

# QTL Mapping using Diversity Outbred Mice

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## 1 Introduction

Quantitative Trait Locus (QTL) mapping in DO mice is performed in several steps. First, we use the founder haplotype contributions to perform linkage mapping. In the mapping model, we adjust for kinship between DO mice using the R package QTLRel. Then, we perform permutations to determine and empirical significance threshold. Next, we select chromosomes with QTL peaks above the significance threshold, examine the founder allele effects and determine support intervals. Finally, we impute the founder SNPs onto the DO genomes to perform association mapping in the QTL intervals.

## 2 Mapping Models

### 2.1 Linkage Mapping

Linkage mapping involves the use of founder haplotype probabilities. We perform point mapping at each marker on the array. We fit an additive model that regresses the phenotype on the eight founder haplotype contributions and incorporates an adjustment for the kinship between samples.

$$y = X\alpha + H\beta + Zu + \varepsilon \tag{1}$$

where:

- $n$  is the number of samples
- $y$  is an  $n \times 1$  vector of phenotype values for each sample
- $X$  is an  $n \times p$  matrix of  $p$  fixed covariates (sex, diet, etc.)
- $\alpha$  is a  $p \times 1$  vector of fixed effects
- $H$  is an  $n \times 8$  matrix of founder haplotype contributions (each row sums to 1)
- $\beta$  is an  $8 \times 1$  vector of founder haplotype effects
- $Z$  is an  $n \times n$  matrix of error covariances between samples
- $u$  is an  $n \times 1$  vector of ???
- $\varepsilon$  is an  $n \times 1$  vector of residual errors

## 2.2 Association Mapping

Between each pair of markers, we assign the genotype state with the highest probability to each DO sample. We then query the Sanger Mouse Genomes SNP file to obtain all of the founder SNPs in the interval.

For each Sanger SNP, we impute the Sanger SNPs onto DO genomes as follows:

$$a_j = \sum_{i=1}^8 s_i h_{ij} \quad (2)$$

where:

- $a$  is the allele call (coded as 0, 1 or 2) for sample  $j$
- $s$  is the Sanger founder allele call (coded as 0 or 1)
- $h$  is the founder haplotype contribution of founder  $i$  for sample  $j$

$$y = X\alpha + A\beta + Zu + \varepsilon \quad (3)$$

where:

- $n$  is the number of samples
- $y$  is an  $n \times 1$  vector of phenotype values for each sample
- $X$  is an  $n \times p$  matrix of  $p$  fixed covariates (sex, diet, etc.)
- $\alpha$  is a  $p \times 1$  vector of fixed effects
- $A$  is an  $n \times 3$  matrix of imputed allele calls
- $\beta$  is an  $3 \times 1$  vector of allele effects
- $Z$  is an  $n \times n$  matrix of error covariances between samples
- $u$  is an  $n \times 1$  vector of ???
- $\varepsilon$  is an  $n \times 1$  vector of residual errors

## 3 QTL Mapping

We will use example data from Svenson et.al, *Genetics*, 2012. Briefly, 149 mice (75 F, 74 M) were placed on either a chow ( $n = 100$ ) or a high fat diet ( $n = 49$ ). A variety of clinical phenotypes were measured at two time points, roughly 14 weeks apart. In this example, we will map the hemoglobin distribution width (HDW) at the second time point. We will load this data from the Bioconductor data package `MUGAExampleData`.

```
> library(DOQTL)
> library(MUGAExampleData)
> data(pheno)
> data(model.probs)
```

QTL mapping requires phenotype and genotype data. Here, we have a `data.frame` of phenotypes called `pheno` and a 3D array of founder haplotype contributions (num.samples x 8 founders x num.markers) called `model.probs`. The sample IDs must be in `rownames(pheno)` and `dimnames(model.probs)[[1]]` and they must match each other. We will map the hemoglobin distribution width at time point 2 (HDW2).

First, we need to create a kinship matrix using the founder contributions.

```
> K = kinship.probs(model.probs)
```

Second, we need to create a matrix of additive covariates to run in the model. In this case, we will use sex, diet and CHOL1. Note that the sample IDs must be in `rownames(covar)`.

```
> covar = data.frame(sex = as.numeric(pheno$Sex == "M"), diet = as.numeric(pheno$Diet == "hf"))
> rownames(covar) = rownames(pheno)
```

Third, we need to get the marker locations on the array.

```
> load(url("ftp://ftp.jax.org/MUGA/muga_snps.Rdata"))
```

Fourth, we map the phenotype using `scanone`.

```
> qtl = scanone(pheno = pheno, pheno.col = "HDW2", probs = model.probs, K = K,
+               addcovar = covar, snps = muga_snps)
```

```
[1] "HDW2"
```

Fifth, we run permutations to determine significance thresholds. We recommend running at least 1,000 permutations. In this demo, we run 100 permutations to save time.

```
> perms = scanone.perm(pheno = pheno, pheno.col = "HDW2", probs = model.probs,
+                      addcovar = covar, snps = muga_snps, nperm = 100)
> thr = quantile(perms, probs = 0.95)
```

We then plot the LOD curve for the QTL.

```
> plot(qtl, sig.thr = thr, main = "HDW2")
```

The largest peak appears on Chr 9. The linkage mapping model (Eqn. 1) produces an estimate of the effect of each founder allele at each marker. We can plot these effects (model coefficients) on Chr 9 to see which founders contribute to a high HDW.

```
> coefplot(qtl, chr = 9)
```

Note that the DO mice with alleles from three strains, 129S1/SvImJ, NZO/HILtJ and WSB/EiJ, have lower changes in cholesterol than the other five strains. Remember these strains because they will appear again below. We then determine the width of the QTL support interval using `bayesint`. Note that this

function only provides reasonable support intervals if there is a single QTL on the chromosome.

```
> interval = bayesint(qtl, chr = 9)
> interval
```

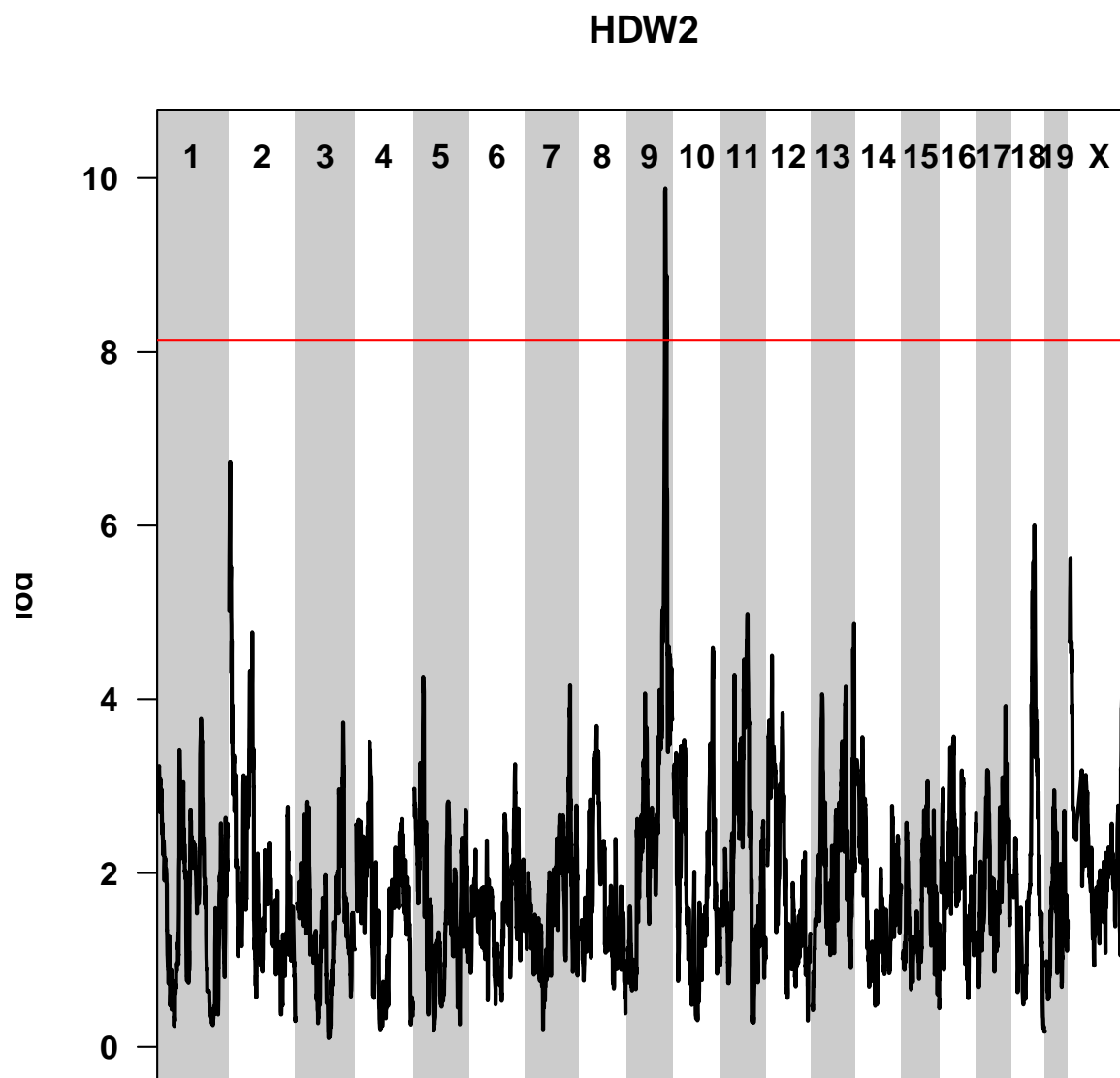


Figure 1: QTL plot of HDW2. The LOD of the mode in Eqn. 1 is plotted along the mouse genome. The red line is the  $p < 0.05$  significance threshold.

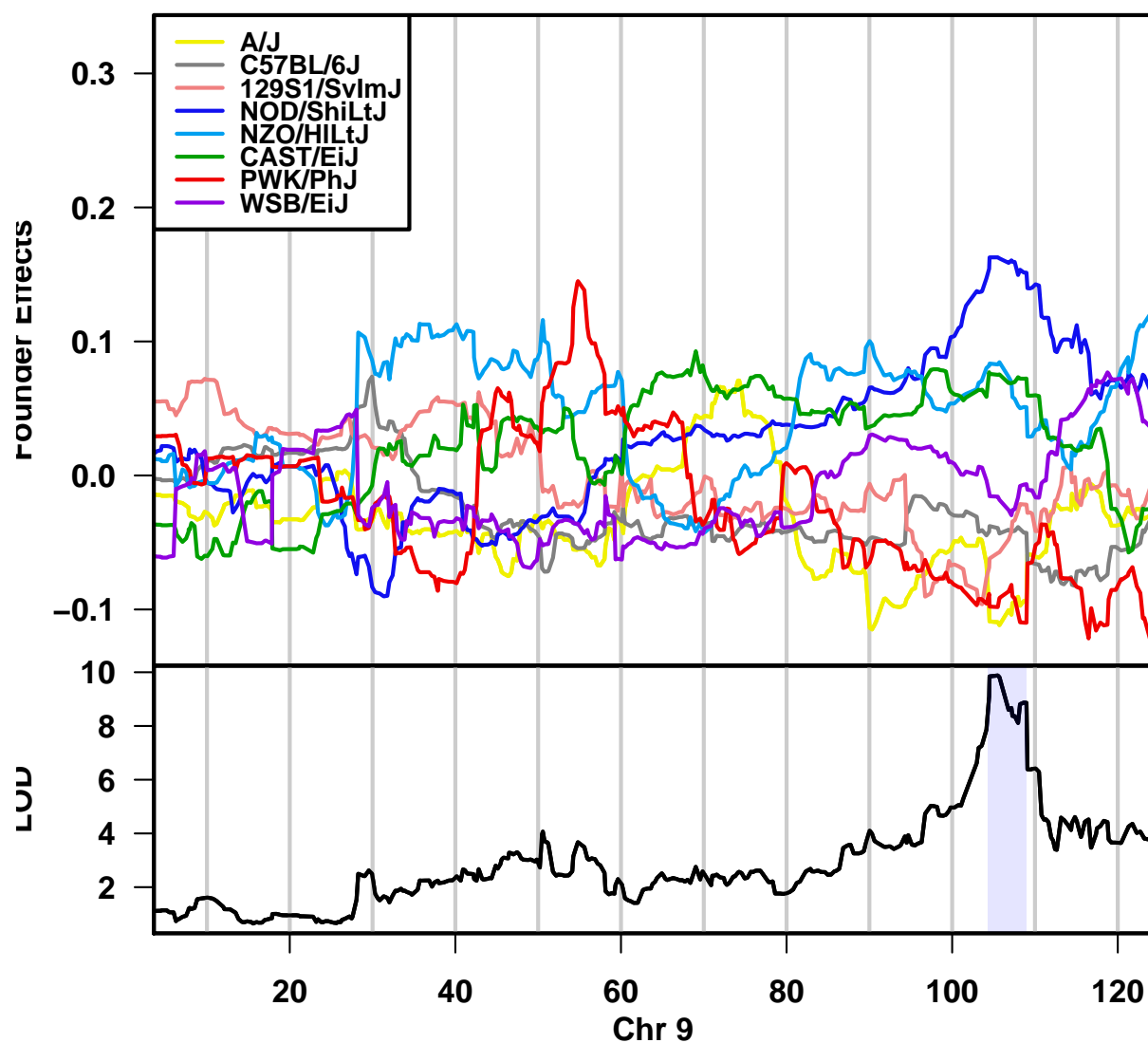


Figure 2: Coefficient plot of HDW2 on Chr 9. The top panel shows the 8 estimated founder allele effects along Chr 9. The NOD/ShiLtJ allele contributes to high values and the A/J and PWK/PhJ alleles contribute to low values. The bottom panel shows the LOD score.

	SNP_ID	Chr	Mb_NCBI38	cM	perc.var	lrs	lod
1	<NA>	9	104.3100	56.6120	25.16321	39.16096	8.503694
4105	UNC091160886	9	105.5128	56.7432	28.60966	45.49603	9.879338
3	<NA>	9	108.9811	59.6310	25.69325	40.11901	8.711732

	p	neg.log10.p
1	<NA>	9
4105	1.09550863036677e-07	6.96038419673778
3	<NA>	9

The QTL support interval is 4.7 Mb wide. Finally, we narrow the candidate gene list by imputing

the founder SNPs onto the DO genomes. This idea is essentially association mapping in an outbred population.

```
> ma = assoc.map(pheno = pheno, pheno.col = "HDW2", probs = model.probs, K = K,
+               addcovar = covar, snps = muga_snps, chr = interval[1,2],
+               start = interval[1,3], end = interval[3,3])
```

```
[1] "Mapping with 135 samples."
[1] "Retrieving SNPs..."
[1] "Retrieved 66750 SNPs."
[1] "Finding unique SNP patterns..."
[1] "Calculating mapping statistic"
```

```
> tmp = assoc.plot(ma, thr = 4)
> unique(tmp$sdps)
```

```
[1] 16 20 60 24
```

We can get the genes in the QTL interval using the `get.mgi.features()` function.

```
> mgi = get.mgi.features(chr = interval[1,2], start = interval[1,3],
+                       end = interval[3,3], type = "gene", source = "MGI")
> nrow(mgi)
```

```
[1] 168
```

```
> head(mgi)
```

	seqid	source	type	start	stop	score	strand	phase	ID
1	9	MGI	gene	104288240	104337728	.	-	.	MGI:MGI:1928480
135	9	MGI	gene	104426113	104426187	.	+	.	MGI:MGI:4358922
142	9	MGI	gene	104481368	104481510	.	-	.	MGI:MGI:5455681
146	9	MGI	gene	104547286	105034544	.	+	.	MGI:MGI:1921270
305	9	MGI	gene	104994916	104995019	.	+	.	MGI:MGI:5453168
379	9	MGI	gene	105053239	105077476	.	+	.	MGI:MGI:2137204

	Name	Parent
1	Acpp	NA
135	Mir2136	NA
142	Gm25904	NA
146	Cpne4	NA
305	Gm23391	NA

```

[1] "Mapping with 135 samples."
[1] "Retrieving SNPs..."
[1] "Retrieved 66750 SNPs."
[1] "Finding unique SNP patterns..."
[1] "Calculating mapping statistic"

```

```

[1] 16 20 60 24

```

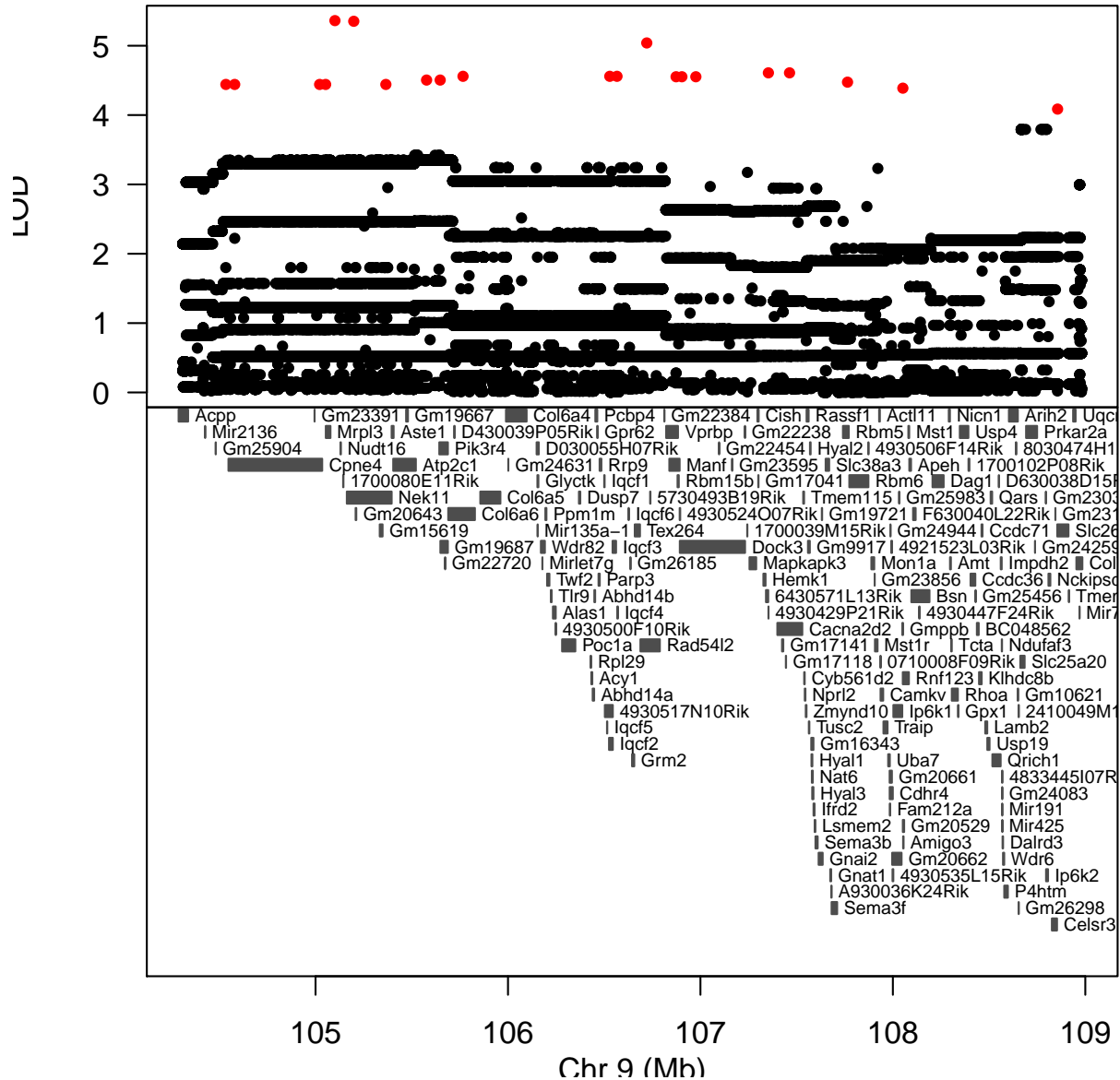


Figure 3: Association mapping plot of HDW2 in the Chr 9 support interval. The top panel shows the LOD score from association mapping (Eqn. 3) in the QTL support interval. The bottom panel shows the genes and non-coding RNAs from the Mouse Genome Informatics database.

```

379 Mrpl3      NA
                                           Dbxref
1  VEGA:OTTMUSG00000024988,NCBI_Gene:56318,ENSEMBL:ENSMUSG00000032561
135                                NCBI_Gene:100316725,ENSEMBL:ENSMUSG00000089406
142                                ENSEMBL:ENSMUSG00000089116
146 VEGA:OTTMUSG00000023466,NCBI_Gene:74020,ENSEMBL:ENSMUSG00000032564
305                                ENSEMBL:ENSMUSG00000088204
379 VEGA:OTTMUSG00000023521,NCBI_Gene:94062,ENSEMBL:ENSMUSG00000032563
      mgiName      bioType
1      acid phosphatase%2c prostate protein coding gene\r
135      microRNA 2136      miRNA gene\r
142      predicted gene%2c 25904      snoRNA gene\r
146      copine IV protein coding gene\r
305      predicted gene%2c 23391      miRNA gene\r
379 mitochondrial ribosomal protein L3 protein coding gene\r

```

There are 169 genes in the QTL support interval. Several SNPs have LOD scores above 4. This is a somewhat arbitrary cutoff and an appropriate threshold will be supplied in future version of DOQTL. In this case, there may be more than one variant that influences the phenotype.

## 4 SessionInfo

```

> sessionInfo()

R version 3.2.0 RC (2015-04-08 r68161)
Platform: x86_64-apple-darwin13.4.0 (64-bit)
Running under: OS X 10.9.5 (Mavericks)

locale:
[1] C/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8

attached base packages:
[1] stats      graphics  grDevices  utils      datasets  methods    base

other attached packages:
[1] MUGAExampleData_0.101.1 DOQTL_1.2.0      RSQLite_1.0.0
[4] DBI_0.3.1

loaded via a namespace (and not attached):
 [1] AnnotationDbi_1.30.0  XVector_0.8.0      zlibbioc_1.14.0
 [4] GenomicRanges_1.20.0 BiocGenerics_0.14.0 annotationTools_1.42.0
 [7] IRanges_2.2.0        mclust_5.0.0       xtable_1.7-4
[10] org.Hs.eg.db_3.1.2    hwriter_1.3.2      GenomeInfoDb_1.4.0
[13] tools_3.2.0          parallel_3.2.0     QTLRel_0.2-14
[16] Biobase_2.28.0        corpcor_1.6.7      gtools_3.4.2
[19] org.Mm.eg.db_3.1.2    S4Vectors_0.6.0    bitops_1.0-6
[22] RUnit_0.4.28          RCurl_1.95-4.5     biomaRt_2.24.0
[25] gdata_2.13.3          Biostrings_2.36.0  Rsamtools_1.20.0
[28] stats4_3.2.0          XML_3.98-1.1       annotate_1.46.0

```