMatrix() to speed up the down-scaling.

Value

Display a plot.

See Also

[plotStackProfile](#), [plotAverageProfile](#), [redimMatrix](#), [normalizeToMatrix](#), [addMetricAndArrangeGRanges](#), [addBins](#)

Examples

```
data("stackepi")
plotEpistack(stackepi,
  metric_col = "exp",
  ylim = c(0, 1),
  metric_transfunc = function(x) log10(x+1))
```

plotMetric	<i>plotMetric()</i>
------------	---------------------

Description

Plot a vertical line chart of the metric column, in the same order as the input.

Usage

```
plotMetric(
  x,
  trans_func = function(x) x,
  title = "Metric",
  ylim = NULL,
  xlab = NULL
)
```

Arguments

x	a numeric vector.
trans_func	a function to transform x values before plotting. Useful to apply log10 transformation (i.e. with trans_func = function(x) log10(x+1)).
title	Title of the plot.

ylim limit of the y axis; format: ylim = c(min,max)
 xlab x-axis title

Value

Display a plot.

See Also

[plotEpistack](#), [plotBoxMetric](#)

Examples

```
data("stackepi")
plotMetric(stackepi$exp)
```

plotStackProfile *plotStackProfile()*

Description

Display a heatmap of an epigenetic track centered at genomic anchors such as Transcription Start Sites or peak center.

Usage

```
plotStackProfile(
  gr,
  pattern = "^window_",
  x_labels = c("Before", "Anchor", "After"),
  title = "",
  zlim = NULL,
  palette = function(n) viridisLite::viridis(n, direction = -1),
  target_height = 650,
  summary_func = function(x) mean(x, na.rm = TRUE),
  n_core = 1
)
```

Arguments

gr a GRanges input
 pattern a character vector of length 1 of a column prefixe (can be regular expressions) that should match columns of gr.
 x_labels a character vectors of length 3 used to label the x-axis.
 title The title of the heatmap

zlim	The minimum and maximum z values to match color to values. Format: zlim = c (min, max)
palette	a palette of color, (i.e. a function of parameter n that should return n colors).
target_height	The matrix height is reduced to this number of rows before plotting. Useful to limit overplotting artefacts. It should roughly be set to the pixel height in the final heatmap.
summary_func	function passed to <code>redimMatrix()</code> . Usually mean, but can be set to median or max for sparse matrices.
n_core	multicore option, passed to <code>redimMatrix()</code> .

Details

The visualisation is centered on an anchor, a set of genomic coordinates that can be transcription start sites or peak center for example. Anchor coordinates are those of the GRanges used as an input (hereafter `gr`).

Anchors are plotted from top to bottom in the same order as in `gr`. One should sort `gr` before plotting if needed.

The matrix used to display the heatmap should be passed as additional metadata columns of `gr`. Such matrix can be obtained using `EnrichedHeatmap::normalizeToMatrix()` for example. The matrix column names are then specified through `what_pattern` using a prefix, a suffix or a regular expressions.

This function scale reasonably well up to hundred thousands of regions. Overplotting issues are solved by last-minute reduction of the matrix size using `redimMatrix()`.

Value

Display a plot.

See Also

[plotAverageProfile](#), [plotEpistack](#), [normalizeToMatrix](#), [plotStackProfileLegend](#)

Examples

```
data("stackepi")
plotStackProfile(stackepi,
                 target_height = 650,
                 zlim = c(0, 1),
                 palette = colorRampPalette(c("white", "dodgerblue", "black")),
                 title = "DNA methylation")
```

plotStackProfileLegend
plotStackProfileLegend()

Description

Utility function to plot the corresponding legend key of plotStackProfile()'s plots.

Usage

```
plotStackProfileLegend(  
  zlim,  
  palette = colorRampPalette(c("white", "grey", "black")),  
  title = NA  
)
```

Arguments

zlim	the limits of the values to be displayed. Format: c(min,max)
palette	a palette of color, (i.e. a function of parameter n that should return n colors).
title	an optional title to be displayed below the color legend.

Value

Display a plot.

See Also

[plotStackProfile](#)

Examples

```
plotStackProfileLegend(zlim = c(0, 2),  
  palette = colorRampPalette(c("white", "grey", "black")))
```

redimMatrix *redimMatrix()*

Description

Reduce the input matrix size by applying a summary function on cells to be fused.

Usage

```
redimMatrix(
  mat,
  target_height = 100,
  target_width = 100,
  summary_func = function(x) mean(x, na.rm = TRUE),
  output_type = 0,
  n_core = 1
)
```

Arguments

<code>mat</code>	the input matrix.
<code>target_height</code>	height of the output matrix (should be smaller than or equal to <code>nrow(mat)</code>).
<code>target_width</code>	width of the output matrix (should be smaller than or equal to <code>ncol(mat)</code>).
<code>summary_func</code>	how to summarize cells? A function such as <code>mean</code> , <code>median</code> , <code>max</code> , or <code>meanColors</code> .
<code>output_type</code>	Type of the output, to be passed to <code>vapply</code> 's <code>FUN.VALUE</code> .
<code>n_core</code>	number of core to use for parallel processing.

Details

This function is used to reduce matrix right before plotting them in order to avoid overplotting issues as well as other plotting artefacts.

Value

a resized matrix of size `target_width` x `target_height` where the `summary_fun` was apply to adjacent cells.

See Also

[meanColor](#)

Examples

```
data("stackepi")
mat <- S4Vectors::mcols(stackepi)
whichCols <- grepl("^window_", colnames(mat))
mat <- as.matrix(mat[, whichCols])
dim(mat)
smallMat <- redimMatrix(mat, target_height = 10, target_width = ncol(mat))
dim(smallMat)

mat <- matrix(sample(1:40,100,replace=TRUE),nrow=10,ncol=10)
dim(mat)
smallMat <- redimMatrix(mat, target_height = 5, target_width = ncol(mat),
  summary_func = function(x) max(x, na.rm = TRUE))
dim(smallMat)
```

`stackepi`*epistack example and test dataset*

Description

DNA methylation profiles (from MBD-seq data) around transcription start sites of the 693 chr18 genes annotated on the pig genome (Sscrofa11.1), as well as gene expression levels in Transcript Per Million (TPM) measured by RNA-seq in the same duodenum sample.

Usage

```
data("stackepi")
```

Format

A GRanges of the 693 rows and 54 metadata columns

Source

This dataset was generated from ENSEMBL annotation data and data generated by our lab (publicly available soon).

Index

* datasets

stackepi, [15](#)

addBins, [2](#), [10](#)

addMetricAndArrangeGRanges, [2](#), [3](#), [10](#)

meanColor, [4](#), [14](#)

normalizeToMatrix, [10](#), [12](#)

par, [9](#)

plotAverageProfile, [5](#), [10](#), [12](#)

plotBinning, [6](#)

plotBoxMetric, [7](#), [11](#)

plotEpistack, [8](#), [11](#), [12](#)

plotMetric, [10](#)

plotStackProfile, [10](#), [11](#), [13](#)

plotStackProfileLegend, [12](#), [13](#)

redimMatrix, [4](#), [10](#), [13](#)

stackepi, [15](#)