# **CGHnormaliter**

April 20, 2011

CGHnormaliter-package

This package implements the CGHnormaliter algorithm which is a strategy for improved normalization of array Comparative Genomic Hybridization (aCGH) data

# **Description**

CGHnormaliter is an iterative algorithm for normalization of aCGH data displaying imbalanced aberrations. First, provisory balanced copy numbers are identified and subsequently used for normalization based on LOWESS. These two steps are then iterated to refine the normalization. The assumption here is that the temporary exclusion of aberrations allows for a more appropriate calculation of the LOWESS regression curve. As a result, after normalization, the log2 intensity ratios of the normals will generally be closer to zero and better reflect the biological reality.

#### **Details**

Package: CGHnormaliter

Type: Package
Version: 1.3.8
Date: 2010-10-11
License: GPL (>= 3)

The package contains two public functions. The function CGHnormaliter performs the iterative normalization of aCGH data, while the function CGHnormaliter.write.table prints normalized aCGH data to a file. See function documentation for details.

# Author(s)

Thomas W. Binsl, Bart P.P. van Houte, Hannes Hettling

#### References

Bart P.P. van Houte, Thomas W. Binsl, Hannes Hettling, Walter Pirovano and Jaap Heringa. CGH-normaliter: an iterative strategy to enhance normalization of array CGH data with imbalanced aberrations. BMC Genomics, 10:401, 2009.

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#### See Also

CGHcall, DNAcopy

CGHnormaliter

Iterative normalization of aCGH data

# **Description**

Implementation of an iterative algorithm for normalization of aCGH data displaying imbalanced aberrations.

#### Usage

```
CGHnormaliter(data, nchrom = 24, stop_threshold = 0.01, max_iterations = 5, pl
```

#### **Arguments**

either a dataframe or character string containing a filename. See section Details data

for the format.

number of chromosomes. nchrom

stop\_threshold

threshold value for the mean difference between the LOWESS regression curves from two consecutive iterations. The iteration is terminated if this difference is

below the stop\_threshold for all samples.

max\_iterations

maximum number of iterations.

logical. If 'plotMA = TRUE', MA-plots before and after normalization are genplotMA

erated of each sample. The plots are saved into a PDF file.

#### **Details**

The input should be either a data. frame or the file name of a tabseparated text file (text files must contain a header). The first four columns should contain the name, chromosome and the start and end position in bp for each array target respectively. The position columns must contain numbers only. Following these are two columns with the raw test and reference intensities for each of your samples. These intensities must be numeric as well. If the input type is a text file, missing values should be represented as 'NA' or an empty field. There is a CGHnormaliter.write.table method that prints the results in a tabular format.

#### Value

This function returns a matrix of objects of class cghCall with dimension (number of clones) \* (number of samples). Each object contains the following components (See section Examples on how to access them):

normalized data

A matrix with the normalized log2 intensity ratios for each profile.

A matrix with the segments for each profile. segments

A data.frame with the calls for each profile. Values are -1 (loss), 0 (normal) calls

or 1 (gain).

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```
probabilities
```

A data.frame with 3 columns of probe information (name, chromosome and position), followed by 3 columns with aberration probabilities for each sample.

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#### References

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#### **Examples**

```
data(Leukemia)
## Normalize the raw intensity values of the first 3 chromosomes.
result <- CGHnormaliter(Leukemia, nchrom=3)
## Get the normalized log2 intensity ratios, segments and calls
normalized.data <- copynumber(result)
segmented.data <- segmented(result)
called.data <- calls(result)
## Plot the normalization result of sample 2
plot(result[,2])
## Write the normalized log2 intensity ratios to file
CGHnormaliter.write.table(result)</pre>
```

CGHnormaliter.write.table

Print normalized aCGH data to a file

# **Description**

This function stores the results of a CGHnormaliter normalization into a plain text file.

# Usage

```
CGHnormaliter.write.table(input, data.type=c("normalized", "segmented", "called"
```

# **Arguments**

input result object of a CGHnormaliter normalization.

data.type type of aCGH data to store. file name to store the data in.

# **Details**

Available aCGH data types are:

- normalized: The normalized log2 intensity ratios
- segmented: The segmented log2 intensity ratios (after normalization)
- called: The calls (after normalization)

The selected data are stored into a tabseparated file with the specified file name.

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#### Author(s)

Thomas W. Binsl, Bart P.P. van Houte, Hannes Hettling

#### References

Bart P.P. van Houte, Thomas W. Binsl, Hannes Hettling, Walter Pirovano and Jaap Heringa. CGH-normaliter: an iterative strategy to enhance normalization of array CGH data with imbalanced aberrations. BMC Genomics, 10:401, 2009.

#### See Also

CGHnormaliter

# **Examples**

```
data(Leukemia)
## Normalize the intensity ratios
## Not run: result <- CGHnormaliter(Leukemia)
## Write the result to a file
## Not run: CGHnormaliter.write.table(result)
## Not run: CGHnormaliter.write.table(result, type="segmented")
## Not run: CGHnormaliter.write.table(result, type="called", file="mycalls.txt")</pre>
```

Leukemia

Array CGH experiment data on childhood acute lymphoblastic leukemia (ALL) in humans

#### **Description**

A dataframe containing 30180 rows and 10 columns, representing the array CGH data of 3 ALL samples.

#### Usage

Leukemia

#### **Format**

A dataframe containing the following 10 columns:

**CloneID** The unique identifiers of array elements.

Chromosome Chromosome number of each array element.

**Start** Chromosomal start position in bp of each array element.

**End** Chromosomal end position in bp of each array element.

Case1.test Background corrected test intensity values for sample 1.

Case1.ref Background corrected reference intensity values for sample 1.

Case2.test Background corrected test intensity values for sample 2.

Case2.ref Background corrected reference intensity values for sample 2.

Case3.test Background corrected test intensity values for sample 3.

Case3.ref Background corrected reference intensity values for sample 3.

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#### **Source**

Provided by the authors (see references).

# References

Paulsson K, Heidenblad M, Morse H, Borg A, Fioretos T, Johansson B: Identication of cryptic aberrations and characterization of translocation breakpoints using array CGH in high hyperdiploid childhood acute lymphoblastic leukemia. Leukemia 2006, 20:2002-2007.

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