

Introduction to RBM package

Dongmei Li

October 27, 2020

Clinical and Translational Science Institute, University of Rochester School of Medicine and
Dentistry, Rochester, NY 14642-0708

Contents

| | | |
|----------|--|----------|
| 1 | Overview | 1 |
| 2 | Getting started | 2 |
| 3 | RBM_T and RBM_F functions | 2 |
| 4 | Ovarian cancer methylation example using the RBM_T function | 6 |

1 Overview

This document provides an introduction to the RBM package. The RBM package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the RBM package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

2 Getting started

The RBM package can be installed and loaded through the following R code.
Install the RBM package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

3 RBM_T and RBM_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The p -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

| | Length | Class | Mode |
|---------------|--------|--------|---------|
| ordfit_t | 1000 | -none- | numeric |
| ordfit_pvalue | 1000 | -none- | numeric |
| ordfit_beta0 | 1000 | -none- | numeric |
| ordfit_beta1 | 1000 | -none- | numeric |
| permutation_p | 1000 | -none- | numeric |
| bootstrap_p | 1000 | -none- | numeric |

```
> sum(myresult$permutation_p<=0.05)
```

```

[1] 58

> which(myresult$permutation_p<=0.05)

[1] 3 11 16 18 27 38 42 46 63 84 86 93 113 114 129 133 142 163 197
[20] 229 245 259 299 309 332 354 391 394 397 404 427 444 458 470 479 508 527 534
[39] 600 608 638 652 669 677 678 703 722 755 820 836 840 864 877 894 907 943 996
[58] 998

> sum(myresult$bootstrap_p<=0.05)

[1] 12

> which(myresult$bootstrap_p<=0.05)

[1] 6 33 124 150 314 359 431 635 678 796 907 996

> permutation_adj_p <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adj_p<=0.05)

[1] 3

> bootstrap_adj_p <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adj_p<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 11

> which(myresult2$bootstrap_p<=0.05)

[1] 83 176 271 359 494 533 598 622 767 832 978

> bootstrap2_adj_p <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adj_p<=0.05)

[1] 0

```

- Examples using the `RBM_F` function: `normdata_F` simulates a standardized gene expression data and `unifdata_F` simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```
> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)
```

| | Length | Class | Mode |
|---------------|--------|--------|---------|
| ordfit_t | 3000 | -none- | numeric |
| ordfit_pvalue | 3000 | -none- | numeric |
| ordfit_beta1 | 3000 | -none- | numeric |
| permutation_p | 3000 | -none- | numeric |
| bootstrap_p | 3000 | -none- | numeric |

```
> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 52

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 58

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 56

> which(myresult_F$permutation_p[, 1]<=0.05)

[1] 10 33 60 71 90 103 104 105 113 129 207 208 211 223 226 238 239 275 287
[20] 293 334 394 395 400 432 437 471 481 485 488 534 543 548 551 562 566 611 654
[39] 657 664 665 672 706 720 733 777 791 815 836 923 967 980

> which(myresult_F$permutation_p[, 2]<=0.05)

[1] 10 33 60 71 94 100 103 104 105 113 129 180 205 208 211 226 239 247 275
[20] 287 293 306 328 387 394 400 414 432 437 471 485 488 497 500 528 534 543 551
[39] 562 579 611 615 626 639 654 664 672 720 791 793 815 832 836 923 945 960 964
[58] 980

> which(myresult_F$permutation_p[, 3]<=0.05)

[1] 5 10 33 60 71 76 100 103 104 105 113 129 208 211 226 238 239 275 287
[20] 293 306 334 355 387 394 400 432 437 471 485 488 497 528 534 543 548 551 562
[39] 611 626 657 664 672 706 720 777 791 793 815 832 836 923 960 964 967 980
```

```

> con1_adj_p <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adj_p<=0.05/3)

[1] 4

> con2_adj_p <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adj_p<=0.05/3)

[1] 2

> con3_adj_p <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adj_p<=0.05/3)

[1] 11

> which(con2_adj_p<=0.05/3)

[1] 226 239

> which(con3_adj_p<=0.05/3)

[1] 105 113 226 239 287 394 432 551 562 791 836

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

              Length Class  Mode
ordfit_t      3000   -none- numeric
ordfit_pvalue 3000   -none- numeric
ordfit_beta1  3000   -none- numeric
permutation_p 3000   -none- numeric
bootstrap_p   3000   -none- numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 57

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 50

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 55

```

```

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 14 22 40 70 80 85 96 102 109 128 132 168 184 192 211 247 259 303 335
[20] 397 407 416 427 430 444 456 480 501 521 523 542 554 569 611 614 648 654 661
[39] 680 700 731 739 757 761 762 784 841 842 844 856 861 890 928 929 936 956 981

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 14 22 40 70 85 109 128 132 168 184 192 210 247 259 303 327 391 397 407
[20] 416 427 430 444 456 499 501 521 523 542 554 569 611 614 648 654 661 680 731
[39] 739 783 784 841 842 844 856 880 890 893 936 981

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 2 14 22 53 68 70 80 85 96 109 128 168 184 192 211 247 303 327 391
[20] 397 407 415 416 427 430 444 456 480 499 501 521 523 542 554 569 611 614 648
[39] 654 661 680 731 739 761 783 784 841 842 844 861 890 936 954 965 981

> con21_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adj_p<=0.05/3)

[1] 3

> con22_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adj_p<=0.05/3)

[1] 3

> con23_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adj_p<=0.05/3)

[1] 5

```

4 Ovarian cancer methylation example using the RBM_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the gemone-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the RBM_T function and presenting the results for further validation and investigations.

```

> system.file("data", package = "RBM")

[1] "/private/tmp/RtmpOccJCc/Rinst1493265bd0050/RBM/data"

> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)

      IlmnID      Beta      exmdata2[, 2]      exmdata3[, 2]
cg00000292: 1   Min.    :0.01058   Min.    :0.01187   Min.    :0.009103
cg00002426: 1   1st Qu.:0.04111   1st Qu.:0.04407   1st Qu.:0.041543
cg00003994: 1   Median :0.08284   Median :0.09531   Median :0.087042
cg00005847: 1   Mean    :0.27397   Mean    :0.28872   Mean    :0.283729
cg00006414: 1   3rd Qu.:0.52135   3rd Qu.:0.59032   3rd Qu.:0.558575
cg00007981: 1   Max.    :0.97069   Max.    :0.96937   Max.    :0.970155
(Other)      :994                NA's     :4
exmdata4[, 2]      exmdata5[, 2]      exmdata6[, 2]      exmdata7[, 2]
Min.    :0.01019   Min.    :0.01108   Min.    :0.01937   Min.    :0.01278
1st Qu.:0.04092   1st Qu.:0.04059   1st Qu.:0.05060   1st Qu.:0.04260
Median :0.09042   Median :0.08527   Median :0.09502   Median :0.09362
Mean    :0.28508   Mean    :0.28482   Mean    :0.27348   Mean    :0.27563
3rd Qu.:0.57502   3rd Qu.:0.57300   3rd Qu.:0.52099   3rd Qu.:0.52240
Max.    :0.96658   Max.    :0.97516   Max.    :0.96681   Max.    :0.95974
                NA's     :1
exmdata8[, 2]
Min.    :0.01357
1st Qu.:0.04387
Median :0.09282
Mean    :0.28679
3rd Qu.:0.57217
Max.    :0.96268

> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)

      Length Class  Mode
ordfit_t      1000   -none- numeric
ordfit_pvalue 1000   -none- numeric
ordfit_beta0   1000   -none- numeric
ordfit_beta1   1000   -none- numeric
permutation_p 1000   -none- numeric
bootstrap_p    1000   -none- numeric

> sum(diff_results$ordfit_pvalue<=0.05)

[1] 45

```

```

> sum(diff_results$permutation_p<=0.05)

[1] 59

> sum(diff_results$bootstrap_p<=0.05)

[1] 44

> ordfit_adj_p <- p.adjust(diff_results$ordfit_pvalue, "BH")
> sum(ordfit_adj_p<=0.05)

[1] 0

> perm_adj_p <- p.adjust(diff_results$permutation_p, "BH")
> sum(perm_adj_p<=0.05)

[1] 4

> boot_adj_p <- p.adjust(diff_results$bootstrap_p, "BH")
> sum(boot_adj_p<=0.05)

[1] 0

> diff_list_perm <- which(perm_adj_p<=0.05)
> diff_list_boot <- which(boot_adj_p<=0.05)
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t)
> print(sig_results_perm)

      IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
83  cg00072216 0.04505377    0.04598964    0.04000674    0.03231534
103 cg00094319 0.73784280    0.73532960    0.75574900    0.73830220
627 cg00612467 0.04777553    0.03783457    0.05380982    0.05582291
764 cg00730260 0.90471270    0.90542290    0.91002680    0.91258610
      exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
83      0.04965089    0.04833366    0.03466159    0.04390894
103     0.67349260    0.73510200    0.75715920    0.78981220
627     0.04740551    0.05332965    0.05775211    0.05579710
764     0.90575890    0.88760470    0.90756300    0.90946790
      diff_results$ordfit_t[diff_list_perm]
83                                     2.514109
103                                    -2.268711
627                                    -2.239498
764                                    -1.808081
      diff_results$permutation_p[diff_list_perm]
83                                     0
103                                    0
627                                    0
764                                    0

```



```

> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t
> print(sig_results_boot)

[1] IlmnID
[2] Beta
[3] exmdata2[, 2]
[4] exmdata3[, 2]
[5] exmdata4[, 2]
[6] exmdata5[, 2]
[7] exmdata6[, 2]
[8] exmdata7[, 2]
[9] exmdata8[, 2]
[10] diff_results$ordfit_t[diff_list_boot]
[11] diff_results$bootstrap_p[diff_list_boot]
<0 rows> (or 0-length row.names)

```