

Introduction to RBM package

Dongmei Li

April 16, 2015

Clinical and Translational Science Institute, University of Rochester School of Medicine and
Dentistry, Rochester, NY 14642-0708

Contents

1 Overview	1
2 Getting started	2
3 RBM_T and RBM_F functions	2
4 Ovarian cancer methylation example using the RBM_T function	6

1 Overview

This document provides an introduction to the RBM package. The RBM package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the RBM package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

2 Getting started

The RBM package can be installed and loaded through the following R code.
Install the RBM package with:

```
> source("http://bioconductor.org/biocLite.R")
> biocLite("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

3 RBM_T and RBM_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The p -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(myresult$permutation_p<=0.05)
```

```
[1] 19
```

```

> which(myresult$permutation_p<=0.05)

[1] 24 57 180 193 200 204 238 434 482 483 537 539 673 734 782 817 847 848 895

> sum(myresult$bootstrap_p<=0.05)

[1] 3

> which(myresult$bootstrap_p<=0.05)

[1] 128 242 618

> permutation_adj_p <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adj_p<=0.05)

[1] 0

> bootstrap_adj_p <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adj_p<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 30

> which(myresult2$bootstrap_p<=0.05)

[1] 54 65 105 146 160 190 194 201 214 229 284 358 380 386 437 462 568 588 604
[20] 655 656 667 717 842 862 863 922 926 943 988

> bootstrap2_adj_p <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adj_p<=0.05)

[1] 0

```

- Examples using the RBM_F function: normdata_F simulates a standardized gene expression data and unifdata_F simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```

> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)

              Length Class  Mode
ordfit_t      3000   -none-  numeric
ordfit_pvalue 3000   -none-  numeric
ordfit_beta1   3000   -none-  numeric
permutation_p 3000   -none-  numeric
bootstrap_p    3000   -none-  numeric

> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 64

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 55

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 52

> which(myresult_F$permutation_p[, 1]<=0.05)

[1] 28 49 64 65 77 92 117 121 140 150 171 174 193 195 197 258 277 287 324
[20] 325 335 355 356 367 378 385 421 431 437 454 472 476 509 557 581 582 599 614
[39] 620 630 697 706 730 738 745 767 781 800 805 807 810 827 834 839 852 873 880
[58] 887 893 908 920 923 950 977

> which(myresult_F$permutation_p[, 2]<=0.05)

[1] 28 49 65 92 117 121 150 171 174 193 195 197 258 287 324 335 355 367 385
[20] 421 437 454 472 476 509 557 581 582 614 620 647 675 715 730 738 740 745 765
[39] 767 781 800 807 810 819 834 839 852 873 880 887 908 920 923 967 977

> which(myresult_F$permutation_p[, 3]<=0.05)

[1] 28 49 77 92 117 121 150 171 174 193 194 195 197 258 324 335 355 367 378
[20] 385 421 437 454 476 557 581 582 614 706 715 730 738 745 767 781 800 805 807
[39] 809 810 819 827 834 839 852 873 880 885 908 923 967 977

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

[1] 13

```

```

> con2_adj_p <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adj_p<=0.05/3)

[1] 12

> con3_adj_p <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adj_p<=0.05/3)

[1] 10

> which(con2_adj_p<=0.05/3)

[1] 28 92 150 258 355 367 454 614 730 745 839 908

> which(con3_adj_p<=0.05/3)

[1] 92 150 454 614 730 839 873 880 908 923

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

      Length Class  Mode
ordfit_t      3000   -none- numeric
ordfit_pvalue 3000   -none- numeric
ordfit_beta1  3000   -none- numeric
permutation_p 3000   -none- numeric
bootstrap_p    3000   -none- numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 48

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 57

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 42

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 6 16 38 58 75 97 109 134 168 177 181 190 200 210 235 248 268 289 427
[20] 484 489 513 542 544 577 606 631 637 660 678 707 728 739 763 785 812 820 845
[39] 880 885 914 918 930 941 944 953 956 979

```

```

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 2 6 16 38 75 86 97 109 119 133 134 163 177 181 190 200 232 248 262
[20] 268 289 360 427 439 484 489 513 542 577 604 606 637 660 667 678 707 708 716
[39] 728 729 739 767 775 785 812 820 824 845 880 881 911 918 930 941 953 979 982

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 6 16 38 75 86 97 109 134 177 181 190 200 210 248 252 268 289 427 486
[20] 489 513 542 577 604 606 637 660 678 728 739 767 785 820 845 885 914 918 930
[39] 941 944 953 979

> con21_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adj_p<=0.05/3)

[1] 3

> con22_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adj_p<=0.05/3)

[1] 3

> con23_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adj_p<=0.05/3)

[1] 2

```

4 Ovarian cancer methylation example using the RBM_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the RBM_T function and presenting the results for further validation and investigations.

```

> system.file("data", package = "RBM")

[1] "/private/tmp/RtmpRJzS1r/Rinst31bf31d0f3a2/RBM/data"

> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)

```

IlmnID		case1	case2	control1
cg00000292:	1	Min. :0.01058	Min. :0.01138	Min. :0.009103
cg00002426:	1	1st Qu.:0.04111	1st Qu.:0.04290	1st Qu.:0.041543
cg00003994:	1	Median :0.08284	Median :0.10438	Median :0.087042
cg00005847:	1	Mean :0.27397	Mean :0.29086	Mean :0.283729
cg00006414:	1	3rd Qu.:0.52135	3rd Qu.:0.54436	3rd Qu.:0.558575
cg00007981:	1	Max. :0.97069	Max. :0.96901	Max. :0.970155
(Other)	:994			
		case3	case4	control3
control2				
Min.	:0.01019	Min. :0.01108	Min. :0.009753	Min. :0.01278
1st Qu.:	0.04092	1st Qu.:0.04059	1st Qu.:0.041818	1st Qu.:0.04260
Median	:0.09042	Median :0.08527	Median :0.092807	Median :0.09362
Mean	:0.28508	Mean :0.28482	Mean :0.283113	Mean :0.27563
3rd Qu.:	0.57502	3rd Qu.:0.57300	3rd Qu.:0.558211	3rd Qu.:0.52240
Max.	:0.96658	Max. :0.97516	Max. :0.963620	Max. :0.95974
		NA's :1	NA's :1	
control4				
Min.	:0.01357			
1st Qu.:	0.04387			
Median	:0.09282			
Mean	:0.28679			
3rd Qu.:	0.57217			
Max.	:0.96268			

```

> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)

```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```

> sum(diff_results$ordfit_pvalue<=0.05)

```

```

[1] 31

```

```

> sum(diff_results$permutation_p<=0.05)

```

```

[1] 62

```

```

> sum(diff_results$bootstrap_p<=0.05)

```

```
[1] 50
```

```
> ordfit_adjp <- p.adjust(diff_results$ordfit_pvalue, "BH")
> sum(ordfit_adjp<=0.05)
```

```
[1] 0
```

```
> perm_adjp <- p.adjust(diff_results$permutation_p, "BH")
> sum(perm_adjp<=0.05)
```

```
[1] 14
```

```
> boot_adjp <- p.adjust(diff_results$bootstrap_p, "BH")
> sum(boot_adjp<=0.05)
```

```
[1] 1
```

```
> diff_list_perm <- which(perm_adjp<=0.05)
> diff_list_boot <- which(boot_adjp<=0.05)
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t)
> print(sig_results_perm)
```

	IlmnID	case1	case2	control1	control2	case3
66	cg00059424	0.02742616	0.02554150	0.03049395	0.02910234	0.02547771
76	cg00065408	0.03952223	0.03967472	0.04799694	0.04929252	0.04064262
110	cg00098239	0.02698720	0.02142180	0.01856646	0.01934917	0.02510008
129	cg00121158	0.03045297	0.02728770	0.03573820	0.03316130	0.02853104
245	cg00224508	0.04479948	0.04477529	0.04152814	0.04189373	0.04208405
396	cg00393585	0.06725091	0.07328911	0.09169522	0.07633878	0.06005587
432	cg00419564	0.03638860	0.03661916	0.04101457	0.04065540	0.03283922
460	cg00445824	0.14782870	0.16655800	0.14393210	0.13479670	0.20038750
660	cg00634577	0.03182804	0.03432180	0.03525499	0.03612398	0.03384897
690	cg00661202	0.01639344	0.01586308	0.01876500	0.02097005	0.01490915
764	cg00730260	0.90471270	0.90207400	0.91002680	0.91258610	0.90575890
772	cg00743372	0.03922780	0.03499011	0.02187972	0.02568053	0.02796053
906	cg00886554	0.06788907	0.06822703	0.06576263	0.05319272	0.07086407
908	cg00887547	0.06519115	0.08241367	0.08613525	0.08758982	0.07862745
	case4	control3	control4	diff_results\$ordfit_t[diff_list_perm]		
66	0.02523713	0.04478458	0.03391813			-2.203752
76	0.03622954	0.04778213	0.04327211			-3.115264
110	0.02291811	0.01784160	0.02109290			1.976098
129	0.02993313	0.03318921	0.03345607			-2.118418
245	0.05731476	0.03775905	0.03955271			1.811314
396	0.05180907	0.11621550	0.07849753			-2.839104
432	0.03934095	0.04413387	0.04462037			-2.407553
460	0.16185300	0.11630830	0.13912630			2.993440
660	0.03062112	0.03502489	0.03710039			-1.402274


```

690 0.01764273 0.01847447 0.01803320 -1.267423
764 0.90290550 0.90756300 0.90946790 -2.477349
772 0.03001808 0.02575992 0.02093909 2.824452
906 0.07034121 0.06106147 0.06624816 2.175684
908 0.06954336 0.09504429 0.08221681 -2.812745

```

```

diff_results$permutation_p[diff_list_perm]
66 0
76 0
110 0
129 0
245 0
396 0
432 0
460 0
660 0
690 0
764 0
772 0
906 0
908 0

```

```

> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t)
> print(sig_results_boot)

```

```

      IlmnID   case1   case2 control1 control2   case3   case4
815 cg00792849 0.126742 0.1725518 0.1276962 0.114806 0.158568 0.1469312
      control3 control4 diff_results$ordfit_t[diff_list_boot]
815 0.1209293 0.1190965 3.20947
      diff_results$bootstrap_p[diff_list_boot]
815 0

```