## Package 'gsean'

October 17, 2020

**Description** Biological molecules in a living organism seldom work individually. They usually interact each other in a cooperative way. Biological process is too complicated to understand without considering such interactions. Thus, network-based procedures can be seen as powerful methods for studying complex process. However, many methods are devised for analyzing in-

Type Package

Title Gene Set Enrichment Analysis with Networks

dividual genes. It is said that techniques based on biological networks such as gene co- expression are more precise ways to represent information than those us-
ing lists of genes only. This package is aimed to integrate the gene expression and biological net-
work. A biological network is constructed from gene expres-
sion data and it is used for Gene Set Enrichment Analysis.
Version 1.8.0
<b>Date</b> 2019-04-29
Author Dongmin Jung
Maintainer Dongmin Jung <dmdmjung@gmail.com></dmdmjung@gmail.com>
<b>Depends</b> R (>= 3.5), fgsea, PPInfer
Suggests SummarizedExperiment, knitr, plotly, RANKS, WGCNA
License Artistic-2.0
<b>biocViews</b> Software, StatisticalMethod, Network, GraphAndNetwork, GeneSetEnrichment, GeneExpression, NetworkEnrichment, Pathways, DifferentialExpression
NeedsCompilation no
VignetteBuilder knitr
git_url https://git.bioconductor.org/packages/gsean
git_branch RELEASE_3_11
git_last_commit c5cd885
git_last_commit_date 2020-04-27
Date/Publication 2020-10-16
R topics documented:
gsean-package
centrality_gsea         2           exprs2adj         3

2 centrality\_gsea

	GO_dme	4
	gsean	5
	KEGG_hsa	6
	label_prop_gsea	7
Index		8

gsean-package

Gene Set Enrichment Analysis with Networks

#### **Description**

Biological molecules in a living organism seldom work individually. They usually interact each other in a cooperative way. Biological process is too complicated to understand without considering such interactions. Thus, network-based procedures can be seen as powerful methods for studying complex process. However, many methods are devised for analyzing individual genes. It is said that techniques based on biological networks such as gene co-expression are more precise ways to represent information than those using lists of genes only. This package is aimed to integrate the gene expression and biological network. A biological network is constructed from gene expression data and it is used for Gene Set Enrichment Analysis.

#### **Details**

The DESCRIPTION file: This package was not yet installed at build time.

Index: This package was not yet installed at build time.

#### Author(s)

Dongmin Jung

Maintainer: Dongmin Jung <dmdmjung@gmail.com>

centrality\_gsea

Gene Set Enrichment Analysis with centrality measure

## **Description**

GSEA is performed with centrality measure

#### Usage

exprs2adj 3

#### **Arguments**

geneset list of gene sets

x Named vector of gene-level statistics. Names should be the same as in gene sets.

adjacency adjacency matrix

pseudo pseudo number for log2 transformation (default: 1)

nperm number of permutations (default: 1000)

centrality centrality measure, degree centrality or node strength is default

weightParam weight parameter value for the centrality measure, equally weight if weight-

Param = 0 (default: 1)

minSize minimal size of a gene set (default: 1)
maxSize maximal size of a gene set (default: Inf)
gseaParam GSEA parameter value (default: 1)

nproc see fgsea::fgsea
BPPARAM see fgsea::fgsea

#### Value

GSEA result

#### Author(s)

Dongmin Jung

## See Also

fgsea::fgsea

## **Examples**

```
data(examplePathways)
data(exampleRanks)
exampleRanks <- exampleRanks[1:100]
adjacency <- diag(length(exampleRanks))
rownames(adjacency) <- names(exampleRanks)
set.seed(1)
result.GSEA <- centrality_gsea(examplePathways, exampleRanks, adjacency)</pre>
```

exprs2adj Convert gene expression data to adjacency matrix by using correlation

coefficients

## Description

A biological network is constructed from gene expression data and it is used for Gene Set Enrichment Analysis.

#### Usage

```
exprs2adj(x, pseudo = 1, ...)
```

4 GO\_dme

## **Arguments**

x gene expression data

pseudo pseudo number for log2 transformation (default: 1)
... additional parameters for correlation; see WGCNA::cor

## Value

adjacency matrix

#### Author(s)

Dongmin Jung

## See Also

```
fgsea::fgsea, WGCNA::cor
```

## **Examples**

```
data(exampleRanks)
Names <- names(exampleRanks)
exprs <- matrix(rnorm(10*length(exampleRanks)), ncol = 10)
adjacency <- exprs2adj(exprs)</pre>
```

GO\_dme

Gene Ontology terms with gene ID for Drosophila melanogaster

## Description

The data set contains all Gene Ontology terms for Drosophila melanogaster and genes are identified by gene ID. There are 2823 categories.

## Usage

GO\_dme

#### **Format**

a list of gene sets

#### Value

GO gene sets

## Author(s)

Dongmin Jung

## Source

http://www.go2msig.org/cgi-bin/prebuilt.cgi?taxid=7227

gsean 5

## **Examples**

```
load(system.file("data", "GO_dme.rda", package = "gsean"))
```

gsean

Gene Set Enrichment Analysis with Networks

## Description

GSEA or ORA is performed with networks from gene expression data

## Usage

```
gsean(geneset, x, exprs, pseudo = 1, threshold = 0.99, nperm = 1000,
    centrality = function(x) rowSums(abs(x)), weightParam = 1,
    minSize = 1, maxSize = Inf, gseaParam = 1, nproc = 0,
    BPPARAM = NULL, corParam = list(), tmax = 10, ...)
```

## Arguments

geneset	list of gene sets
x	Named vector of gene-level statistics for GSEA or set of genes for ORA. Names should be the same as in gene sets.
exprs	gene expression data
pseudo	pseudo number for log2 transformation (default: 1)
threshold	threshold of correlation for nodes to be considered neighbors for ORA (default: $0.99$ )
nperm	number of permutations (default: 1000)
centrality	centrality measure, degree centrality or node strength is default
weightParam	weight parameter value for the centrality measure, equally weight if weight-Param = $0$ (default: 1)
minSize	minimal size of a gene set (default: 1)
maxSize	maximal size of a gene set (default: Inf)
gseaParam	GSEA parameter value (default: 1)
nproc	see fgsea::fgsea
BPPARAM	see fgsea::fgsea
corParam	additional parameters for correlation; see WGCNA::cor
tmax	maximum number of iterations for label propagtion (default: 10)
	additional parameters for label propagation; see RANKS::label.prop

## Value

GSEA result

## Author(s)

Dongmin Jung

6 KEGG\_hsa

#### See Also

```
exprs2adj, label_prop_gsea, centrality_gsea
```

## **Examples**

```
data(examplePathways)
data(exampleRanks)
exampleRanks <- exampleRanks[1:100]
Names <- names(exampleRanks)
exprs <- matrix(rnorm(10*length(exampleRanks)), ncol = 10)
rownames(exprs) <- names(exampleRanks)
set.seed(1)
result.GSEA <- gsean(examplePathways, exampleRanks, exprs)</pre>
```

KEGG\_hsa

KEGG pathways with gene symbol for human

## **Description**

The data set contains 186 KEGG pathways for Drosophila melanogaster and genes are identified by gene symbol.

## Usage

KEGG\_hsa

## **Format**

a list of gene sets

#### Value

KEGG gene sets

## Author(s)

Dongmin Jung

## Source

http://software.broadinstitute.org/gsea/msigdb/collections.jsp

## **Examples**

```
load(system.file("data", "KEGG_hsa.rda", package = "gsean"))
```

label\_prop\_gsea 7

label\_prop\_gsea

Over-representaion analysis with the label propagation algorithm

## Description

ORA is performed by GSEA with the label propagation algorithm

## Usage

## **Arguments**

geneset	list of gene sets
X	set of genes
adjacency	adjacency matrix
threshold	threshold of correlation for nodes to be considered neighbors (default: 0.99)
nperm	number of permutations (default: 1000)
minSize	minimal size of a gene set (default: 1)
maxSize	maximal size of a gene set (default: Inf)
gseaParam	GSEA parameter value (default: 1)
nproc	see fgsea::fgsea
BPPARAM	see fgsea::fgsea
• • •	additional parameters for label propagation; see RANKS::label.prop

#### Value

GSEA result

## Author(s)

Dongmin Jung

## See Also

```
fgsea::fgsea, RANKS::label.prop
```

## **Examples**

```
data(examplePathways)
data(exampleRanks)
exampleRanks <- exampleRanks[1:100]
geneNames <- names(exampleRanks)
set.seed(1)
x <- sample(geneNames, 10)
adjacency <- diag(length(exampleRanks))
rownames(adjacency) <- geneNames
result.GSEA <- label_prop_gsea(examplePathways, x, adjacency)</pre>
```

# Index

```
centrality_gsea, 2
exprs2adj, 3

G0_dme, 4
gsean, 5
gsean-package, 2

KEGG_hsa, 6

label_prop_gsea, 7
```