# Package 'globalSeq'

October 17, 2020
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Title Global Test for Counts
<b>Description</b> The method may be conceptualised as a test of overall significance in regression analysis, where the response variable is overdispersed and the number of explanatory variables exceeds the sample size. Useful for testing for association between RNA-Seq and high-dimensional data.
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globalSeq-package

Negative binomial global test

## **Description**

Testing for association between RNA-Seq and other genomic data is challenging due to high variability of the former and high dimensionality of the latter.

Using the negative binomial distribution and a random effects model, we developed an omnibus test that overcomes both difficulties. It may be conceptualised as a test of overall significance in regression analysis, where the response variable is overdispersed and the number of explanatory variables exceeds the sample size.

The proposed method can detect genetic and epigenetic alterations that affect gene expression. It can examine complex regulatory mechanisms of gene expression.

## **Getting started**

```
omnibus tests entire covariate sets
proprius shows individual contributions
cursus analyses the whole genome
```

The following command opens the vignette: utils::vignette("globalSeq")

#### More information

A Rauschenberger, MA Jonker, MA van de Wiel, and RX Menezes (2016). "Testing for association between RNA-Seq and high-dimensional data", *BMC Bioinformatics*. 17:118. html pdf (open access)

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cursus

Genome-wide analysis

## **Description**

This function tests for associations between gene expression or exon abundance (Y) and genetic or epigenetic alterations (X). Using the locations of genes (Yloc), and the locations of genetic or epigenetic alterations (Xloc), the expression of each gene is tested for associations with alterations on the same chromosome that are closer to the gene than a given distance (window).

## Usage

```
cursus(Y, Yloc, X, Xloc, window,
    Ychr = NULL, Xchr = NULL,
    offset = NULL, group = NULL,
    perm = 1000, nodes = 2,
    phi = NULL, kind = 0.01)
```

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#### **Arguments**

Y **RNA-Seq data:** numeric matrix with q rows (genes) and n columns (samples); or a SummarizedExperiment object

Yloc **location RNA-Seq:** numeric vector of length q (point location); numeric matrix

with q rows and two columns (start and end locations)

X **genomic profile:** numeric matrix with p rows (covariates) and n columns (sam-

ples)

Xloc **location covariates:** numeric vector of length p
window **maximum distance:** non-negative real number
Ychr chromosome RNA-Seq: factor of length q
Xchr chromosome covariates: factor of length p

offset numeric vector of length n

group confounding variable: factor of length n
perm number of iterations: positive integer

nodes number of cluster nodes for parallel computation

phi dispersion parameters: vector of length q kind computation: number between 0 and 1

#### **Details**

Note that Yloc, Xloc and window must be given in the same unit, usually in base pairs. If Yloc indicates interval **locations**, and window is zero, then only covariates between the start and end location of the gene are of interest. Typically window is larger than one million base pairs.

If Y and X include data from a single chromosome, Ychr and Xchr are redundant. If Y or X include data from **multiple chromosomes**, Ychr and Xchr should be specified in order to prevent confusion between chromosomes.

For the simultaneous analysis of **multiple genomic profiles** X should be a list of numeric matrices with n columns (samples), Xloc a list of numeric vectors, and window a list of non-negative real numbers. If provided, Xchr should be alist of numeric vectors.

The offset is meant to account for different **libary sizes**. By default the offset is calculated based on Y. Different library sizes can be ignored by setting the offset to rep(1,n).

The user can provide the **confounding** variable group. Note that each level of group must appear at least twice in order to allow stratified permutations.

Efficient alternatives to classical **permutation** (kind=1) are the method of control variates (kind=0) and permutation in chunks (0 < kind < 1) details.

#### Value

The function returns a dataframe, with the p-values in the first row and the test statistics in the second row.

## References

A Rauschenberger, MA Jonker, MA van de Wiel, and RX Menezes (2016). "Testing for association between RNA-Seq and high-dimensional data", *BMC Bioinformatics*. 17:118. html pdf (open access)

RX Menezes, M Boetzer, M Sieswerda, GJB van Ommen, and JM Boer (2009). "Integrated analysis of DNA copy number and gene expression microarray data using gene sets", *BMC Bioinformatics*. 10:203. html pdf (open access)

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#### See Also

The function omnibus tests for associations between an overdispersed response variable and a high-dimensional covariate set. The function proprius calculates the contributions of individual samples or covariates to the test statistic. All other function of the R package globalSeq are internal.

## **Examples**

omnibus

Omnibus test

## **Description**

Test of association between a count response and one or more covariate sets. This test may be conceptualised as a test of overall significance in regression analysis, where the response variable is overdispersed, and where the number of explanatory variables (p) exceeds the sample size (n). The negative binomial distribution accounts for overdispersion and a random effect model accounts for high dimensionality (p»n).

## Usage

```
omnibus(y, X, offset = NULL, group = NULL,
    mu = NULL, phi = NULL,
    perm = 1000, kind = 1)
```

#### **Arguments**

У	response variable: numeric vector of length n
X	<ul><li>one covariate set: numeric matrix with n rows (samples) and p columns (covariates);</li><li>multiple covariate sets: list of numeric matrices with n rows (samples)</li></ul>
offset	numeric vector of length n
group	confounding variable: factor of length n
mu	mean parameters: numeric vector of length 1 or n
phi	dispersion parameter: non-negative real number
perm	number of iterations: positive integer
kind	computation: number between 0 and 1

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#### **Details**

The user can provide a common mu for all samples or sample-specific mu, and a common phi. Setting phi equal to zero is equivalent to using the Poisson model. If mu is missing, then mu is estimated from y. If phi is missing, then mu and phi are estimated from y. The offset is only taken into account for estimating mu or phi. By default the offset is rep(1,n).

The user can provide the **confounding** variable group. Note that each level of group must appear at least twice in order to allow stratified permutations.

Efficient alternatives to classical **permutation** (kind=1) are the method of control variates (kind=0) and permutation in chunks (0 < kind < 1) details.

#### Value

The function returns a dataframe, with the p-value in the first column, and the test statistic in the second column.

#### References

A Rauschenberger, MA Jonker, MA van de Wiel, and RX Menezes (2016). "Testing for association between RNA-Seq and high-dimensional data", *BMC Bioinformatics*. 17:118. html pdf (open access)

RX Menezes, L Mohammadi, JJ Goeman, and JM Boer (2016). "Analysing multiple types of molecular profiles simultaneously: connecting the needles in the haystack", *BMC Bioinformatics*. 17:77. html pdf (open access)

S le Cessie, and HC van Houwelingen (1995). "Testing the fit of a regression model via score tests in random effects models", *Biometrics*. 51:600-614. html pdf (restricted access)

#### See Also

The function proprius calculates the contributions of individual samples or covariates to the test statistic. The function cursus tests for association between RNA-Seq and local genetic or epigenetic alternations across the whole genome. All other functions of the R package globalSeq are internal.

#### **Examples**

```
# simulate high-dimensional data
n <- 30; p <- 100
y <- rnbinom(n,mu=10,size=1/0.25)
X <- matrix(rnorm(n*p),nrow=n,ncol=p)
# hypothesis testing
omnibus(y,X)</pre>
```

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proprius	Decomposition		
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## **Description**

Even though the function omnibus tests a single hypothesis on a whole covariate set, this function allows to calculate the individual contributions of n samples or p covariates to the test statistic.

## Usage

## **Arguments**

у	response variable: numeric vector of length n
Χ	covariate set: numeric matrix with n rows (samples) and p columns (covariates)
type	character 'covariates' or 'samples'
offset	numeric vector of length n
group	confounding variable: factor of length n
mu	mean parameters: numeric vector of length 1 or n
phi	dispersion parameter: non-negative real number
alpha	significance level: real number between 0 and 1
perm	number of iterations: positive integer
plot	plot of results: logical

#### Details

The user can provide a common mu for all samples or sample-specific mu, and a common phi. Setting phi equal to zero is equivalent to using the Poisson model. If mu is missing, then mu is estimated from y. If phi is missing, then mu and phi are estimated from y. The offset is only taken into account for estimating mu or phi.

The user can provide the confounding variable group. Note that each level of group must appear at least twice in order to allow stratified permutations.

## Value

If alpha=NULL, then the function returns a numeric vector, and else a list of numeric vectors.

## References

A Rauschenberger, MA Jonker, MA van de Wiel, and RX Menezes (2016). "Testing for association between RNA-Seq and high-dimensional data", *BMC Bioinformatics*. 17:118. html pdf (open access)

JJ Goeman, SA van de Geer, F de Kort, and HC van Houwelingen (2004). "A global test for groups of genes: testing association with a clinical outcome", *Bioinformatics*. 20:93-99. html pdf (open access)

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#### See Also

The function omnibus tests for associations between an overdispersed response variable and a high-dimensional covariate set. The function cursus tests for association between RNA-Seq and local genetic or epigenetic alternations across the whole genome. All other functions of the R package globalSeq are internal.

## **Examples**

```
# simulate high-dimensional data
n <- 30; p <- 100
y <- rnbinom(n,mu=10,size=1/0.25)
X <- matrix(rnorm(n*p),nrow=n,ncol=p)
# decomposition
proprius(y,X,type="samples")
proprius(y,X,type="covariates")</pre>
```

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